

SMASH 2017

Conference Program

September 17th-20th 2017

Baveno, Italy

SMASH 2017 NMR Conference

Dear SMASH 2017 attendees,

We are pleased to extend a warm welcome to the 2017 Small Molecules Are Still Hot NMR Conference. Once again, we find ourselves in the delightful setting of Baveno, on the shores of Lake Maggiore. Nestling in the foothills of the Alps, surrounded by natural wonders, it's hard to believe we are only a short drive from Milan, one of Italy's most important and vibrant cities. For those of you that attended the previous meeting here, we are certain you will agree with the above sentiments, for those who didn't, we are sure you will appreciate this wonderful venue.

Of course, the attractive location for SMASH would be nothing if the scientific program was not strong and relevant to you; we think that we have put together an interesting and varied program that will be exactly that.

The formal program for SMASH begins on Sunday evening, with registration, a mixer and dinner (though we are sure many of you will have attended the various user meetings being held in the daytime of Sunday). This will be the perfect opportunity to catch up with old colleagues and make new acquaintances. The scientific sessions begin on Monday morning, and we have seven varied and exciting oral sessions covering a broad range of topics relevant to small molecules – from structural elucidation to solid state NMR, from reaction monitoring to exotic nuclei, from binding interactions to metabolomics (all tunefully named, as you may have noted). There is also a session which will revisit some of the 'hot' topics from previous SMASH meetings. As for previous meetings, each oral session also includes one short talk which is an 'upgrade' of a poster. We will also have three non-concurrent, interactive workshops, covering calculation of NMR parameters, solid-state NMR for solution-phase specialists and pure shift NMR. The formal program is completed by two poster sessions, which we are sure will provoke plenty of lively discussion – always a highlight at SMASH.

We are also very pleased to be presenting the 4th James Shoolery award to Dr James Keeler, who will give his award presentation on Monday evening before dinner. We look forward to a fascinating journey through the history of his involvement with NMR.

As always, key vendors in the NMR arena will be available at the meeting to showcase their products and discuss their use. For those of you wanting to explore the local area more, there is free time on Tuesday afternoon. Tuesday afternoon will also feature a roundtable discussion concerning ongoing initiatives in improving and standardising NMR data reporting, which although not part of the core program, may be of interest to many of you.

We are delighted that registrations for the meeting are as high as they've ever been, with record numbers of posters submitted and students attending, demonstrating the vibrancy of our field and - dare we say - that small molecules really are still hot. We sincerely hope that this meeting will continue the amazing tradition SMASH has forged since its establishment almost two decades ago.

On behalf of the organising committee, we extend a warm welcome to all of you and thank you for choosing to attend SMASH 2017.

Gary Sharman and Tim Claridge
Co-chairs, SMASH 2017 NMR conference

SMASH 2017 Conference Program

Sunday, September 17th

09:00 - 16:00	Vendor User Meetings (Mestrelab Research, Bruker BioSpin)
17:00 - 18:00	Registration – Grand Hotel Dino
18:00 - 20:00	Mixer – Grand Hotel Dino
20:00 - 22:00	Dinner – Grand Hotel Dino

Monday, September 18th

09:00 - 09:15	Welcome, Announcements and Opening Remarks
09:15 - 10:50	I Still Haven't Found What I'm Looking For - New Approaches to Structure Elucidation Chair: Peter Howe, <i>Syngenta, UK</i> <u>Complex Structural Problems and Unconventional NMR Methods to Solve Them</u> Josep Sauri , <i>Merck Sharp & Dohme (Structure Elucidation Group), Boston, USA</i> <u>New NMR Methods to Improve the Measurement of One-Bond Proton-Carbon Coupling Constants</u> Teo Parella , <i>Universitat Autònoma de Barcelona, Spain</i> <u>Synergistic Combination of CASE Algorithms and DFT Chemical Shift Predictions for Structure Elucidation, Verification and Revision</u> Alexei Beuvich , <i>Merck Sharp & Dohme (Discovery & Preclinical Trials), Kenilworth, USA</i> <u>Computer-Assisted 3D Structure Elucidation of Small Molecules Using Residual Dipolar Couplings and Isotropic ¹³C Chemical Shifts</u> (Upgraded Poster) Roberto Gil , <i>Carnegie Mellon University, USA</i>
10:50 - 11:30	Break
11:30 - 12:30	Workshop 1: Pure and Simple - Understanding Pure Shift NMR Methods Chair: Laura Castañar-Acedo, <i>University of Manchester, UK</i>
12:30 - 14:00	Lunch
14:00 - 15:35	All Shook Up - Metabolites and Multi-Component Samples Chair: Carla Marchioro, <i>Research4Rent (R4R), Italy</i> <u>Mutual Diffusion Driven Experiments and Cheminformatics to Demystify Complex Mixtures</u> Julien Wist , <i>Universidad del Valle, Colombia</i> <u>Complementary Approaches to Mixture Analysis</u> Jean-Marc Nuzillard , <i>Institut de Chimie Moléculaire de Rains, CNRS, France</i> <u>¹H-NMR to Evaluate the Metabolome of Bronchoalveolar Lavage Fluid (BALF) in Bronchiolitis Obliterans Syndrome (BOS)</u> Carlotta Ciaramelli , <i>University of Milano-Bicocca, Italy</i> <u>Robust and Reliable Quantification of Phospholipids in Edible Oils</u> (Upgraded Poster) Joep Van Rijn , <i>DSM Biotechnology Center, Netherlands</i>
15:35 - 16:00	Break
16:00 - 17:00	Workshop 2: Got to Get You into My Life - A Guide to Solid-State NMR for Solution State Spectroscopists Chair: Peter Gierth, <i>Bruker UK</i>
17:00-17:30	Free time
17:30-18:30	Shoolery Award Lecture - 'An NMR Retrospective' James Keeler , <i>University of Cambridge, UK</i>

- 18:30 – 20:00 **Poster Session 1 - Even Better Than the Real Thing**
Even numbered posters to be presented - *Drinks and snacks available*
- 20:00 – 22:00 **Dinner**

Tuesday, September 19th

- 09:00 - 10:35 **Let's Dance - Reaction Monitoring and Kinetics**
Chair: Guy Lloyd-Jones, *University of Edinburgh, UK*
[Catch Me If You Can - Watching Homogeneous Catalysis with Real-time High Resolution FlowNMR](#)
Ulrich Hintermair, *University of Bath, UK*
[Quantitative NMR Spectroscopic Study of Highly Diluted Key Components in Complex Reactive Mixtures: Aqueous Amine Solutions Loaded with CO₂](#)
Erik von Harbou, *University of Kaiserslautern, Germany*
[Chances and Pitfalls of In-Situ Irradiation NMR Spectroscopy](#)
Jonas Kind, *Technische Universität Darmstadt, Germany*
[Ultrafast DOSY NMR of Hyperpolarised Mixtures](#) (Upgraded Poster)
Ludmilla Guduff, *ICSN-CNRS, Gir-sur-Yvette, France*
- 10:35 - 11:00 **Break**
- 11:00 - 12:35 **We Will Rock You - Solid-State NMR Applications**
Chair: Steven Brown, *University of Warwick, UK*
[No Heavy Metal - NMR Crystallography of Metal Salts and Organometallic Compounds](#)
Ann-Christin Poeppler, *Universität Würzburg, Germany*
[Investigation of Powders at Natural Isotopic Abundance using Solid-State NMR and Dynamic Nuclear Polarization](#)
Pierre Thureau, *University of Marseille, France*
[Combined Solid-State NMR, Diffraction and Modeling Studies of Small Molecule Pharmaceuticals](#)
Luis Mafra, *University of Aveiro, Portugal*
[Insights into Amorphous Solid Dispersion of Felodipine Using Solid-state NMR Spectroscopy: Miscibility and Molecular Interactions](#) (Upgraded Poster)
Kanika Sarpal, *University of Kentucky, USA*
- 12:35 - 18:00 **Lunch followed by free afternoon**
- 17:00 - 18:00 **Round Table Discussion on NMR Data Reporting**
(not a formal SMASH program event, but all interested parties are welcome to attend)
- 18:00 – 19:30 **Poster session 2 – Against All Odds**
Odd numbered posters to be presented - *Drinks and light buffet menu of starters available*

Wednesday, September 20th

- 09:00 - 10:35 **Should I Stay or Should I Go? - Non-Covalent Interactions and Complexes**
Chair: Elisabetta Chiarparin, *AstraZeneca, UK*
[Fragment Evolution Without Routine Crystallography](#)
Ben Davis, *Vernalis, UK*
[NMR² for Fast 3D Structure Determination of Protein-Ligand Binding Site Without Protein Resonance Assignment](#)
Julien Orts, *ETH, Switzerland*
[NMR Free Ligand Conformations for Enhanced Structure Based Drug Design](#)
Rodrigo Carbajo, *AstraZeneca, UK*
[Differential EPitope Mapping \(DEEP\) STD NMR to Reveal the Pharmacophore of a Protein Target](#) (Upgraded Poster)
Serena Monaco, *School of Pharmacy, University of East Anglia, UK*

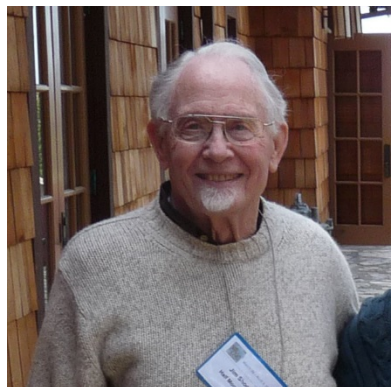
- 10:35 - 11:00 **Break**
- 11:00 - 12:35 **It's Not Unusual - Multinuclear and Inorganic Methods**
 Chair: Michael John, *Georg-August Universität Göttingen, Germany*
[NMR Analysis of Small Paramagnetic Metal Complexes with Large Hyperfine Shifts](#)
 Markus Enders, *University of Heidelberg, Germany*
[Elucidation of the Topological and Chemical Order in Materials by Multi-Nuclear Solid-State NMR](#)
 Pierre Florian, *CEMHTI-CNRS Orleans, France*
[A Hyphenated Computational Protocol for Analysis of Natural Abundance ²H Residual Quadrupolar Couplings in \(Chiral\) Oriented Solvents](#)
 Armando Navarro-Vázquez, *Universidade Federal de Pernambuco, Brazil*
[Hyperpolarised Low-Field NMR for Industrial Reaction Monitoring](#) (Upgraded Poster)
 Olga Semenova, *Center of Hyperpolarisation of Magnetic Resonance, University of York, UK*
- 12:35 - 14:00 **Lunch, Free Time & Vendor Discussions**
- 14:00 - 15:00 **Workshop 3: We can work it out- Calculation of structures and NMR parameters**
 Chair: Giuseppe Bifulco, *Università di Salerno, Italy*
- 15:00 - 15:30 **Break**
- 15:30 – 17:05 **Go Your Own Way - Past SMASH Hot Topics Revisited**
 Chair: Christina Thiele, *Technische Universität Darmstadt, Germany*
[Broadband Pulses Revisited](#)
 Burkhard Luy, *Karlsruhe Institut für Technologie, Germany*
[Small Microcoils Are Still Hot](#)
 Aldrik Velders, *Wageningen University, Netherlands*
[Hot Blooded, Check Out CPMG](#)
 Sarah Robinson, *Genentech, South San Francisco, USA*
[General Approach to Access Long-Range 1H-1H RDCs](#) (Upgraded Poster)
 Davy Sinnaeve, *Ghent University, Ghent, Belgium*
- 17:05 - 17:15 **Closing Remarks**

Thursday, September 21st

- 09:30 – 15:30 **qNMR Minisymposium (Not a formal SMASH event)**
 Chairs: Michael Maiwald, *BAM* and Michael Bernstein, *Mestrelab*
 This is not a formal SMASH program event, but all interested parties are welcome (no registration fee but please register).
[Symposium program](#)

The James Shoolery Award

In 2014, SMASH established the James Shoolery Award as a grant, in honor of James N. Shoolery, to recognize the important contributions by an individual to the field of small molecule NMR spectroscopy.



In 1952, Jim Shoolery joined Varian Associates to set up an applications laboratory for NMR spectroscopy. His main initial goals were to develop applications of NMR in chemistry and to educate the wider chemistry community in the potential value of NMR spectroscopy in their research. In pursuit of these goals during the 1950's, he published a series of highly popular ads entitled "NMR at Work," initially in *Analytical Chemistry* and later on the back page of the *Journal of the American Chemical Society*. These illustrated a wide range of applications of NMR in chemistry and were based on work that he carried out in the applications lab. He also wrote a number of "Technical Information Bulletins" to help spectrometer owners in the operation of their instruments. Finally, he gave numerous lectures at conferences and research laboratories and at the annual NMR and EPR workshops that Varian Associates held in Palo Alto starting in 1958. In a 1993 article on the early history of NMR, he estimated that about 20,000 scientists had attended these different lectures by the end of the 1950's.

At the same time, Jim interacted with the R & D division of Varian on NMR instrument improvement, including the progression of ^1H operating frequency on Varian spectrometers from 30 to 40 to 60 and finally to 100 MHz by 1959. He was also involved in important technical improvements, including sample spinning, shim coils, spin decoupling, a flux stabilizer, and an electronic integrator. However, even with these improvements, the HR series of spectrometers were still extremely tricky to operate, requiring a significant amount of training, operating experience and patience. Jim realized that NMR spectroscopy would not reach its full potential as an analytical technique in chemistry until a spectrometer was developed that would be much easier to use, similar to the routine IR spectrometers that were already available from other manufacturers. Therefore, in 1957, Jim teamed with Emery Rogers of the marketing division of Varian to propose to the R & D division the development of a lower cost NMR spectrometer, which could use calibrated chart paper, which was rugged and reliable, and which could be run by graduate students and laboratory technicians with no training other than that provided by the spectrometer manual. He was heavily involved in this project, which resulted in 1961 in the introduction of the Varian A-60. This was a truly revolutionary development whose ease of operation triggered a dramatic increase in the use of NMR spectroscopy by chemists, in general, and by organic chemists, in particular. To illustrate its impact, the 1960 volume of the *Journal of Organic Chemistry* contained only one paper reporting the use of NMR while the 1967 volume included 220 papers, which used NMR data. In 2011, the seminal role of the A-60 in the development of NMR as a valuable analytical technique was recognized by the American Chemical Society as a National Historical Chemical Landmark in a ceremony at the Agilent facility in Santa Clara.

After the initial demonstration of FT NMR at Varian, Jim was involved in the development of the CFT-20 and FT-80 Varian spectrometers. These followed in the footsteps of the A-60 in being low cost and easy-to-use instruments for chemistry labs. In 1972, his book, "A Basic Guide to NMR," was published by Varian Associates and helped to educate many young chemists in the use of NMR. Later, with the

development of multi-pulse sequences and 2D NMR, Jim was among the first to recognize the great value of these techniques for identifying unknown organic chemical structures, particularly in the natural products field. Jim, along with Steve Patt, developed the APT sequence for spectral editing ^{13}C spectra of organic compounds and, through the 1980's, he collaborated with a number of natural products groups in establishing structures and assigning spectra of the compounds which they had isolated. He also, in 1984, published an important review article in the *Journal of Natural Products*, which clearly demonstrated the value of modern NMR techniques in the natural products field.

SMASH 2017 Scholarship Recipients



The following students received a scholarship to attend SMASH 2017

- **Christian Adam**, Karlsruhe Institute of Technology, Germany
- **Jasper Adamson**, National Institute of Chemical Physics and Biophysics, Estonia
- **Martina Bugáňová**, Czech Academy of Sciences, Czech Republic
- **Guilherme Dal Poggetto**, University of Manchester, United Kingdom
- **Lydia Dewis**, University of Bristol, United Kingdom
- **Simone Di Micco**, University of Salerno, Italy
- **Claire Dickson**, University of Bristol, United Kingdom
- **Trent Graham**, Washington State University, United States of America
- **Ludmilla Guduff**, ICSN-CNRS, France
- **Albert Hofstetter**, École Polytechnique Federale de Lausanne, Switzerland
- **Julian Ilgen**, Technische Universität Darmstadt, Germany
- **Tangi Jezequel**, University of Nantes, France
- **Simon Kern**, Bundesanstalt für Materialforschung und – prüfung, Germany
- **Jonas Kind** Technische Universität Darmstadt, Germany
- **Raul Laasner**, Duke University, United States of America
- **Inês Martins**, Universidade de Lisboa, Portugal
- **Serena Monaco**, University of East Anglia, United Kingdom
- **Pinelopi Moutzori**, University of Manchester, United Kingdom
- **Keyla Ortiz**, Universidad Industrial de Santander, Colombia
- **Francesco Puig-Castellví**, IDAEA-CSIC, Spain
- **Kanika Sarpal**, University of Kentucky, United States of America
- **Mira Schwab**, Technische Universität Darmstadt, Germany
- **Olga Semenova**, University of York, United Kingdom
- **Alexandra Shchukina**, University of Warsaw, Poland
- **Davy Sinnaeve**, University of Ghent, Belgium
- **Pablo Trigo Mourino**, Max-Planck-Institute for Biophysical Chemistry, Germany
- **Siyang Zhong**, University of Bristol, United Kingdom

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Technische Universität Darmstadt

Monday, September 18th
09:15 - 10:50

**I Still Haven't Found What I'm
Looking for – New Approaches to
Structure Elucidation**

Chair: Peter Howe

Speakers:

Josep Sauri
Merck – Boston (US)

Teo Parella
UAB (ES)

Alexei Buevich
Merck – Kenilworth (US)

Roberto Gil (Upgraded Poster)
Carnegie Mellon University (US)

Complex Structural Problems and Unconventional NMR Methods to Solve Them

Josep Saurí¹, Yizhou Liu², Kirk R. Gustafson³, Emily Mevers⁴, Jon Clardy⁴, Gary E. Martin²,
and R. Thomas Williamson²

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Merck Sharp & Dohme, Boston, MA 02115, USA
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4. Department of Biological Chemistry and Molecular Pharmacology,
Harvard Medical School, Boston, MA 02115, USA

Routine small and medium-sized molecules can generally be characterized by conventional NMR techniques such as 1D and 2D NMR experiments including COSY, NOESY/ROESY, HSQC and HMBC. However, when dealing with unknown and/or more complex molecular structures – especially proton-deficient compounds, an assignment strategy beyond the "normal" suite of NMR experiments is often required. In this presentation, we will discuss several examples of complex structural problems that were successfully addressed by the acquisition of novel NMR methods.

New NMR Methods to Improve the Measurement of One-Bond Proton-Carbon Coupling Constants

Teodor Parella

Servei de Ressonància Magnètica Nuclear, Universitat Autònoma de Barcelona, E-08193 Bellaterra (barcelona), Catalonia, ES

A set of NMR methods based on the regular HSQC pulse scheme for the accurate determination of one-bond proton-carbon coupling constants are presented. A discussion about the pros and cons to measure them from the direct F2 or the indirect F1 dimensions is made. First, some F2-based methods (PIP-HSQC [1] and perfectHSQC [2] experiments) yielding pure in-phase multiplets are discussed. Then, tips and tricks to implement J-F1-resolved HSQC experiments are introduced, and some practical matters concerning the measurement on diastereotopic CH₂ systems, solutions to avoid signal overlap and details about automation/accuracy are evaluated [3-7]. Some concepts including perfect NMR, J/ δ -scaling or the generation of user-friendly 1JCH NMR profiles are presented. Finally, examples to determine these couplings in isotropic and weakly aligned anisotropic media are shown. In particular, the simultaneous and direct in-situ determination of scalar, total and dipolar coupling constants and the impact of using 1DCH and 2DHH in the efficient structural discrimination of small molecules and natural products are illustrated.

1. L. Castañar, J. Saurí, R.T. Williamson, A. Virgili and T. Parella, *Angew. Chem. Intl. Ed.* 53, 8379-8382 (2014).
2. L. Castañar, E. Sistaré, A. Virgili, R.T. Williamson and T. Parella, *Magn. Reson. Chem.* 53, 115-119 (2015).
3. J. Saurí, L. Castañar, P. Nolis, A. Virgili and T. Parella, *J. Magn. Reson.* 242, 33-40 (2014).
4. L. Castañar, M. García, E. Helleman, P. Nolis, R.R. Gil and T. Parella, *J. Org. Chem.*, 81, 11126-11131 (2016).
5. N. Marcó, R.R. Gil and T. Parella, *Magn. Reson. Chem.*, in press (2017); DOI: 10.1002/mrc.4575.
6. N. Marcó, A.A. Souza, P. Nolis, R.R. Gil and T. Parella, *J. Magn. Reson.*, 276, 37-42 (2017).
7. N. Marcó, A.A. Souza, P. Nolis, C. Cobas, R.R. Gil and T. Parella, *J. Org. Chem.*, 82, 2040-2044 (2017).

Synergistic Combination of CASE Algorithms and DFT Chemical Shift Predictions for Structure Elucidation, Verification and Revision

Alexei V. Buevich¹ and Mikhail E. Elyashberg²

1. Department of Discovery and Preclinical Sciences, Process Research and Development, NMR Structure Elucidation, Merck & Co., Inc., Kenilworth, NJ, USA
2. Moscow Department, Advanced Chemistry Development (ACD/Labs), Moscow, RU

Structure elucidation of complex natural products and organic compounds remains a challenging problem. Even when equipped with advanced spectroscopic methods, structure elucidation methodology is still not free from errors and, hence, the development of better, more robust methods remains in high demand. To support this endeavor, CASE (Computer-Assisted Structure Elucidation) expert systems were developed [1]. These systems are capable of generating an ensemble of all possible structures consistent with the molecular formula and the set of 2D NMR data followed by selection of the most probable structure on the basis of empirical NMR chemical shift prediction.

However, in some cases, CASE systems failed to distinguish the correct structure when average deviations of chemical shifts were too large or when the correct structure was among several top-ranked structures with acceptable but very similar deviations. Herein, we demonstrate for the first time that the combination of CASE and density functional theory (DFT) methods for NMR chemical shift prediction [2] allows unequivocal determination of correct structure even in such difficult situations [3]. This approach has been tested on three natural products: aquatolide, coniothyron and epoxyrousoenone. All three cases represent rather challenging structural problems: the structures of the first two molecules were originally misassigned and the third structure, in addition to being a proton-deficient molecule, has four chiral centers.

We demonstrated that the proposed synergistic combination of CASE and DFT methodologies is an unbiased, reliable, and very efficient structure verification and de novo structure elucidation method that can be potentially applied to difficult structural problems when molecular systems are chiral, conformationally flexible and/or when other experimental methods (X-ray analysis, INADEQUATE, etc.) would be difficult or impossible to use.

1. Elyashberg M. E., Williams A. J. *Computer-based Structure Elucidation from Spectral Data. The Art of Solving Problems*; Springer, Heidelberg, 2015.
2. Lodewyk M. W., Siebert M. R., Tantillo D. J. *Chem. Rev.*, 112, 1839-1862, 2012.
3. Buevich A. V., Elyashberg M. E. *J. Nat. Prod.*, 79 (12), 3105–3116, 2016.

Computer-Assisted 3D Structure Elucidation of Small Molecules Using Residual Dipolar Couplings and Isotropic ^{13}C Chemical Shifts

Roberto R. Gil¹, Kirill Blinov², and Armando Navarro-Vazquez³

1. Department of Chemistry, Carnegie Mellon University, Pittsburgh, PA, US
2. MestReLab Research S. L., Santiago de Compostela, ES
3. Departamento de Química Fundamental, Universidade Federal de Pernambuco, Recife, BR

The 2D structure (molecular constitution) of most small molecules can be in principle straightforwardly determined by manual or automatic analysis of a set of experimental data that includes the molecular formula, a series of 1D and 2D NMR experiments providing through-bond connectivity (COSY, TOCSY, HSQC, HMBC and ADEQUATE/INADEQUATE), and empirical chemical shift predictions. This is the main concept embedded in CASE (Computer Assisted Structure Elucidation) programs. Once the 2D structure is available, the determination of the relative spatial arrangement (configuration and preferred conformation) of all atoms in the molecule is a more challenging task commonly addressed by using NOE and 3J coupling constants analysis, as well as recent developments on DFT calculations of ^{13}C chemical shifts (DP4). The development of the application of Residual Dipolar Couplings (RDCs) to the configurational and conformational analysis of small molecules has matured enough in the recent years to perform this task in an almost straightforward way, without even the need of using NOE and 3J coupling analysis, for the analysis of rigid and semi-rigid small molecules. We have recently shown that it is possible to go from molecular constitution to configuration without human intervention by feeding the program with an SDF structure file from a CASE-based program and a table containing only one-bond proton-carbon RDCs.[1] The process involves a) generation of the configurational space, b) conformational analysis for each configuration, c) automatic superposition of conformers for single-tensor analysis, d) fitting of RDC data to a set of conformation/configurations using model selection to prevent overfitting, e) plot of a bar diagram with quality factors indicating the selection of the correct 3D structure. This work led to the creation of StereoFitter, a complete computational module that can perform multiple-nmr parameters fitting (scalar couplings, NOE-derived distances, chemical shifts, RDCs, RCSAs) to combinations of configuration/conformations using model selection (Akaike Information Criterion (AIC)) to prevent overfitting. StereoFitter can be trivially extended to the use of chiroptics parameters (ECD, VCD) to determine absolute configuration. Generation of configurations, conformational analysis, DFT geometry optimizations, and analysis of DFT GIAO calculations of NMR parameters (J couplings, Chemical Shift Tensors) is also performed by StereoFitter by interacting with third party computational chemistry packages. In the present work we would like to show that, after the determination of the molecular constitution by the CASE program, a combined use of RDCs and isotropic ^{13}C chemical shifts is enough to determine configuration, particularly useful for proton-deficient molecules. For the antimalarial drug Artemisinin,[2] the correct structure was determined using only ^{13}C chemical shifts. The whole process from 2D structure to the generation of 38 diastereomers, followed by DFT energy minimization and DFT GIAO calculation of CSA tensor for each diastereomer, and finalizing with the fitting of chemical shifts to the set of diastereomers, took only 2 hours in a 5 years old PC equipped with 2 Xeon Processor with 6 cores each. Results shown in Figure 1.

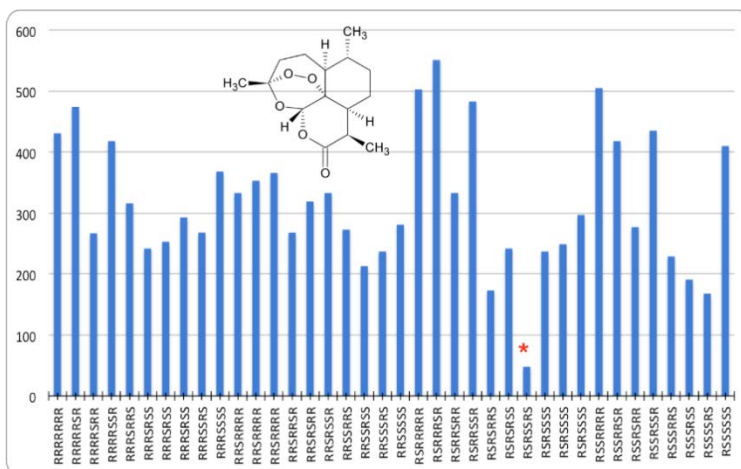


Figure 1: Determination of the configuration of Artemisinin using only isotropic ^{13}C chemical shift. Fitting DFT calculated chemical shift with the experimental data using StereoFitter. The red asterisk shows the correct structure with the lowest χ^2 value.

We have also applied the method to other relevant natural products such as the sesquiterpene lactone 10-epi-8-deoxycumambrin B [3] and the fungal metabolite Homodimericin A.[4] The structure of this natural product was determined using a combination of CASE, RDCs and RCSAs, since it has an hexacyclic core with fourteen quaternary carbons, eleven of them contiguous. For this particular compound we used the data reported by the authors.[4] For Homodimericin A we observed that it is possible to use ^{13}C chemical shifts as an alternative to RCSAs to determine its configuration.

In contrast to methodologies such as DP4 our method allows the fitting of populations to all the available experimental data including chemical shifts. Potential overfitting problems are treated by using the Akaike Information Criterion (AIC) where models with increasing number of populations are penalized.

CASE analysis was performed using the MestReNova Structure Elucidator. Most of the results presented here used Poly(methylmethacrylate) (PMMA) based flexible gels, whose degree of alignment can be easily tuned by variable and reversible compression.[5]

Acknowledgments: ANV thanks the UFPE for a visiting professor contract and FACEPE (APQ-0507-1.06/15) for financial support. NMR instrumentation at CMU was partially supported by the NSF(CHE-0130903 and CHE-1039870). R.R.G. acknowledges support from the NSF (CHE-1111684). We thanks Leandro F. Gil-Silva from Alignment Technologies LLC / Mestrelab Research SL for the provision of aligning gels.

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2. Coordinating Group for Research on the Structure of Qing Hau Sau. *Kexue Tongbao* (Chinese Edition) 22(3), 142, **1977**.
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Monday, September 18th
11:30 - 12:30

Workshop 1

**Pure and Simple – Understanding
Pure Shift NMR Methodology**

Coordinated by:
Laura Castañar-Acedo, The University of Manchester

Understanding Pure Shift NMR Methodology

Coordinated by:

Laura Castañar-Acedo, The University of Manchester

Currently, pure shift NMR is an area of high interest. The aim of this workshop is to cover the practical aspects of these experiments. First, we will briefly describe the different methods available, their implementation in conventional 1D and multidimensional NMR experiments, and we will show several practical applications reported in recent years. The main part of the workshop will then deal with the practical features of pure shift experiments, such as optimal acquisition parameters and post-processing. No NMR experiment is perfect, and pure shift experiments are no exception. Some of the problems/limitations, such as sensitivity and spectral quality, will therefore also be discussed, as well as the techniques available for their removal/reduction. Finally, we will have an open question and answer session about all of the aspects covered, and about the challenges and possible next steps in the amazing adventure of the development and application of pure shift NMR experiments.

Monday, September 18th
14:00 – 15:35

All Shook Up – Metabolites and Multi-Component Samples

Chair: Carla Marchioro

Speakers:

Julien Wist
Universidad del Valle, Cali (CO)

Jean-Marc Nuzillard
Université de Reims Champagne-Ardenne, Reims (FR)

Carlotta Ciaramelle
University of Milano-Bicocca, Milan (IT)

Joep van Rijn (Upgraded Poster)
DSM Biotechnology Center, Delft (NL)

Mutual Diffusion Driven Experiments and Cheminformatics to Demystify Complex Mixtures

Christian Pantoja¹, Michael Zasso², Daniel Kostro², Andrés Castillo¹, Andrés Bernal¹, Alejandro Bolaños¹, Luc Patiny², and **Julien Wist**¹

1. Chemistry Department, Universidad del Valle, Cali, CO
2. Institute of Chemical Sciences and Engineering, Ecole Polytechnique Fédérale de Lausanne (EPFL), CH-1015 Lausanne, CH

NMR earned its place in every chemistry department because it enabled the identification and characterization of compounds, better and quicker than any other technique. As NMR spectrometers got more mature, more complex samples such as macromolecules or mixtures of many compounds started being studied. We thus moved from the task of assigning each signal in a spectrum to atoms of a molecule to the task of assigning each signal in a spectrum to atoms in hundreds of molecules. Being successful at such a complex process requires combining efforts to improve both the resolution and overall quality of the experimental data and the accuracy of posterior analysis. Here we would like to present two contributions aiming at both directions; the first one being a new approach to experimentally discriminate different molecules in a mixture based on mutual diffusion coefficients, the second one being a collection of computational building blocks that can be used to elaborate specific tools to automate complex analysis pipelines.

MDD experiments. Deconvolution of mixtures may be achieved *in-situ* with the help of a physical driving force. Think of DOSY, where the self-diffusion coefficient is used to discriminate signals from different molecules depending on their size. Recently we have shown that it is possible to establish a gradient of concentration for a binary solution of triethylamine-deuterium oxide inside an NMR tube. The gradient is induced by transition from a single phase to a biphasic state and will fade away as the system is allowed to return to its thermodynamical equilibrium (single phase). Dissolving species in such a system will have them spread along the tube according to their mutual or cooperative diffusion coefficients. Spatial encoding NMR experiments allow to capture the gradient for each individual species, thereby providing a mean to discriminate them. Results obtained in triethylamine-D₂O were similar to those obtained by PFG-NMR, but not identical: water and methanol have similar self-diffusion coefficients and are poorly resolved in the DOSY experiment, while they have different mutual diffusion rates and appear well separated in the Mutual Diffusion Driven (MDD)-NMR experiment.

Although still limited to a few binary systems (triethylamine-water, nitrobenzene-hexane, methanol-cyclohexane and others), such an approach offers the possibility to pick a system according to the affinity of the molecules that should be separated. Finally, initial conditions, such as molar fraction, mixing time and temperature can be tuned to improve separation.

Cheminformatics. Despite the acquisition of NMR data is highly automated, posterior analysis is still mainly performed by hand and very few computational tools exist that perform them automatically. Assisted assignment has been around for a while but requires a lot of manual input from users. On the other hand, fully automatic assignment procedures do not rely on human intervention, but do rely on large amounts of manually-curated data. Similarly, only few tools exist that assist researchers in the task of matching signals with the different components of a mixture, or better, that perform it automatically. Here we present two examples of web applications build over our open source framework for cheminformatics.

The first tool tries to assign $^1\text{H-NMR}$ spectra in an iterative manner. Once atoms have been confidently assigned to signals, this knowledge is used to learn and predict expected chemical shifts. A new iteration can start and more atom-signal pairs are assigned. As a result, the accuracy of the predictions was found to increase after each iteration, as is the number of atoms assigned. Training this system with less 2341 pairs of molecule-spectrum let to predict over 60% of test pairs within 0.2 ppm, competing with the popular Spinus predictor. To the best of our knowledge, this is the first self-learning algorithm of its kind and it may in the future, once fed with enough data, compete with the best available predictors.

As a second example, we present a tool to assist researchers in the task of identifying metabolites in mixtures. A database of reference compounds is used to propose putative candidates. First, a fully automated pipeline allows to schedule the acquisition of reference spectra and store them in the database, where they are duly peak-picked. A manual revision of the result is made easy by a complete user graphical interface. On the other end of the pipeline, metabolic profiles can be uploaded and analyzed using o-PLS and STOCSY approaches. PLS loadings and STOCSY rows trigger queries in the database resulting in a list of possible candidates.

These tools are free to use and open source projects, written in JavaScript and thus natively designed for the web, the best place ever to share information.

Complementary Approaches to Mixture Analysis

Ali Bakiri¹, Ilhem Zebiri¹, Jane Hubert¹, Romain Reynaud², Laurence Voutquenne¹, Jean-Hugues Renault¹, and **Jean-Marc Nuzillard¹**

1. Institut de Chimie Moléculaire, Université de Reims, FR
2. Soliance-Givaudan, Pomacle, Marne, FR

The topic of mixture analysis is considered here for the identification of known compounds and for the discovery of new ones. The complementary methods we use or develop may not necessarily involve a physical separation step.

The most straightforward way to get around the difficulty of analyzing a mixture is to fractionate it into pure compounds and to analyze them separately. The LC-NMR technique was created for this purpose. The availability of always higher static magnetic fields and of cyprobes partly solved the NMR sensitivity problem that arises from the low amounts of sample injected in high resolution analytical LC columns. The concentration of LC effluents by their trapping in a solid phase extraction (SPE) cartridge led to the LC-SPE-NMR protocol. We used it for the re-investigation of the chemical content of a Peruvian plant, *Dendrobangia boliviana*, and it allowed us to characterize five new saponins [1]. Another way to solve the sensitivity problem is to increase the column effluent concentration. This approach was explored about 20 years ago by the coupling of Centrifugal Partition Chromatography (CPC) with NMR [2].

The combination of CPC fractionation (without any obligation to obtain a pure compound in any fraction), ¹³C NMR analysis of fractions, resonance binning and hierarchical clustering of evolution profiles of ¹³C resonance intensities leads to the constitution of lists of chemical shifts values for the individualized components of a mixture. These lists are then used as search targets for a locally developed spectrochemical database of natural products. The whole process, called CAMEL (CARActérisation de MÉLanges, mixture characterization in French), was already applied successfully, mostly to plant extracts selected for dermocosmetic activities [3]. The CAMEL strategy identifies compounds already reported in literature and alerts its user for a possible discovery of new compounds. More recently, an algorithm was designed for the direct identification of known compounds from the ¹³C NMR spectrum of a complex mixture, such as a plant extract, but without any purification step [4].

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2. Spraul, M, Braumann, U, Renault, J-H, Thépenier, P, Nuzillard, J-M, *J. Chromatogr. A*, 776(1-2), 255-260, 1997.
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¹H-NMR to Evaluate the Metabolome of Bronchoalveolar Lavage Fluid (BALf) in Bronchiolitis Obliterans Syndrome (BOS)

Carlotta Ciaramelli¹, Marco Fumagalli², Simona Viglio², Anna Maria Bardoni², Davide Piloni^{2,3}, Federica Meloni^{2,3}, Paolo Iadarola², and Cristina Airoidi¹

1. University of Milano-Bicocca, Milan, IT
2. University of Pavia, Pavia, IT
3. IRCCS Foundation Policlinico San Matteo, Pavia, IT

Bronchiolitis obliterans syndrome (BOS) is the main phenotype of an irreversible obstructive graft dysfunction known as chronic lung allograft dysfunction (CLAD), which challenges patient survival after lung transplantation. CLAD diagnosis relies on functional parameters and presents a significant heterogeneity in the pathology evolution, varying from a gradual decline in graft function to a more abrupt onset and more severe decline. Given this heterogeneity, tools that will help unravel the complexity of the disease and identify useful predictive markers are urgently needed. Metabolomics is indeed a platform capable of capturing disease-relevant metabolic profile changes and molecular signatures of disease processes.

To this purpose, NMR spectroscopy was employed for the metabolic profiling of bronchoalveolar lavage fluid (BALf) from lung transplant recipients without BOS (stable subjects, n=10), and with BOS at different degree of severity: potential BOS (BOS Op, n=10) and established BOS (BOS I, n=10). [1] The tuning of a number of parameters concerning both sample preparation/processing and variations of spectra acquisition modes was performed to design an efficient and reproducible protocol for the screening of metabolites in a pulmonary fluid that should reflect the status of airway inflammation/injury.

Exploiting the combination of mono and bi-dimensional NMR experiments, 38 polar metabolites, including amino acids, Krebs cycle intermediates, mono- and di-saccharides, nucleotides and phospholipid precursors were unequivocally identified, in substantial agreement with previously reported assignments. [2,3] In order to correlate the metabolic signature with the onset of BOS, metabolites' content of the above recipients was analyzed by multivariate (PCA and OPLS-DA) statistical methods. PCA analysis differentiated stable subjects from BOS I patients and this discrimination was significantly improved by the application of OPLS-DA. The analysis of stable vs BOS Op and of BOS Op vs BOS I samples showed a clear discrimination of considered cohorts, although with a poorer efficiency compared to those measured for stable vs BOS I patients, these results being in agreement with the development of the disease.

The NMR data pointed out the potential of this methodology for the identification of predictive biomarkers to unravel and monitor serious post-transplant lung conditions, including BOS. These preliminary results strongly support the possibility to afford a metabolic signature of BOS from NMR BALf analysis.

Acknowledgements

This work was financially supported by Fondazione CARIPLO (Milan, Italy) grant # 2013-0820.

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3. Wolak, J. E.; Esther, C. R. Jr.; O'Connell, T. M. *Biomarkers* 14(1), 55-60, **2009**.

Robust and Reliable Quantification of Phospholipids in Edible Oils

Joep van Rijn, Paul Groen, Remco Muntendam, and Adriana Carvalho de Souza

DSM Biotechnology Center, Delft, NL

Crude plant oils will form a solid sediment called gum upon storage. These gums contain mainly phospholipids. To obtain commercial edible oil the gum has to be removed, this process is known as degumming. The oldest form degumming is obtained by mixing crude oils with water and the solid formed is removed by centrifugation. Enzymatic degumming can be applied to reduce oil losses during degumming. To monitor degumming processes a robust and accurate phospholipid quantification is needed. Several techniques have been applied for the phospholipids quantification in edible oils, such as ICP, LC and TLC. However, in the past decade ^{31}P NMR has shown to have advantages above the previously mentioned techniques because of the possibility to quantify different phospholipids directly from oil in an accurate and fast manner. In this report, we show the application and performance of 1D ^{31}P NMR method for quantification of several phospholipids in crude and refined oils. The method is robust and has a variation lower than 5% for most phospholipids.

Monday, September 18th
16:00 – 17:00

Workshop 2

**Got to Get You into My Life – A Guide
to Solid-State NMR for Solution State
Spectroscopists**

Coordinated by:
Peter Gierth, Bruker

A Guide to Solid-State NMR for Solution State Spectroscopists

Coordinated by:

Peter Gierth, Bruker

Solid state NMR is a tool with a large range of applications to small molecules, including providing spectra of insoluble compounds, polymorphism studies, analysis of solid mixtures, investigation of protonation states. Historically solid-state NMR has been seen as an exotic technique, requiring large amounts of specialised hardware and considerable operator experience, limiting its wider use. In recent years the improvements of general NMR hardware have allowed sophisticated solid-state NMR experiments on standard instruments with simple addition of a probe and pneumatic spinning unit, and this combined with an increasing array of relevant experimental techniques has increased interest in solid-state NMR among the general NMR community. This workshop will cover hardware requirements, some basic experimental procedures, and examples of applications of classical and recently developed techniques to small molecule problems.

Monday, September 18th
18:30 – 20:00

Presentation of the James N. Shoolery Award

SMASH 2017 Recipient

James Keeler

University of Cambridge, UK



James Keeler was born and raised in rural Norfolk, the second son of a farming family. He attended the local grammar school where an inspiring teacher first sparked an interest in chemistry and encouraged James to pursue this at University. He was an undergraduate in Oxford (at John's College) and then won a scholarship from Merton College to continue with a doctorate under the supervision of Ray Freeman. On completing his doctorate in 1984, James was appointed to a 'new blood' lectureship in the Department of Chemistry, University of Cambridge and a Fellowship at Selwyn College. He has continued in these roles since that date.

James' research interests have covered a wide range of topics in the broad area of 'new techniques' in high-resolution NMR. Particular themes have included improving lineshapes, the measurement of coupling constants, the suppression of zero-quantum coherence, the application of pulsed field gradients (especially to NOE experiments), and pure shift techniques. In recent years he has shifted his focus more to teaching and is the author of three undergraduate texts as well as the introductory NMR text *Understanding NMR Spectroscopy*.

James has been awarded the Royal Society of Chemistry Meldola Medal, the University of Cambridge Pilkington Teaching Prize, and the Royal Society of Chemistry Silver Medal for contributions in Magnetic Resonance.

Tuesday, September 19th

09:00 - 10:35

Let's Dance – Reaction Monitoring and Kinetics

Chair: Guy Lloyd Jones

Speakers:

Ulrich Hintermair

University of Bath, Bath (UK)

Erik von Harbou

University of Kaiserslautern, Kaiserslautern (DE)

Jonas Kind

TU Darmstadt, Darmstadt (DE)

Ludmilla Guduff (Upgraded Poster)

ICSN-CNRS, Gir-sur-Yvette (FR)

Catch Me If You Can - Watching Homogeneous Catalysis with Real-Time High Resolution FlowNMR

Andrew M. R. Hall¹, Peilong Dong², Anna Codina³, John P. Lowe², and Ulrich Hintermair¹

1. Centre for Sustainable Chemical Technologies, University of Bath, UK
2. Department of Chemistry, University of Bath, UK
3. Bruker UK Ltd., Coventry, UK

Molecular solution-phase catalysis is a key technology for addressing sustainability issues in the chemical industry. The development and optimization of homogeneous catalysts is, however, often hampered by limited insight into the kinetics of the reaction and the transformation of the catalyst, enforcing empirical optimization. Rational catalyst and reaction development is only possible through thorough understanding of catalyst activation and de-activation mechanisms, potential resting or dormant states, and the kinetics of the productive cycle (i.e. rate-limiting steps). While rather laborious techniques are available to investigate the afore-mentioned aspects separately, there is no readily applicable technique that may be used universally in early stages of catalyst development. We will present the development of such a technique based on NMR spectroscopy as one of the most informative solution-phase analysis method available.

We have built a reaction setup in which a reaction vessel is coupled to a capillary NMR probe via small diameter HPLC tubing. With this we can continuously circulate a reaction mixture through the spectrometer, thereby follow the reaction progresses and catalyst transformation under catalytically relevant conditions in real time.

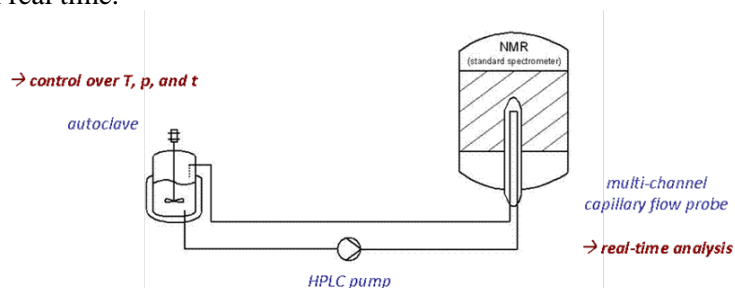


Figure 1. Schematic of the FlowNMR setup.

We have characterised the hydrodynamic flow characteristics of the setup and measured flow effects on continuous ¹H NMR acquisition to quantify changes in T₁, T₂ and signal intensity as function of volumetric flow velocity. Application in real-time reaction and catalyst monitoring under strictly inert conditions has been demonstrated, and multiple solvent suppression and selective excitation techniques allow the detection of minor intermediates even in non-deuterated solvents. [1]

Using the described setup, we have followed the asymmetric transfer-hydrogenation of acetophenone using Noyori's chiral TsDPEN-ligated (arene)Ru^{II} complexes as catalysts in basic iso-propanol (figure 2).[2]

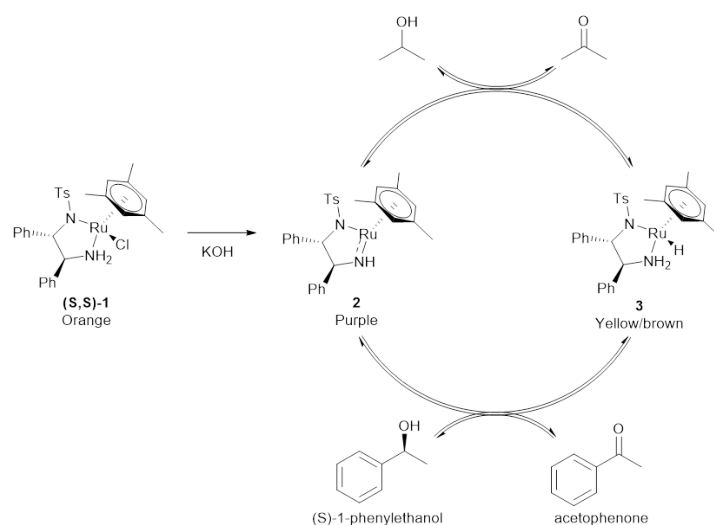


Figure 2. Catalytic system investigated.

As continuous NMR acquisition can be started on pure solvent flow, and the reaction initiated by sequential injection of reagents, the entire reaction can be followed without any lag phases. High quality kinetic data is obtained for this air-sensitive transition metal catalysed reaction (figure 3). Importantly, both conversion and enantioselectivity at the end of the reaction are identical to the results obtained in a sealed Schlenk flask, demonstrating that several hundred pumping cycles do not affect the chemistry in solution.

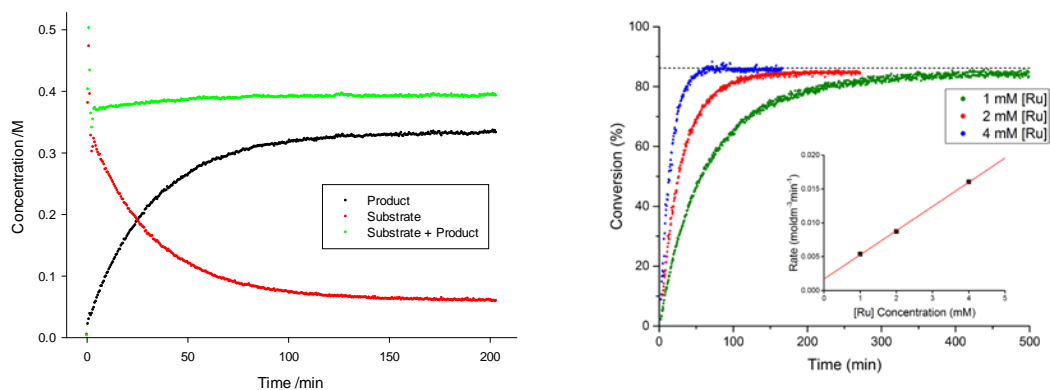


Figure 3. Reaction kinetics of the system shown in fig. 2 as obtained by ^1H FlowNMR.

Furthermore, using selective excitation techniques, we were able to observe metal-hydride intermediates during the reaction under the same conditions. These can be quantified over time and directly correlated to product formation kinetics (figure 4).

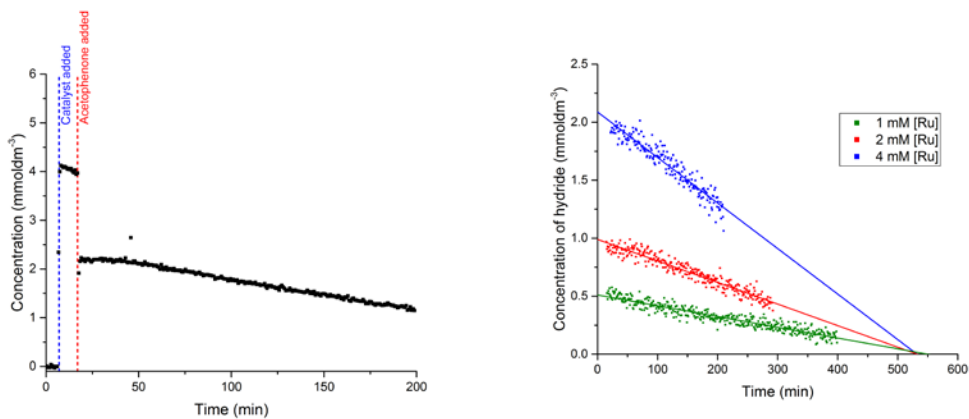


Figure 4. Ru-H intermediates detected during the reaction shown in fig. 2.

We will present a full account of our data and discuss their implications for the mechanism of this widely used reaction.

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2. K-J. Haack, S. Hashiguchi, A. Fujii, T. Ikariya, R. Noyori, *Angew. Chem. Int. Ed.* **1997**, *36*, 285.

Quantitative NMR Spectroscopic Study of Highly Diluted Key Components in Complex Reactive Mixtures: Aqueous Amine Solutions Loaded with CO₂

Erik von Harbou, Richard Behrens, Elmar Kesler, and Hans Hasse

Laboratory of Engineering Thermodynamics, University of Kaiserslautern, Kaiserslautern, DE

NMR spectroscopy is widely applied in chemistry for example to elucidate chemical structures. Its potential for quantitative investigation of complex reactive multi-phasic systems, however, has been hardly exploited in chemical reaction engineering or thermodynamics so far even though it offers many advantages for quantitative investigations of complex systems. On the one hand, NMR spectroscopy features non-invasive measurements so that the investigation of reactive systems, which are typically very sensitive to changes of temperature or pressure, are not disturbed by sampling. On the other hand, in contrast to optical spectroscopy methods, the composition of the sample can be determined in most cases directly from the acquired NMR signal without the need for calibration prior to the analysis. Furthermore, the concentrations of several different analytes can be determined simultaneously even though their values differ by orders of magnitude. Thus, NMR spectroscopy is often not only the best but also the only applicable method to quantify complex reactive systems with intermediates or products that cannot be isolated from the mixture.

In this work, we will present an *in-situ* NMR spectroscopic method to investigate the liquid phase reactions in aqueous amine solutions loaded with CO₂ under high pressure. These systems are widely applied in reactive absorption processes for example for scrubbing CO₂ from synthesis or flue gas. In these processes, the CO₂ is absorbed from the gas phase into the liquid phase where it undergoes immediately several consecutive reactions with the solvent. In order to be able to describe the absorption process reliably, a comprehensive picture of the reaction network is necessary. In addition, even though the concentration of the molecular CO₂ dissolved in the liquid phase is very low, it determines both the rate of mass transfer from the gas to the liquid phase and the rate of reaction in the liquid phase. Therefore, its value together with the concentration of the reaction products has to be known precisely in order to be able to understand the overall reactive absorption process.

Chances and Pitfalls of *In-Situ* Irradiation NMR Spectroscopy

Jonas Kind¹, Ann-Kathrin Schönbein², Jasper J. Michels², Stefan Ortgies³, Alexander Breder³, and Christina M. Thiele¹

1. Clemens-Schöpf-Institut für Organische Chemie und Biochemie, TU Darmstadt, DE
2. Max Planck Institute for Polymer Research, Department of Molecular Electronics, Mainz, DE
3. Institut für Organische und Biomolekulare Chemie, Georg-August-Universität, Göttingen, DE

NMR spectroscopy is widely used for (*in-situ*) reaction characterisation and reaction monitoring e.g. to identify intermediate species or to quantify side product formation.

Irradiation of NMR samples inside the magnet is well known from photo-CIDNP experiments, where light is used to enhance NMR signal intensities. Furthermore, the combination of NMR and *in-situ* irradiation offers the possibility to characterise irreversible and reversible photochemical conversions on a large temporal scale [1,2].

By using recent results of two photochemical reactions we discuss benefits of *in-situ* irradiation NMR using LEDs and silica waveguides as well as unexpected pitfalls of this technique. We want to illustrate the scope of *in-situ* irradiation NMR in terms of irradiation intensities, sample homogenities and quality of the resulting spectra as well as comparability to batch experiments.

Firstly we show kinetic data of a reaction cascade yielding poly(2,5-bis-(2'-ethylhexyl)-1,4-phenylenevinylene) (BEH-PPV), a well-known OLED emitter material. A reactive *p*-quinodimethane is produced *in-situ* from the premonomer by adding potassium *tert*-butoxide below the threshold temperature of the thermal polymerization reaction. Polymerization of the *p*-quinodimethane can be induced by irradiation of the reaction mixture with UV light [3].

Secondly we show results on a dye sensitized seleno mediated photo catalytic lactonization of an alkenoic acid in the presence of oxygen [4]. Production of the lacton occurs via an isolatable seleno substituted lacton, which yields the final product after photo catalytic elimination. From batch experiments it is known that either the activation of the intermediate or the elimination to the product are rate limiting. Here we present data of an initial rate approach to identify the rate limiting step in the catalytic cycle.

1. Feldmeier, Christian; Bartling, Hanna; Riedle, Eberhard; Gschwind, Ruth M., *J. Magn. Reson.*, 232, 39-44, 2013.
2. Kind, Jonas; Kaltschnee, Lukas; Leyendecker, Martin; Thiele, Christina M.; *Chem. Commun.*, 52, 12506-12509, 2016.
3. Kuch, Serena; Vilbrandt, Nicole; Rehahn, Matthias, *Macromol Rapid Commun.*, 37 (10), 820-825, 2016.
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Ultrafast DOSY NMR of Hyperpolarised Mixtures

Ludmilla Guduff¹, Dennis Kurzbach², Daniel Abergel², Ilya Kuprov³, Carine van Heijenoort¹, and Jean-Nicolas Dumez¹

1. ICSN-CNRS, Gif-sur-Yvette, FR
2. ENS, Paris, FR
3. University of Southampton, UK

The sensitivity of solution-state NMR experiments can be increased by several orders of magnitude with dissolution dynamic nuclear polarization (D-DNP). This hyperpolarisation strategy is of great potential use for the analysis of low concentrated mixtures of small molecules. [1]. The amplified polarisation, however, only lasts for a few tens of seconds or less, because of longitudinal relaxation. Diffusion-ordered spectroscopy (DOSY), a method to separate the NMR spectra of components in a mixture, would be useful for the analysis of hyperpolarised mixtures. Conventional DOSY experiments, however, require long acquisition durations that are not compatible with D-DNP.

Here we show that hyperpolarised ^{13}C DOSY experiments can be recorded in a single scan, using a spatial encoding of the diffusion dimension. We first describe a pulse sequence for spatially encoded (SPEN) DOSY, based on the work of Keeler and Frydman [2,3], which accelerates experiments by an order of magnitude. The SPEN DOSY concept is generalised to 3D DOSY. [4]. We then use D-DNP to hyperpolarise a mixture of small molecules, together with SPEN DOSY for the acquisition of a time-series of ^{13}C DOSY spectra. Several modifications are made to the original sequence, in order to compensate for the effect of convection [5], and to make it possible to acquire a time-series of spectra from a single dissolution experiments. Throughout the analysis, numerical simulations are used, based on a Fokker-Plank formalism to describe simultaneously the spin and spatial dynamics. These simulations are key to characterise the pulse sequence and derive an improved model to obtain more accurate diffusion coefficients from SPEN DOSY experiments. Hyperpolarised ^{13}C DOSY is a promising tool for the monitoring of chemical reactions and the analysis of molecular interactions.

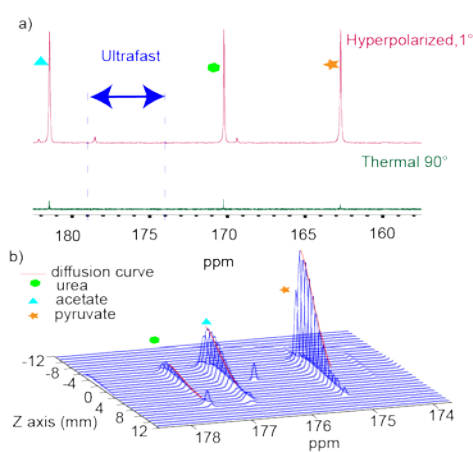


Figure 2: a) 1D ^{13}C spectrum. b) SPEN DOSY data.

1. Dumez, J.-N. et al., *P. Analyst* **2015**, *140*, 5860–5863.
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Tuesday, September 19th

11:00 - 12:35

We Will Rock You – Solid-State NMR Applications

Chair: Steven Brown

Speakers:

Ann-Christin Poeppler

Universität Würzburg, Würzburg (DE)

Pierre Thureau

Marseille (FR)

Luis Mafra

University of Aveiro, Aveiro (PT)

Kanika Sarpal (Upgraded Poster)

University of Kentucky, Lexington (US)

No Heavy Metal - NMR Crystallography of Metal Salts and Organometallic Compounds

Ann-Christin Pöppler^{1,2}, David Walker¹, and Steven P. Brown¹

1. Department of Physics, University of Warwick, Coventry, UK
2. Institute of Organic Chemistry, University of Würzburg, Würzburg, DE

Initially chosen as a robust proof of concept system for the study of very air- and moisture sensitive organolithium compounds, understanding the solid-state structure and properties of orotic acid monohydrate and its lithium and magnesium salts was not straightforward.

All three compounds are known to form hydrate structures [1], an important class of chemicals due to the influence of the incorporated water molecules on the physicochemical properties, e.g. bioavailability and stability. Based on their connectivities, hydrate structures can be divided into three subsets: i) isolated hydrates, ii) channel hydrates and iii) (metal) ion assisted hydrates.

The three compounds, represent a set of structures with, in principle, known single crystal X-ray structures, in which each individual compound belongs to a different class of hydrate. Orotic acid monohydrate contains water molecules in isolated sites, while both salt structures show water molecules coordinated to a metal ion. Additionally, in magnesium orotate octahydrate, water molecules are also arranged in channels. Their pharmaceutical potential is subject to discussion in literature [2].

A combined study by solid-state NMR, GIPAW (CASTEP) [3] calculations, powder X-ray diffraction and thermogravimetric analysis shows complexities in structure and dynamics of these compounds that go beyond the static view of the available crystal structures. Furthermore, first data for the analysis of the sensitive organolithium compounds is also presented.

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Investigation of Powders at Natural Isotopic Abundance using Solid-State NMR and Dynamic Nuclear Polarization

Pierre Thureau

Aix-Marseille University, CNRS, Marseille, France

We demonstrate here that solid-state NMR can be used to investigate powders, which cannot be studied using standard techniques such as single-crystal X-ray diffraction. More specifically, an efficient procedure for the assignment of ^{13}C resonances in natural abundance powders will be shown.[1] Furthermore, it will be shown that the sensitivity enhancement obtained using dynamic nuclear polarization (DNP) can be used to determine the conformation and crystal packing of pharmaceutical compounds.[2] The benefit of combining crystal structure prediction and DNP experiments will also be discussed. Finally, we will show that DNP experiments can be useful to reveal the early stage of polymorphic transformation in powders.

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Combined Solid-State NMR, Diffraction and Modeling Studies of Small Molecule Pharmaceuticals

Luís Mafra

CICECO - Aveiro Institute of Materials, Department of Chemistry, University of Aveiro, Campus Universitário de Santiago, 3810-193 Aveiro, PT

Solid-state NMR (SSNMR) is a powerful atomic-level characterization technique able to study the local crystalline/amorphous structure of bulk and dosage forms of active pharmaceutical ingredients (API).[1] Toward a better understanding of how small molecule pharmaceuticals self-assemble in the solid-state to yield distinct polymorphic/pseudopolymorphic forms, a combined experimental 1D/2D SSNMR, X-ray diffraction (XRD), and computational study of selected small molecule pharmaceuticals is presented.[2-5]

Although X-ray diffraction (XRD) provides a full description of the intra- and intermolecular distances and angles, the technique presents severe limitations in probing local interactions involving light atoms such as hydrogens. In contrast, SSNMR is particularly useful to study hydrogens and probe the strength and nature of protons engaged in hydrogen bond (HB) interactions, having the potential to discriminate among distinct polymorphs. Here we report the effect of crystal packing on the ^1H and ^{13}C chemical shifts (CS) of drug hydrates and anhydrides, including nonconventional HBs, $\pi\cdots\pi$ and $\text{CH}\cdots\pi$ contacts, are studied through periodic DFT calculations using the GIPAW-DFT formalism. It will be shown that NMR CSs can be sensitive detectors of hydration/dehydration states in highly insoluble antibiotics.[2]

Current approaches to modify physicochemical properties of APIs, without impacting its pharmacological behavior, include the development of new “multicomponent crystalline solid forms” that can be obtained using mechanochemical processes, namely liquid assisted grinding (LAG).[3,4] This presentation highlights the NMR discrimination between cocrystals and salts of distinct APIs, revealing that ^1H and ^{15}N SSNMR combined with GIPAW-DFT calculations can locate “XRD-disordered” hydrogen atoms. ^1H SSNMR detected unusually strong HBs associated with such disordered hydrogens through the presence of ^1H resonances shifted to very high frequencies (up to ca. 20.1 ppm).[3]

Recently, SSNMR became an important gadget in the process of crystal structure solution in powders. This is a non-trivial task and using powder XRD methods alone may often lead to the wrong structure solution. In this talk, a new hybrid approach for structure determination of crystalline solids, will be presented, based on the combination of SSNMR, XRD and an ensemble of computational-assisted structure solution tools including a genetic algorithm based on evolution-inspired operators repeatedly applied to populations of possible crystal structure solutions that evolve to eventually produce the best new offspring candidates. Such methodologies are demonstrated in a challenging multiple component crystal structure composed by flexible molecules - *trihydrate β -lactamic* antibiotic. [5]

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Insights into Amorphous Solid Dispersions of Felodipine Using Solid-state NMR Spectroscopy: Miscibility and Molecular Interactions

Kanika Sarpal and Eric J. Munson

College of Pharmacy, University of Kentucky, Lexington, KY, US

Phase separation in amorphous solid dispersions (ASDs) is still not clearly understood on the sub-nanometric scale [1], and further systematic investigations are still required. The phase separation behavior is influenced by many factors, such as the composition of drug-polymer blend, the type and the strength of drug-polymer interactions, and the method of preparation [2-4]. We demonstrated the use of solid-state NMR spectroscopy (SSNMR) to evaluate the role of the strength of drug-polymer hydrogen bonding (H-bonding) on the compositional homogeneity in ASDs of felodipine (FEL), a poorly water soluble drug, with poly(vinylpyrrolidone), or PVP, poly(vinylpyrrolidone-co-vinylacetate), or PVP/VA, and poly(vinylacetate) or PVAc. The dispersions were prepared at various drug loadings (50% to 90% w/w) via melt quenching. The blend scale miscibility was studied by examining the proton spin-lattice relaxation times in the laboratory and rotating frame ($^1\text{H } T_1$ and $T_{1\rho}$) for the drug and the individual polymer for each set of ASDs. Domain sizes were estimated via spin diffusion. The experimental data was used to elucidate the influence of the strength of drug-polymer H-bonding on the phase behavior of resulting dispersions. It was found that FEL:PVP and FEL:PVP/VA systems exhibited weak signs of nano phase separation especially for the compositions with lower polymer loadings with considerably small domain sizes. Whereas FEL:PVAc system showed pronounced signs of nano phase separation as the polymer amount decreased with larger domains. The extent of H-bonding in amorphous FEL and FEL:Polymer blends within each set of ASDs was quantified via deconvolution. The carbonyl region was used to selectively analyze the evolution of the various populations of coexisting species in the samples. The order of the strength/extent of drug-polymer H-bonding interactions was PVP > PVP/VA > PVAc. It was suggested that the strength of drug-polymer H-bonding interaction is one of the key factors in controlling the phase behavior of ASDs.

Our findings indicate that SSNMR is a useful tool for evaluating the spatial homogeneity with sub-50 nm resolution, where other conventional techniques fail, and drug-polymer interactions for the compositions of pharmaceutically relevant systems.

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Tuesday, September 19th
17:00 – 18:00

Round table discussion

NMR Data Reporting

Coordinated by:
Damien Jeannerat, Université de Genève

NMR Data Reporting

Coordinated by:

Damien Jeannerat, Université de Genève

This will be a roundtable discussion (Damien Jeannerat, Chair) based around a number of ongoing initiatives in creating and NMR data storage/reporting standards. These include Damien's NMRdata initiative and there are also RSC and IUPAC discussion around this topic. All SMASH attendees are welcome.

Wednesday, September 20th
09:00 - 10:35

Should I Stay or Should I Go? – Non-Covalent Interactions and Complexes

Chair: Elisabetta Chiarpin

Speakers:

Ben Davis
Vernalis (UK)

Julien Orts
ETH Zurich, Zurich (CH)

Rodrigo Carbajo
AstraZeneca (UK)

Serena Monaco (Upgraded Poster)
University of East Anglia, Norwich (UK)

Fragment Evolution Without Routine Crystallography

Ben Davis

Vernalis, UK

Fragment – and structure- based drug discovery typically require significant numbers of three dimensional structures of ligand/protein complexes. These structures have to be accurate, relevant and timely in order to guide medicinal chemistry efforts. However, many classes of biological target are not amenable to this approach due to a range of factors.

For example, the target may not crystallise at all, or may crystallise only in an apo form, and high-resolution NMR structures of larger proteins remain time consuming to determine. In these cases conventional structure-guided methods cannot readily be applied, increasing the difficulty in discovering and developing novel drugs against these target classes.

We have found that combining NMR data with modelling and chemical SAR can effectively steer a medicinal chemistry campaign from fragment to lead, particularly when accompanied by clear communication between the disparate groups involved.

We will discuss our experiences and development of approaches to identifying and characterising ligands of PPI targets, with particular reference to the anti-apoptotic protein Bcl2. These methods have proven to be generally applicable and successful, with two compounds developed using these methods now in Phase I clinical trials.

MMR² for Fast 3D Structure Determination of Protein-Ligand Binding Site Without Protein Resonance Assignment

Julien Orts¹, Marielle Aulikki Wälti¹, May Marsh², Laura Vera², Alvar D. Gossert³, Peter Güntert^{1,4}, and Roland Riek¹

1. ETH Zürich, Laboratory of Physical Chemistry, HCI F217, Vladimir-Prelog-Weg 2, Zürich, CH
2. Swiss Light Source, Paul Scherrer Institute, Villigen, CH
3. Novartis Institutes for BioMedical Research, Novartis AG, Basel, CH
4. Institute of Biophysical Chemistry Goethe University, Frankfurt, DE

X-ray crystallography molecular replacement (Rossmann et al. 1962) (MR) is a highly versatile tool for the detailed characterization of lead compound and binding modes in the pharmaceutical industry (Hillisch et al 2004). The two major limitations of its application to drug research are (i) the availability of a similar protein structure, which, in the area of structure-based drug design, is most often a complex of the protein with a lead compound, and (ii) obtaining well-diffracting crystals of the ligand-protein complexes of interest. While nowadays the first point is often not a limitation anymore, obtaining well-diffracting crystals might be difficult. In such situations structure determination of protein-ligand complexes by liquid-state NMR is a good option. Unfortunately, the established standard structure determination protocol (Cavanagh et al. 2007) is in general time-consuming, and a shortcut using available structural data as in the case of MR in X-ray crystallography is not available.

Here, we present *NMR*² (**NMR Molecular Replacement**), a MR-like approach in NMR to determine the structures of the binding pockets of ligands at atomic resolution. The calculation of structures of protein-ligand complexes relies on the collection of unassigned semi-quantitative inter-molecular NOE distance restraints and on previously solved structures. The *NMR*² method uses a high throughput structure calculation protocol, rather than a docking-scoring simulation. It is fast since it requires only a few days of measuring time and bypasses the time-consuming sequential assignment steps for the protein. When applied to the cancer-relevant HDMX protein, the *NMR*² method yielded the structure of a ligand protein complex with an accuracy below 1 Ångstrom for the binding pocket irrespective of the starting protein structure templates used. We will present multiple *NMR*² applications covering a peptidomimetic inhibitor and small molecules that bind strongly or weakly to protein receptors fully or partially labelled using methyl-specific isotope labelling. Our findings demonstrate that *NMR*² may open an avenue for the fast and robust determination of the binding pocket structure of ligand-protein complexes at atomic resolution.

NMR Free Ligand Conformations for Enhanced Structure Based Drug Design

Rodrigo J. Carbajo

AstraZeneca, Analytical & Structural Chemistry, Oncology IMED, Cambridge, UK

The increasing availability of protein-ligand x-ray crystallography data has transformed the ability to rationally design drugs and a number of success stories show how more quickly we can build ligand affinity in a structure enabled drug discovery project. However, despite the knowledge of the experimental bioactive ligand conformation and protein-ligand interactions, a challenge remains during a lead optimisation program to maintain or increase ligand affinity while tinkering with physical chemical properties. Although a number of computational tools are available to calculate the ligand conformational landscape in solution, these frequently fail to predict the actual most thermodynamically stable form, resulting in lack of progress with ligand affinity. We will show with examples from AstraZeneca oncology discovery projects, how measurement by NMR spectroscopy of the conformational landscape of free lead molecules in solution can aid and focus drug design hypothesis, leading to accurate predictions of affinity and physical chemistry properties.

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DiffErential EPitope Mapping- (DEEP) STD NMR to reveal the pharmacophore of a protein target

Serena Monaco¹, Louise E. Tailford², Nathalie Juge², and Jesus Angulo¹

1. School of Pharmacy, University of East Anglia, Norwich, UK
2. Institute of Food Research, Norwich Research Park, Norwich, UK

Saturation Transfer Difference (STD) NMR spectroscopy is extensively used to obtain epitope maps of ligands binding to protein receptors under fast exchange conditions [1], [2]. STD NMR reveals structural details of biomolecular recognition processes, which are fundamental to direct lead optimisation efforts in drug discovery.

Standard procedures seek uniform saturation of the receptor to identify regions of the ligand contacting the protein binding pocket. However, in this way, the experiment does not provide information about the “nature” of the amino acids surrounding the ligand in the bound state.

Here we report a novel protocol (DiffErential EPitope Mapping-STD NMR or DEEP-STD NMR) to identify the type of protein residues contacting the ligand [3]. We demonstrate that the approach constitutes a novel versatile method to orthogonally explore the nature (aliphatic, aromatic, polar or hydrophobic) of the amino acid residues lining the surface of the binding pocket and their orientation relative to the ligand.

As a proof of principle, we selected two relevant protein-ligand interactions from different areas of interest: i) the interaction of 3-nitrophenyl- α -galactoside (3NPG) with subunit B of Cholera Toxin (CTB) [4], well known to CTB inhibitor designers; and ii) the interaction of 2,7 anhydro Neu5Ac with the glycosyl hydrolase GH33 from *Ruminococcus Gnavus* [5], of great interest in gut microbiota fundamental research, as it is over-represented in individuals affected by Inflammatory Bowel Disease [6]. For both systems, high resolution X-Ray structures were available, allowing us to validate the protocol. The approach overall seems solid, versatile and promising for expanding the view of STD-NMR and going beyond its current limitations.

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Wednesday, September 20th
11:00 – 12:35

It's Not Unusual – Multinuclear and Inorganic Methods

Chair: Michael John

Speakers:

Markus Enders
Universität Heidelberg, Heidelberg (DE)

Pierre Florian
CEMHTI-CNRS Orleans, Orleans (FR)

Armando Navarro-Vázquez
Universidade Federal de Pernambuco, Recife (BR)

Olga Semenova (Upgraded Poster)
University of York, York (UK)

NMR Analysis of Small Paramagnetic Metal Complexes with Large Hyperfine Shifts

Marko Damjanovic, Markus Hiller, and **Markus Enders**

Anorganisch-Chemisches Institut, Heidelberg University, Heidelberg, DE

Solution NMR spectra of paramagnetic molecules have been recorded already in the early days of NMR spectroscopy and the basic theory describing the interaction of the unpaired electron with the NMR nucleus was developed soon after [1]. This has led to many applications where paramagnetic centers influence the appearance of NMR signals (e.g. Lanthanide shift reagents, paramagnetic relaxation enhancement (pre), pseudocontact shifts (pcs), etc). Especially in large biomolecules, the introduction of paramagnetic centers adds a valuable tool for obtaining additional structural data.

Some paramagnetic compounds show small paramagnetic shifts combined with small signal linewidths so that the typical toolbox of NMR experiments can be used for a straightforward interpretation of the data. However, many metal complexes show large paramagnetic shifts (i.e. hyperfine shifts) which makes the measurement and interpretation of these spectra more difficult. Therefore, NMR spectroscopy of small paramagnetic metal complexes has not become a standard technique in many laboratories. Large ^1H or ^{13}C NMR hyperfine shifts are the result of considerable unpaired electron delocalization to the measured nuclei leading to Fermi-contacts shifts (*fcs*) or non-averaged dipolar electron-nuclei interactions leading to pseudocontact shifts (*pcs*). The latter is large if the magnetic susceptibility is strongly anisotropic, which is the case for many Lanthanide complexes and for a few d-block complexes with spin orbit coupling.

Anisotropy of the Magnetic Susceptibility (*AMS*) leads to partial alignment of the molecules in the magnetic field of the NMR spectrometer so that Residual Dipolar or Quadrupolar Couplings (*RDC* or *RQC*, respectively) become visible [2,3]. *AMS* is the prerequisite for so-called Single Molecule Magnets (*SMMs*). Many recently developed *SMMs* consist of one or only a few Lanthanoid or d-block metal ions and these systems usually give well-resolved ^1H or ^{13}C NMR spectra. The difficulty in the interpretation of such NMR spectra arises from the presence of more than one dominating NMR shift contribution. This problem can be solved by using quantum chemical calculations combined with strategies, which increase the number of available experimental NMR data. The latter is achieved by measuring ^1H and ^{13}C NMR spectra, by the introduction of a more complex ligand substitution pattern or by synthesis of an isostructural series of Lanthanide complexes [4].

The comprehensive paramagnetic NMR analysis, allows not only the extraction of many structural parameters in solution but also the determination of important magnetic properties like size and sign of the magnetic anisotropy [5], determination of zero-field splitting [6] or extraction of crystal field parameters. The recent developments for measuring and interpreting NMR spectra of small paramagnetic metal complexes helps to quickly evaluate new candidates for better *SMMs* but also improves the NMR method for paramagnetic molecules in general.

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Elucidation of the Topological and Chemical Order in Materials by Multi-Nuclear Solid-State NMR

Pierre Florian¹, Franck Fayon¹, Sohei Sukenaga², Yann Morizet³, and Dominique Massiot¹

1. Conditions Extrêmes et Matériaux: Haute température et Irradiation (CEMHTI-CNRS), Orléans, FR
2. Institute of Multidisciplinary Research for Advanced Materials (IMRAM), Sendai, JP
3. Université de Nantes, Laboratoire de Planétologie et Géodynamique, Nantes, FR

Local order, as opposed to the long-range order of the ideal crystalline structures, can be considered as an intrinsic characteristic of real materials, being often the clue to the tuning of their properties and their final applications. While ordering can be easily assessed in two-dimensional imaging techniques with resolution approaching the atomic level, the diagnostic, description, and qualification of local order in three dimensional systems is much more challenging.

Solid-state nuclear magnetic resonance and its panel of constantly developing new instruments and methods enable local, atom selective characterization of structures and assemblies ranging from atomic to nanometer length scales. This opens unique opportunities for characterizing a variety of materials, ranging from crystalline compounds to amorphous or glassy materials, for which we show that it becomes possible to separate topologic, geometric and chemical contributions to the order or disorder.[1]

Among the most efficient technics available in our toolbox are the multi-nuclear double-resonance experiments. In the context of material science, those approaches imply working with a wide type of nuclei such as ²⁷Al, ¹¹B, ²⁹Si, ¹³P, ¹⁷O, ¹⁹F, etc. and can therefore appears to be somewhat difficult to. We will show that the quality of the retrieved information is worth the efforts especially when disorder is present in the system, hiding the fine structures of the solid-state NMR spectra.

Using examples extracted from our work on oxide glasses we will show how scalar- or dipolar-based sequences can decipher the complex spectral line shapes and provide new structural insights. Technics such as REDOR, refocused INEPT or HMQC involving two “X” nuclei will be illustrated for spectral editing as well as two-dimensional correlations addressing their specificities with regards to their use for solid-state materials.[2,3] The case of quadrupolar nuclei will be largely discussed in the context of dipolar recoupling schemes or presence of scalar couplings.[4]

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A Hyphenated Computational Protocol for Analysis of Natural Abundance ^2H Residual Quadrupolar Couplings in (Chiral) Oriented Solvents

Armando Navarro-Vázquez^{1,2}, Philippe Berdagué², and Philippe Lesot²

1. Departamento de Química Fundamental, Universidade Federal de Pernambuco, Recife, BR
2. Institute of Organic Chemistry and Institute for Biological Interfaces Karlsruhe, Institute of Technology (KIT), Karlsruhe, DE
3. RMN en Milieu Orienté, ICMMO, UMR CNRS 8182, Université Paris-Sud / Université Paris-Saclay, FR

Natural abundance deuterium 2D-NMR (NAD 2D-NMR) in (chiral) oriented solvents is a powerful spectroscopic approach, essential for multiple and original analytical applications, from stereochemistry to the determination of site-specific natural (D/H) isotopic distribution.[1] Besides, due to the large value of the quadrupolar interaction the ^2H residual quadrupolar couplings (^2H -RQC) can furnish high-precision information about the structural, conformational and orientational properties of small organic chiral or prochiral molecules in aligning chiral media such as polyglutamate-based lyotropic systems (PBLG, PCBLL, PELG).[2]

To fully exploit and further explore the potential of anisotropic NAD NMR, we developed a new integrated computational protocol, based on MSpin-RDC software,[3] was here specifically designed for the analysis of ^2H -RQC extracted from anisotropic NAD-NMR spectra[3] in small molecules. ^2H -RQC anisotropic data, under-exploited so far, can be easily merged in this approach with other anisotropic NMR observables such as residual dipolar couplings (RDC) or even residual chemical shift anisotropies (RCSA) for linear fitting of the alignment tensor components.[2,4]

From experimental data recorded on a 14 T NMR spectrometer equipped with a ^2H cryogenic probe,[1,2] subtle but detectable differences can be observed and quantified for the orientational distribution of enantiomers or enantiotopic directions of prochiral molecules interacting with chiral liquid crystals as presented here. Additionally, structural elucidation problems, such as prochiral assignment in methylene groups can be easily handled using the proposed approach.[3]

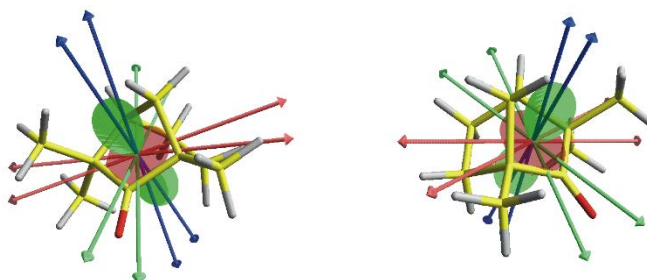


Figure 3. Relative orientation of inertia and alignment tensors principal frame in fenchone enantiomers from ^2H -RQC data obtained in PBLG.

Acknowledgments

P.L. and P.B. acknowledges both the CNRS and the University of Paris-Sud for their recurrent funding of fundamental research. A.N.-V. thanks KIT for a GästwissenschaftlerStipendium and UFPE for a visiting professorship as well as the HGF programme BIFTM, ChemPhysChem 10.1002/cphc.201601423 This article is protected by copyright. All rights reserved. - 25 - the DFG 555 (instrumentation facility Pro2NMR, LU 835/11), and FACEPE (APQ-0507- 1.06/15) for financial support.

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Hyperpolarised Low-Field NMR for Industrial Reaction Monitoring

Olga Semenova¹, Peter Richardson¹, Andrew Parrott², Alison Nordon², Meghan Halse¹,
and Simon Duckett¹

1. Centre of hyperpolarisation of Magnetic Resonance, Chemistry Department
University of York, York, UK
2. Chemistry Department, Strathclyde University, Glasgow, UK

Reaction monitoring is an important part of synthetic development, scale-up procedures and industrial quality control [1]. Process analytical tools (PAT) serve to control these steps in industry. Reliable analytical tools that can be used to follow a process are required. Low-cost benchtop NMR spectrometers that are based on permanent magnets with field strengths of around 1T offer the opportunity to develop NMR as an analytical tool for continuous reaction monitoring within an industrial site [2]. As a consequence of the relatively low magnetic field, these instruments suffer from low sensitivity and reduced chemical shift dispersion. Here we seek to investigate the viability of NMR reaction monitoring at low field by increasing sensitivity via the *para*-hydrogen (*p*-H₂) hyperpolarisation technique SABRE (Signal Amplification By Reversible Exchange) [3].

To illustrate this concept, we identify and follow a well-defined model reaction, such as the conversion of an amine into an amide. In this work, we describe studies on the level of SABRE enhancement seen for mixtures of reactant and product molecules in order to develop methods to quantitatively follow a reaction using hyperpolarisation. In addition, we investigate the effect of the SABRE process on the model reaction and demonstrate that the expected reaction products form in the presence of the SABRE catalyst. We show that the hyperpolarised NMR signals change with time as the starting material and product molecules concentrations in the reaction mixture evolve, indicating the viability of this method for reaction monitoring. In addition to this proof-of-concept work carried out on a standard high-field (400 MHz) NMR spectrometer, we present preliminary data from a benchtop (43 MHz) instrument to illustrate the feasibility of our approach.

Key words: reaction monitoring, NMR, hyperpolarisation, benchtop.

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Wednesday, September 20th
14:00 – 15:00

Workshop 3

**We Can Work It Out – Calculations of
Structures and NMR Parameters**

Coordinated by:
Giuseppe Bifulco, Università di Salerno (IT)

We Can Work It Out – Calculations of Structures and NMR Parameters

Coordinated by:

Giuseppe Bifulco, Università di Salerno (IT)

The recent progress in the fields of NMR and of quantum chemistry has contributed to a great acceleration in the structural determination of complex organic molecules. Recently, the substantial potential offered even from the common personal computer has made possible the use of DFT methods in the optimization of the geometries and calculation of the properties of medium and high molecular weight compounds. For this reason the world of quantum chemical NMR parameter calculation has attracted the interest not only of the theoretical chemists, but also of the experimental NMR spectroscopists. In this workshop the structural study of organic compounds by means of an integrated DFT-NMR approach will be described. In particular, both theoretical and practical aspects of NMR parameter calculation by quantum mechanical methods will be discussed, with special regard toward the modern protocols involved in the structural and stereochemical assignment of natural and synthetic organic compounds. Questions regarding problems/limitations/future directions are encouraged during the workshop.

Wednesday, September 20th
15:30 - 17:05

**Go Your Own Way – Past SMASH
Hot Topics Revisited**

Chair: Christina Thiele

Speakers:

Burkhard Luy
Karlsruhe Institut für Technologie, Karlsruhe (DE)

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Broadband Pulses Revisited

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Based on an overview talk, the various types of broadband compensated pulses are introduced and some hints to their usage given. In addition, several new developments in broadband pulse design, e.g. the design and application of broadband pulses with scalable effective flip angles, J-compensated broadband ^1H , ^{13}C -pulse sandwiches and saturation pulses for ultrabroadband applications, are demonstrated.

Small Microcoils Are Still Hot

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Commercially-available NMR probe heads and tranciever coils are very costly, impeding in-house modification of these for dedicated applications, such as on-flow and mass and/or volume-limited analyses. Since the pioneering work of Sweedler and collaborators on microliter solenoidal NMR coils two decades ago,[1] several groups have looked into alternative approaches to develop small-volume NMR detection devices exploiting different coil geometries.[2]

In the past few years, we have exploited mainly lower nanoliter detection volume planar spiral microcoils for their excellent mass sensitivity, required among others for on-flow reaction monitoring, allowing the rapid determination of kinetic and thermodynamic parameters.[3,4] In addition, these coils show fascinating broad-band properties allowing homo- and heteronuclear multidimensional NMR experiments with a single coil in a non-tuned circuit.[5,6] Although in-house designed for dedicated purposes, these glass NMR chips still require clean-room fabrication steps and we therefore started investigating alternative approaches.

Recently, we developed a simple method for the manufacturing of microfluidic devices in a single block of PDMS, coined *ESCARGOT*: Embedded SCAffold RemovinG Open Technology.[7] Using this procedure it is possible to include not only channels in the PDMS device, but also stirring bars, electronics and even fully functional solenoidal NMR coils. This fabrication method is of interest for those willing to use microfluidic NMR devices without having much knowledge on the topic or access to microfabrication equipment and clean rooms.

Here we will present our latest home-built NMR probes with different coil geometries and detection volumes in the nL-to- μ L range for in-line and in-situ monitoring, with focus on improved concentration sensitivity, offering new horizons for home-built small-volume and low-cost NMR probe designs.[8]

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Hot Blooded, Check Out CPMG

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Purity assessment of small molecules by NMR can be challenging when they exhibit multiple conformations at room temperature. Conformational isomers are observed when chemical exchange is slow on the chemical shift timescale ($k_{ex} < \Delta\omega$). This exchange is overcome and conformational isomers recognized by increasing the temperature until signal coalescence is observed ($k_{ex} \sim \Delta\omega$), followed by signal narrowing once the fast exchange regime is reached ($k_{ex} > \Delta\omega$). An alternative way to detect conformational changes is through the application of Carr-Purcell-Meiboom-Gill relaxation dispersion (CPMG-RD) sequences. These NMR experiments are traditionally used for protein conformational studies,¹ although application to small molecules through ligand-target binding studies have been reported.² Presented herein is the application of CPMG-RD to the dynamics that underlie chemical exchange phenomena in a class of structurally complex antibiotics. Some salient features of CPMG-RD are the ability to precisely quantitate the chemical exchange rate constants and populations, which is demonstrated with this compound class.

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General Approach to Access Long-Range ^1H - ^1H RDCs

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Residual dipolar couplings (RDCs) are a powerful means for conformational and configurational analyses [1, 2]. For small molecules, ^1H - ^{13}C one-bond couplings are mostly exploited. However, the quality of, for instance, configurational analysis greatly depends on the number of RDCs that sample independent orientations, which may not always be provided by one-bond couplings alone. Ways to obtain long-range RDCs are thus in high demand. Long-range ^1H - ^1H RDCs would be an obvious choice, but are very challenging to access. The main reason is that, in alignment media such as PBLG, the high abundance of ^1H - ^1H RDCs results in broad multiplets that overlap and from which individual splittings are not resolvable. Here, we propose a general approach to obtain ^1H - ^1H RDCs.

We demonstrate that the recent PSYCHEDELIC experiment [3] gives access to long-range $^n\text{T}_{\text{HH}}$ couplings at a resolution close to the natural linewidth. PSYCHEDELIC delivers a 2D J-resolved spectrum with absorption mode lineshapes and with only ^1H - ^1H couplings to one or more chosen spins, thus allowing coupling measurement as simple doublets at pure shift resolution and resolving both the issues of spectral overlap and multiplet complexity. The experiment is based on PSYCHE [4], which allows broadband homodecoupling at good sensitivity and tolerance to strong coupling. The tolerance to strong coupling can be further enhanced by introducing frequency-swept 180° pulses applied during gradients [5]. Although PSYCHEDELIC delivers the magnitude of the ^1H - ^1H coupling, it offers no sign information, which is crucial for RDC interpretation. In isotropic samples, E.COSY or similar experiments can provide relative sign information, but in weakly aligned samples the overflow of ^1H - ^1H couplings complicate cross-peak analysis. Solutions based on P.E.COSY or z-COSY using similar principles as PSYCHEDELIC to provide relative sign information of only selected couplings will be introduced.

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¹H NMR Experiments in the Conformational Analysis of Amino Acid Derivatives Containing a Sulfur Atom

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The conformational analyses of L-methionine (L-Met) (**1**) and L-cysteine (L-Cys) (**2**), the two essential amino acids containing a sulfur atom, were performed including quantum theory calculations. Recent studies [1,2] have shown that an understanding of the interactions between the side chain and the *backbone* in an amino acid is essential to address the conformational preferences of peptides and proteins. Thus, the study of their *N*-acetylated derivatives using NMR data and quantum chemical calculations, including the solvent effect (IEF-PCM), combined with ³J_{HH} spin-spin coupling constants analyses were carried out.

Theoretical calculations (ω B97X-D/aug-cc-pVTZ) resulted in 8 and 4 more stable conformers for **1** and **2**, respectively, which are in three different dispositions (**a**, **b** and **c**). Furthermore, the calculations showed that the most stable conformers of **1** and **2** are in the form **a**, but when the dielectric constant (ϵ) of the medium is increased, these conformers are destabilized and have their populations decreased, indicating that the conformational equilibrium of **1** and **2** are affected by the solvent changes.

The solvent effect was also evaluated through experimental and theoretical ³J_{HH} coupling constants. The observed values for ³J_{H_aH_{b1}} and ³J_{H_aH_{b2}} confirm the predominance of conformers **a** of **1** and **2** in the less polar solvent (chloroform). When ϵ is increased (from CDCl₃ to DMSO), an increase in the ³J_{H_aH_{b1}} is observed and it indicates that conformers **c** of **1** are favoured in the conformational equilibrium, since this disposition has a hydrogen H_a *anti* to H_{b1}. Unlike **1**, in more polar solvents conformers **b** of **2**, which have H_a *anti* to H_{b2}, have their populations increased due to an increase in the ³J_{H_aH_{b2}}. Thus, the changes in the values of ³J_{H_aH_{b1}} and ³J_{H_aH_{b2}} support our theoretical findings that the conformational equilibria of **1** and **2** are affected by the solvent change. The theoretically calculated ³J_{HH,calc} are in agreement with the experimental data and reproduces well the trend observed by these compounds.

To investigate the effects (steric, hyperconjugation and hydrogen bonding) ruling the conformational preferences of the studied compounds, NBO, QTAIM and NCI analyses were carried out, as in a previous work in this laboratory [3]. These analyses showed that not only a specific interaction rules the conformational preferences of **1** and **2**, but an interplay between steric repulsion and hyperconjugation

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2

Identification of Non-Intentionally Added Substances in Packaging Materials by Pure-Shift NMR

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Non-intentionally added substances (NIAS) are undesired chemical species present in formulated materials. The origin of the NIAS can vary; for example, impurities and/or decomposition of the starting materials, byproducts formed during side reactions etc. Independently of the source of contamination, the presence of NIAS in food packaging materials poses serious questions regarding the potential health risks associated with the consumer user. As consequence, the characterization of the low molecular weight fraction, especially as it relates to oligomers or reaction products of the starting monomers, is of fundamental importance.

Given the complexity of these formulated materials, the identification of NIAS, directly in the final packaging polymer or in the migration samples, represents one of the most challenging and complicated tasks performed by analytical groups within the chemical industry. The very low LOD required (in the ppb range) makes carbon measurements ineffective even when the samples are pre-concentrated several times. On the other hand, the concomitant presence of several species makes the proton NMR spectrum too crowded, limiting the amount of useful information that can be obtained.

In this work, we show a combination of 1D and 2D Pure-Shift techniques we could overcome the intrinsic resolution limitation of proton experiments and were able to identify several NIAS in a particularly challenging case. The implementation of these novel experiments enabled the use of NMR for the identification of NIAS in food packaging materials.

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3

NMR Characterization of Complex Natural Products: Opportunities and Challenges in Structure Elucidation

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NMR provides powerful structural elucidation tools that are particularly well suited for natural products studies. Comprehensive spectroscopic characterization of a native metabolite may be sufficient to fully assign a planar structure. However assignment of the relative and absolute configuration of a molecule when there are multiple stereogenic centers often requires the development and application of additional experimental strategies. Many successful approaches in this regard rely on the formation of appropriate derivatives for more detailed NMR study. Since natural products are often obtained in very limited quantities, micro-scale chemical manipulations and the ability to analyze the structure of the resulting products is often key to the complete structural and configurational assignment of complex metabolites. Anisotropic NMR parameters such as residual dipolar coupling (RDC) and residual chemical shift anisotropy (RCSA) provide a powerful and complementary means to help assign and verify structures deduced from conventional NMR analyses.¹⁻⁶ Application of a wide spectrum of NMR techniques and methodologies will be described in the structural elucidation of a new phosphomacrolide marine natural product with 8 stereogenic centers.

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4

Determination of Covalency in the Bonding of Trivalent Actinides with Soft N-Donor Ligands by NMR

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In possible future actinide recycling processes the separation of the chemically highly similar trivalent actinide (An) ions from lanthanide (Ln) ions by liquid-liquid extraction is a challenging task. A number of extraction ligands with the required selectivity (separation factors >100) for the separation of An(III) from Ln(III) have been developed. Among those, soft N-donor ligands of the bis-triazinylpyridine type (BTP) have been very successful [1, 2]. However, the molecular origin of these ligands' selectivity remains elusive.

NMR spectroscopy of the paramagnetic An(III) complexes and their comparison to structurally analogue Ln(III) complexes offers insights into the bonding situation. The overall paramagnetic chemical shift consists of the pseudo-contact shift (PCS), which is transmitted through-space, and the Fermi contact shift (FCS), transmitted through-bond. By separation of the two contributions, important information on the share of covalency in the An-ligand-bond is obtained [3]. As the coordination involves the nitrogen lone pair in the aromatic BTP core, ¹⁵N NMR is an especially valuable tool. By ¹⁵N isotope enrichment of several extraction ligands the chemical shifts even for the coordinating nitrogen atoms were observed throughout the Ln(III) series and for Am(III) complexes. Our results indicate that an increased share of covalency in the An-ligand bond is the driving force for the observed selectivity [4, 5].

The aliphatic side-chains of the aromatic cores of the most commonly used extraction agents do not only influence the solubility in organic solvents. They are crucial for the stability against hydrolysis and radiolysis, and also have strong effects on the coordination kinetics and complex stability. We showed by comparison of extraction ligands with different side chains (*n*Pr vs. *i*Pr) that differences in coordination behaviour are not due to changes in the electronic structure but rather due to sterical reasons or different complex hydration [6].

While NMR is established as a routine method in inorganic and organic chemistry, it has so far found only limited use on complexes of paramagnetic metal ions in particular of the trans-uranium elements. We will show the merits of this method, which can deliver information that have previously not been available from other methods that have been used in actinide research for years.

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5

Maximising Dispersion in COSY Experiments

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Two-dimensional NMR experiments have traditionally been recorded with limited digital resolution, primarily to limit the size of data sets and the time taken to acquire the data. This has had the effect of limiting the dispersion of signals, and can lead to problems in assigning very crowded spectra.

Recent advances in computing power and in techniques such as Pure Shift methods [1] and compressed sensing [2,3] mean that it is now practical to acquire some two-dimensional NMR experiments at digital resolutions that approach the intrinsic resolution limits of the instrumentation and sample [4].

COSY experiments represent a particular challenge because, in contrast to experiments such as HSQC, the transfer function evolves in the same time period as the chemical shift. This means that if we look to extend the evolution period to achieve high digital resolution we also see correlations due to smaller couplings, often over more than 3 bonds, which make interpretation of the spectra of unknown structures more difficult.

Here we present an experiment that overcomes many of these problems and allows the acquisition of Pure Shift COSY spectra at very high digital resolution while minimising the introduction of unwanted long-range correlations. The utility of such an experiment is demonstrated.

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6

Robust qNMR Automated Analysis

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The use of NMR for quantitation is an everyday occurrence. A popular application is the purity determination of single organic compounds, which is, for example, a staple requirement of the pharmaceutical industry.

The total uncertainty in the measurement is a summation of all stages in the analysis, and is well understood. Remarkably, quantitation by NMR (qNMR) routinely achieves results with <2% total uncertainty with only modest effort. With more effort, and skilled operators, this figure can be quite routinely reduced by an order of magnitude. qNMR is a validated primary procedure.

In many areas of NMR analysis the trend is towards automation, and implementation of more complex NMR analysis tasks to analysts having less NMR training and basic understanding. Indeed, the logical extension is to extend this to fully automated analysis. This imposes new challenges to NMR software capability because of the need for extremely reliable and sophisticated algorithms and work flows. An extension to automated qNMR adds a layer of necessary robustness, given the nature of the analysis and the regulated environments where it is often applied.

Here we will focus on NMR signal processing and analysis requirements in the context of qNMR. Many previously routine tasks require rethinking and new algorithms to ensure effective quantitation.

qNMR analysis itself is not an especially difficult procedure in principle, but there are many ways that it can be adversely influenced through naïve, improper operation. A surprisingly sophisticated analysis system is therefore required to ensure reliable results for a very high percentage of samples. We have developed algorithms and general capabilities that addresses automated processing and analysis even with these quite challenging requirements. We show that fully automated qNMR processing and analysis is achievable when such a combination of elegant techniques are applied together.

7

Hyperpolarised Low-Field NMR for Industrial Reaction Monitoring

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Reaction monitoring is an important part of synthetic development, scale-up procedures and industrial quality control [1]. Process analytical tools (PAT) serve to control these steps in industry. Reliable analytical tools that can be used to follow a process are required. Low-cost benchtop NMR spectrometers that are based on permanent magnets with field strengths of around 1 T offer the opportunity to develop NMR as an analytical tool for continuous reaction monitoring within an industrial site [2]. As a consequence of the relatively low magnetic field, these instruments suffer from low sensitivity and reduced chemical shift dispersion. Here we seek to investigate the viability of NMR reaction monitoring at low field by increasing sensitivity via the *parahydrogen* ($p\text{-H}_2$) hyperpolarisation technique SABRE (Signal Amplification By Reversible Exchange) [3].

To illustrate this concept, we identify and follow a well-defined model reaction, such as the conversion of an amine into an amide. In this work, we describe studies on the level of SABRE enhancement seen for mixtures of reactant and product molecules in order to develop methods to quantitatively follow a reaction using hyperpolarisation. In addition, we investigate the effect of the SABRE process on the model reaction and demonstrate that the expected reaction products form in the presence of the SABRE catalyst. We show that the hyperpolarised NMR signals change with time as the starting material and product molecules concentrations in the reaction mixture evolve, indicating the viability of this method for reaction monitoring. In addition to this proof-of-concept work carried out on a standard high-field (400 MHz) NMR spectrometer, we present preliminary data from a benchtop (43 MHz) instrument to illustrate the feasibility of our approach.

Key words: reaction monitoring, NMR, hyperpolarisation, benchtop.

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Overcoming Overlap in a Low-Field Reaction Monitoring System Using CRAFT (or Times, They Are A Changin')

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There are significant advantages to using NMR in a Process Analytical/Reaction Monitoring role. Given the operating environment and financial constraints involved with superconducting magnets, low-field systems offer significant advantages. [1] Signal overlap is a problem even at high field, however, and the requirement to work in protonated solvents means that every analysis is challenged by the dynamic range of the system. CRAFT (Complete Reduction to Amplitude Frequency Table) [2, 3] offers a unique alternative to the traditional frequency-based approach to NMR data processing. It provides the advantage of returning high-quality qNMR results from complex spectra and powerful post-acquisition tools to handle large solvent resonances. This presentation will demonstrate results from our implementation of a 60 MHz NMR system using CRAFT-based data reduction to better understand reaction chemistries in a process chemistry environment, showing the advantages provided by time domain analysis.

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9

Simplifying the Analysis of Low Level Impurities in High Dynamic Range Mixtures

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The analysis of high dynamic range mixtures is always a challenge, and it is even more demanding when the main interest lies in the detection and quantification of low level components of the sample. In pharmaceutical chemistry, for example, all components above 0.1 % of a main active ingredient need to be identified and quantified. NMR can be very useful in this area, but the presence of ¹³C satellites complicates matters. ¹³C isotopomer signals have similar intensities, and often similar chemical shifts, to the resonances of interest. Broadband ¹³C decoupling is always a possibility, but can cause severe sample heating if good resolution is needed. Moreover, for nuclei with large secondary isotope shifts like ¹⁹F this approach does not solve the problem, merely halving the number of ¹⁹F-¹³C signals. Here, we suggest an alternative approach for dealing with ¹³C isotopomer signals in both ¹H and ¹⁹F NMR, which relies on their suppression rather than decoupling.

In the case of ¹⁹F, a low-pass ¹J_{CF} filter followed by a heteronuclear 2DJ experiment allows both the one-bond and long range satellites to be suppressed [1]. This 2D acquisition scheme bypasses the problem caused by secondary isotope shifts, since the final 1D spectrum is extracted as an integral projection onto the *F*₂ axis for an *F*₁ range close to *F*₁ = 0, which includes only ¹²C isotopomer signals. The final clean spectrum is obtained at only very modest cost in sensitivity. ¹⁹F-¹⁹F coupling is a potential problem, but can usually be dealt with by using selective 180° ¹⁹F pulses.

A related approach can be used in the case of ¹H NMR. Here the problem of homonuclear coupling is dealt with by using a perfect echo [2], during which ¹³C 90° pulses are used to convert ¹H coherence of ¹³C isotopomers into unobservable MQC. This is followed by a z-filter, to allow the acquisition of clean ¹H spectra for ¹²C-bound protons only. The perfect echo provides time for four successive 90° carbon pulses; their timing is optimised to allow suppression over a wide range of ¹J_{CH}. Because ¹H secondary isotope shifts are very small, long range ¹³C satellites are much less of a problem than in ¹⁹F NMR and are generally hidden under the parent signals, but if necessary low power ¹³C decoupling can be applied to collapse them.

In both the ¹⁹F and the ¹H experiment the suppression of the ¹³C satellites signals leads to clean spectra with no visible ¹³C couplings, facilitating the analysis of high dynamic range mixtures.

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Ultraclean Pure Shift NMR: Cyclic Sideband Suppression

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In all but the simplest ¹H spectra, signal overlap is a problem. Pure shift NMR experiments can offer a vital resolution improvement, suppressing the effects of homonuclear couplings J , but the spectral simplicity that results comes at a price. Current pure shift experiments typically work by periodically refocusing the effects of J , either during a 1D acquisition (real-time methods) or in 2D mode (interferogram methods). The pure shift FID is recorded or constructed as a series of “chunks” of data of duration $1/\text{sw1}$, in which J is refocused at the midpoint. The choice of sw1 requires a compromise between the quality of decoupling and either sensitivity (interferogram methods) or linewidth and spectral purity (real-time methods). Because J is only perfectly refocused at the midpoint of each data chunk, the pure shift FID retains a slight periodic modulation, which Fourier transforms into “chunking sidebands” spaced at multiples of sw1 either side of the pure shift peaks. With a typical sw1 of 40 Hz such sidebands are of the order of a few % or smaller and can generally be neglected, but in high dynamic range samples they can obscure real signals of interest.

The impact of such sidebands can be reduced by spreading them out, averaging spectra measured with different values of sw1 [1]. Here in contrast we propose to suppress them completely, by manipulating the phase of the residual J modulation. Small (ms) changes in pulse sequence timing are used to change the phases of sidebands over a 360° range while leaving the parent signals unaffected, so that averaging gives a clean spectrum. Using 2^N different timings during time averaging allows sidebands to be suppressed to arbitrary order N .

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11

Analyzing Highly Complex Mixtures: Combining Tailored Multilinear Relaxation and Diffusion Experiments with PARAFAC Tensor Analysis

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To extract sensible NMR information from very complex mixtures is an important but also a formidable task. In this context PARAFAC (PARAllel FACtor analysis) [1], a powerful extension of component analysis to higher order arrays, has been used successfully for relatively simple mixtures. The power of PARAFAC stems from its ability to analyze multilinear data, i.e data that vary independently as a function of three or more variables. Previously, for NMR data, this has meant a combination of a spectroscopic dimension with diffusion encoding (as in a DOSY experiment) [2] and a third domain such as relaxation [3] or concentration variation [4]. Using changes in concentration can easily cause e.g. chemical shift changes, that are detrimental to multivariate analysis; and relaxation generally violates the multilinearity required for PARAFAC analysis as different spins in a given molecule have different relaxation times, which has limited the use of PARAFAC in earlier experiments. Recently, we have proposed a step towards NMR experiments specifically tailored for PARAFAC analysis: *Relaxation-Encoded Selective TOCSY* (REST) [5], in which differences in relaxation between spins can be exploited to separate the signals of different spin systems by producing multilinear data. It uses a combination of selective excitation and isotropic mixing to ensure that all the signals measured for a given species originate from a single proton [6]. Combining REST₁ and REST₂ in a single experiment (REST₁₋₂), or REST and diffusion (REST-DOSY), should allow the application of PARAFAC to very complex mixtures. Extending the REST experiment to higher order data is significantly more efficient than using bilinear multivariate methods such as SCORE [7]. In principle, this approach could allow efficient analysis of highly complex mixtures with a large number of chemical species at a high dynamic range.

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12

Robust and Reliable Quantification of Phospholipids in Edible Oils

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Crude plant oils will form a solid sediment called gum upon storage. These gums contain mainly phospholipids. To obtain commercial edible oil the gum has to be removed, this process is known as degumming. The oldest form degumming is obtained by mixing crude oils with water and the solid formed is removed by centrifugation. Enzymatic degumming can be applied to reduce oil losses during degumming. To monitor degumming processes a robust and accurate phospholipid quantification is needed. Several techniques have been applied for the phospholipids quantification in edible oils, such as ICP, LC and TLC. However, in the past decade ^{31}P NMR has shown to have advantages above the previously mentioned techniques because of the possibility to quantify different phospholipids directly from oil in an accurate and fast manner. In this report, we show the application and performance of 1D ^{31}P NMR method for quantification of several phospholipids in crude and refined oils. The method is robust and has a variation lower than 5% for most phospholipids

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Ultrafast DOSY NMR of Hyperpolarised Mixtures

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The sensitivity of solution-state NMR experiments can be increased by several orders of magnitude with dissolution dynamic nuclear polarization (D-DNP). This hyperpolarisation strategy is of great potential use for the analysis of low concentrated mixtures of small molecules. [1]. The amplified polarisation, however, only lasts for a few tens of seconds or less, because of longitudinal relaxation. Diffusion-ordered spectroscopy (DOSY), a method to separate the NMR spectra of components in a mixture, would be useful for the analysis of hyperpolarised mixtures. Conventional DOSY experiments, however, require long acquisition durations that are not compatible with D-DNP.

Here we show that hyperpolarised ^{13}C DOSY experiments can be recorded in a single scan, using a spatial encoding of the diffusion dimension. We first describe a pulse sequence for spatially encoded (SPEN) DOSY, based on the work of Keeler and Frydman [2,3], which accelerates experiments by an order of magnitude. The SPEN DOSY concept is generalised to 3D DOSY. [4]. We then use D-DNP to hyperpolarise a mixture of small molecules, together with SPEN DOSY for the acquisition of a time-series of ^{13}C DOSY spectra. Several modifications are made to the original sequence, in order to compensate for the effect of convection [5], and to make it possible to acquire a time-series of spectra from a single dissolution experiments.

Throughout the analysis, numerical simulations are used, based on a Fokker-Plank formalism to describe simultaneously the

spin and spatial dynamics. These simulations are key to characterise the pulse sequence and derive an improved model to obtain more accurate diffusion coefficients from SPEN DOSY experiments. Hyperpolarised ^{13}C DOSY is a promising tool for the monitoring of chemical reactions and the analysis of molecular interactions.

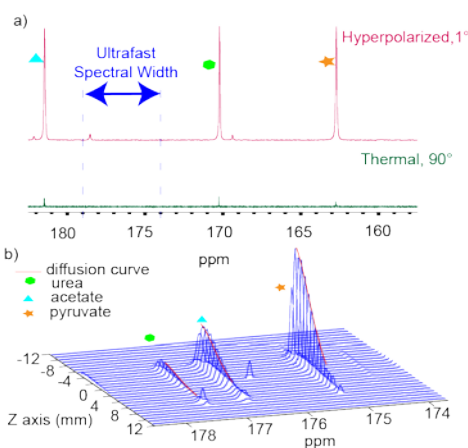


Figure 4: a) 1D ^{13}C spectrum. b) SPEN DOSY data.

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DiffERential EPitope Mapping- (DEEP) STD NMR to Reveal the Pharmacophore of a Protein Target

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Saturation Transfer Difference (STD) NMR spectroscopy is extensively used to obtain epitope maps of ligands binding to protein receptors under fast exchange conditions [1], [2]. STD NMR reveals structural details of biomolecular recognition processes, which are fundamental to direct lead optimisation efforts in drug discovery.

Standard procedures seek uniform saturation of the receptor to identify regions of the ligand contacting the protein binding pocket. However, in this way, the experiment does not provide information about the “nature” of the amino acids surrounding the ligand in the bound state.

Here we report a novel protocol (*DiffERential EPitope Mapping*-STD NMR or DEEP-STD NMR) to identify the type of protein residues contacting the ligand [3]. We demonstrate that the approach constitutes a novel versatile method to *orthogonally* explore the nature (aliphatic, aromatic, polar or hydrophobic) of the amino acid residues lining the surface of the binding pocket and their orientation relative to the ligand.

As a proof of principle, we selected two relevant protein-ligand interactions from different areas of interest: i) the interaction of 3-nitrophenyl- α -galactoside (3NPG) with subunit B of Cholera Toxin (CTB) [4], well known to CTB inhibitor designers; and ii) the interaction of 2,7 anhydro Neu5Ac with the glycosyl hydrolase GH33 from *Ruminococcus Gnavus* [5], of great interest in gut microbiota fundamental research, as it is over-represented in individuals affected by Inflammatory Bowel Disease [6]. For both systems, high resolution X-Ray structures were available, allowing us to validate the protocol. The approach overall seems solid, versatile and promising for expanding the view of STD-NMR and going beyond its current limitations.

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Scalable Chiral Alignment in Supramolecular Lyotropic Liquid Crystalline Phases

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Benzene-1,3,5-tricarboxamides (BTAs) are known to self-assemble into rod like and helical supramolecules.[1] These stiff aggregates act as mesogenes to form thermotropic or lyotropic liquid crystals (LLC).[2, 3] Achiral substituted BTAs aggregate to racemic mixtures of (M)- and (P)-helices. One handed helices can be obtained by introducing enantiopure sidechains to BTAs or via a sergeant-and-soldier-principle (SaS) by mixing chiral and achiral substituted BTAs.[4] In this work several BTAs were synthesized and tested for their behavior as SaS-LLCs.

These are used as weak orienting media, which give access to residual dipolar couplings (RDCs). RDCs offer complementary structure information to the conventional NMR parameters for structure determination (as 3J -couplings and NOEs). For structure determination of small organic molecules the LLCs should be compatible with organic solvents and introduce only a low degree of order.[5] Chiral alignment media can transfer the chiral information to the analyte molecule. As a result of diastereomorphous interactions with chiral alignment media enantiomers can be differentiated.[6]

Until now, primarily polymer based LLCs and anisotropically swollen gels are utilized as alignment media in organic solvents.[7] BTAs are a promising new approach for organic solvent compatible alignment media as synthetic steps for polymerisation[8] and/or crosslinking[9] are redundant. Challenges linked to this steps as well as time consuming sample preparation[7,9] of polymer gels can be avoided.

We show here, that mixtures of chiral and achiral BTAs form LLCs in organic NMR-solvents. To investigate the capabilities of the SaS-LLCs as chiral and enantiodifferentiating alignment-media, the enantiomers of β -pinene were analysed. The characterization of THF- d_8 as molecular probe through 2H -NMR-spectroscopy indicated that SaS-LLCs act as chiral alignment media for small organic molecules with the potential of scalable chiral characteristics.

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Sparse Sampling in Non-Frequency Dimensions

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Serial NMR experiments are often used in chemical and physical analysis. Experiments in a series can differ in temperature (monitoring phase transitions), pulsed field gradient setting (diffusion-ordered spectroscopy) or can be performed at different moments of time (monitoring chemical reactions). Multidimensional spectroscopy, although very informative, is difficult to employ in such experiments. The measurement of multidimensional spectra is time-consuming, because of costly sampling of indirect time dimensions. The non-uniform sampling (NUS) can help to shorten the experiment, but so far was limited to frequency dimensions. In this study we show how to extend it to non-frequency (pseudo)dimensions: temperature, diffusion-encoding gradients and others. A variety of different processing techniques will be discussed: time-resolved NUS[1,2], sparse inverse Laplace transform[3,4] and Radon transform[5,6].

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Novel Developments in Pure Shift NMR Using Perfect-Echo and Zangger-Sterk

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The interpretation of proton spectra of molecules with increasing complexity is often hampered by signal overlap due to homonuclear scalar coupling. Recently, the application of pure shift techniques successfully enhanced the resolution in one- and multidimensional proton detected experiments by collapsing multiplet structures into singlets.

A major drawback of methods for broadband homonuclear decoupling is their inherent sensitivity loss. In Zangger-Sterk[1] based techniques this is caused by a decrease in sample volume contributing to detectable signal. Various approaches have been presented to reduce this sensitivity penalty, such as *multi-slice-excitation*, *nemo-Zangger-Sterk*, *sequential slice selection*, ASAP or PSYCHE.[2-6]

Here we present a modification to Zangger-Sterk decoupling, which yields homonuclear decoupling for coupled pairs of spins even if they share the same volume element. This in turn can drastically improve the sensitivity of the experiment, as larger volume elements can be used for signal detection. These features are achieved by combination of Zangger-Sterk decoupling with the Perfect-Echo[7] experiment. Even in the presence of strong coupling the Perfect-Echo-Zangger-Sterk experiment produce clean spectra with good tolerance towards the occurrence of unwanted artefacts.

We show that this experiment is particularly suitable to study samples with strong signal clustering, a situation which can render classic Zangger-Sterk decoupling inefficient. Applications in one- and two-dimensional experiments are presented for oligomeric structures, which feature small chemical shift differences for atomic positions in the repetitive structural motives.

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Internal Reference Method for Natural Abundance ^{13}C Isotopomics

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Isotope ratio monitoring by NMR spectrometry (irm-NMR) provides the complete ^2H or ^{13}C intramolecular position-specific composition at natural abundance, a strategy belonging to the recent concept of **isotopomics** [1]. Irm- ^2H has been developed in the 1980s [2], but the development of irm- ^{13}C NMR is much more recent. Indeed, this analytical tool requires an accuracy of 1‰ (against 1% for irm- ^2H NMR) due to the very small deviation range of ^{13}C natural abundance values (only 50‰ against 500‰ for ^2H). Numerous methodological developments have been necessary such as ^1H decoupling using adiabatic pulses to reach such accuracy by ^{13}C NMR [3].

Currently, measuring position-specific natural abundance ^{13}C values relies on the combination of ^{13}C acquisitions with isotope ratio mass spectrometry (IRMS) as a reference. This approach has two drawbacks: i) it requires two different analytical techniques on the same sample, ii) it suffers from the low sensitivity of ^{13}C . To circumvent this constraint, we propose a new methodology to perform **^{13}C isotopomics analysis on a single sample, via NMR only**. In a first step, the concentration ratio of the chemical reference and the analyte must be determined by quantitative ^1H NMR with a 1‰ accuracy.⁽¹⁾ Then, a ^{13}C NMR analysis is performed on the same sample to determine the site-specific isotopic composition x_i . The latter can either be performed via direct ^{13}C detection or by polarization transfer methods. In both cases, the analyzed sample has to be highly concentrated in order to reach the required Signal-to-Noise Ratio in a realistic time in ^{13}C NMR. Unfortunately, this is not compatible with the first step, since the analysis of highly concentrated samples by ^1H NMR induces signal intensity and line shape distortions due to radiation damping (RD)⁽⁴⁾ phenomenon which severely hampers the accuracy of quantitative ^1H NMR.

In this context, we developed a new pulse sequence named DWET (Double-WET) to suppress RD and perform quantitative ^1H NMR analysis on concentrated samples with a 1‰ accuracy. DWET detects only a slice from the sample by saturating the signal arising from lateral areas of the sensitive volume. This method has been successfully applied to isotopic analysis through the discrimination of vanillin samples as a function of their geographical origins based on the ^{13}C isotopic profiles.⁽⁵⁾ However, it appears that the DWET pulse sequence is unsuited for samples with short T_1 , which forms a serious limitation for irm- ^{13}C NMR experiments where a paramagnetic agent is added to decrease T_1 . In this context, we developed two variants of the DWET method, called Multi-WET (MWET) and Profiled-WET (PWET), to reach the same accuracy of 1‰ with a better immunity towards T_1 variations. This new analytical package for ^{13}C isotopomics at natural abundance opens a wide range of new applications in the fields of authentication and metabolism.

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New Diagnostic Method for Chagas Disease Using $^1\text{H-NMR}$

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Chagas disease (CD) is caused by the protozoan parasite *Trypanosoma cruzi* [1]. This disease represents an important cause of morbidity and mortality in endemic countries, affects approximately 8 million people worldwide [2,3]. In Colombia, *T. cruzi* infection prevalence is around 5%, corresponding to 700,000 people, and in some areas of the department of Santander the seroprevalence is about 50% [4]. CD include an acute and a chronic phase, which presents a wide spectrum of clinical manifestations including cardiac, digestive and neurological forms [2,3]. The diagnosis of the infection with *T. cruzi* is made through serological and parasitological tests, which are used according to the clinical phase. The diagnosis is complex especially during the chronic phase, due to the lack of symptoms and the low or intermittent parasitemia that leads to direct parasitological methods having a low sensitivity. For this reason, the diagnosis is based on serological methods which detect the presence of specific antibodies directed against antigens of *T. cruzi* combined with clinical and epidemiological findings. However, serological tests present high sensitivity but lack specificity because of antigenic cross-reactivity with other parasites like *Leishmania sp.* and *T. rangeli* [5–7]. In this regard, World Health Organization suggested that at least two assays based on different techniques may be used in parallel to increase diagnostic accuracy because a single assay is not considered sufficiently sensitive and specific. Less invasive methods for diagnosis, such as determination of biomarkers from blood serum would be of significant advantage and useful for primary diagnosis, early detection and surveillance. Furthermore, non-invasive biomarkers could be used for knowledge of the metabolic alterations and it will help to understand the course of the infection. The aim of this work was to study the potential of NMR spectroscopy for the application of metabolomic studies in patients with CD to determine new metabolomic biomarkers of disease diagnostics.

In this work we use liquids state NMR spectroscopy to try to establish a diagnosis method to patients of CD. The NMR experiments were performed on a Bruker Avance at 9.4T, 400 MHz for proton resonance. One-dimensional proton spectra were measured using a CPMG sequence [PRESET-90°-(δ -180°)_n-aq], which was optimized to remove the short T2 components arising due to the presence of high molecular weight proteins as well as to obtain a smooth baseline for multivariate analysis. All NMR spectra were binned to buckets including an equal width of 0.04 ppm. Water signal (4.6-4.8 ppm) was presaturated using low power level of 0.2 mW.

Chemical shift of 4.6 to 5.0 ppm was eliminated to remove water residual. The Principal component analysis (PCA) and partial least-squares discriminant analysis (PLS-DA) was implemented to appraise grouping trends. For this purpose, groups of healthy volunteers (control group, CG) and patients with CD were compared. The potential biomarkers were found using the PLS-DA model. The results showed the metabolites important for the discrimination of the groups CG and CD. The predictive ability and validation of the PLS-DA model were determined using the predictive set of samples. According to the ROC curve

and the AUC values, the PLS-DA model showed excellent discriminant properties for comparing CG and CD. This metabolites can be used like biomarkers of disease diagnostic.

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REST and Multivariate NMR Methods in Beer Analysis

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Diffusion-ordered spectroscopy (DOSY) [1] is one of the most commonly used NMR experiments in the analysis of mixtures. However DOSY is of limited use where mixture components have similar diffusion coefficients (i.e. similar hydrodynamic radii). Moreover, signal overlap can obscure diffusion information and complicate spectral interpretation, although this problem can be alleviated by multivariate analysis [2] if the compounds of interest have sufficiently different diffusion coefficients. When even this fails, differences in relaxation between different compounds can be exploited to separate signals if the combination of selective excitation and isotropic mixing is used to ensure that all the signals measured for a given species originate from a single proton. [3] Exploiting relaxation differences and focusing on selected spin connectivities can give great insight in mixture analysis, in a new class of experiments named *Relaxation-Encoded Selective TOCSY* (REST). The new method can for example use T_1 encoding, e.g. by inversion or saturation recovery (REST₁), or T_2 , e.g. using PROJECT [4] (REST₂). REST has been used to investigate the carbohydrate content of a lager beer sample. The carbohydrate region of the ¹H spectrum is very crowded and impossible to interpret directly, as is often the case for sugar mixtures. Using the REST₂ experiment, followed by multivariate analysis with OUTSCORE, [5] spectra of individual components can be extracted.

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Molecular Spins and Persistent Magnetic Moments

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Some time ago [1,2], the author has proposed the theoretical concept of persistent currents in the electron shells of some diamagnetic molecules. If confirmed, these should be connected with the existence of permanent magnetic moments even in the absence of unpaired electrons, and of persistent molecular spins (angular momenta). A classical analogy for such quantum molecular situations might be, for example, the motorbike clutch: engaging in one sense, and free-running in the other.

From the quantum mechanical point of view, this appears unavoidable, in particular in small molecules which lack cyclic symmetry, such as chloraldehyde or, in general $C(XYZ)$, where C is a central atom and XYZ are three different atoms/groups arranged around it. A bonding electron partially localized near the central atom C , but affected by all three substituents X , Y , and Z , will necessarily behave as a wave moving differently in the cyclic sense (XYZ) rather than in the anti-cyclic sense (XZY). The result is a cyclically oriented ground state and a permanent current loop. In turn, the latter must give rise to a persistent magnetic moment and a persistent angular momentum (spin).

Such persistent current loops are expected to exist also in a-cyclic fragments of large molecules, including proteins. In very small a-cyclic molecules, what makes them particularly interesting in the magnetic resonance context, is the fact that they would make the whole molecule behave as a unique spin particle, endowed with a magnetic moment, and therefore capable of exhibiting magnetic resonance at some (probably very low) Larmor frequency. If so, very-high-field magnetic resonance techniques could be used to study these phenomena. Another interesting aspect is that persistent electric currents in molecular electron shells are not an explicit part of the popular density function theory (though the DFT approximation might in part implicitly account for them). Making persistent electron current density, apart from electron charge density, an explicit feature of DFT might be beneficial for its accuracy.

In this presentation, the author would like to complement the original hypothesis with several quantitative estimates, as well as to point out the similarities and differences (considerable) between the hypothetical molecular spins and the spin-particles-as-we-know-them.

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Rethinking NMR Crystallography

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The physicochemical properties of crystalline solids are determined to a great extent by the atomic-level structural properties of the material of interest. This is especially important for pharmaceutically active materials, where bioavailability depends on crystal structure. This is also the case in many other areas, where exact knowledge of crystal structures is required, and as a result, the development of new and improved strategies for crystal structure determination will have a significant impact.

Although single-crystal X-ray diffraction (SC-XRD) is the most powerful and routine technique for crystal structure determination, it is limited to single-crystals of appropriate quality and size. In the many cases, where the sample is either to some degree amorphous or a microcrystalline powder, structural characterization by SC-XRD is potentially not possible. Solid-state nuclear magnetic resonance (NMR) can overcome this limitation through its sensitivity to the local atomic environment regardless of the degree of long range order.

Over the past decade the combination of NMR and computational methods has made tremendous progress [1] and today there are many examples of structure determination or validation by chemical shift measurements combined with density functional theory calculations (DFT)[2]. Recently there have been examples of *de novo* structure determination, combining NMR chemical shifts, DFT shift calculations and crystal structure prediction (CSP)[3]. However, there is currently a huge bottleneck in chemical shift based NMR crystallography, which is the time required for CSP calculations, which severely limits the complexity of the molecules that can be tackled.

In general, CSP methods have two stages. First, an ensemble of gas-phase conformer structures is generated and sorted by their calculated internal energy. In the second stage the gas-phase conformers with the lowest internal energy are selected, and for each conformer an ensemble of possible crystal structures is generated. This procedure is based on the assumption that the low-energy gas-phase conformers are similar to those in the crystalline structures. Both stages involve searching the potential energy surface (PES) of the system, where the number of minima on the PES increases exponentially with the number of atoms. Often, only fully quantum mechanical calculations suffice to deliver the required level of accuracy[4]. These combined factors are responsible for the high computational cost associated with the CSP calculations.

Here, we present a method that combines solid-state NMR measurements on powder samples with a computational analysis that allows the introduction of unbiased structural constraints restricting the area of interest of the PES that needs to be searched when using CSP in a structure determination process. These constraints are introduced during the gas-phase structure generation, where they can reduce the number of conformers by up to 90%, reducing the time required for the CSP procedure by an order of magnitude. This increase in efficiency allows the chemical shift based NMR crystal structure determination of structures of unprecedented complexity and size. We demonstrate the method on different pharmaceutically active

compounds using constraints derived from a series of (1H-13C) HETCOR experiments. In particular, we show that the restrained CSP method allows the determination of the crystal structure of ampicillin, whereas unrestrained CSP fails to produce the correct structure.

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NMR Metabolomics and Lyme Borreliosis

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Lyme borreliosis (LB), also known as Lyme disease, is the most important tick-transmitted disease globally [1, 2]. LB is caused by the spirochete *Borrelia burgdorferi* sensu lato. *Borrelia* is transferred to the human host via the tick's saliva when the tick attaches itself to the skin of the host.

Clinically the LB infection can roughly be classified into early stage and disseminated stage of the disease. The early diagnosis of LB is possible without any laboratory testing if the patient is aware of the tick bite and a typical early rash lesion is visible at the tick bite site. However, it has been estimated that only 50 % of the patients with LB develop visible rash [3]. Thus, in atypical cases, with no or only a faint rash, laboratory confirmation of the infection would be most helpful. Unfortunately, no such test is currently available for the early stage of the disease, as the antibody production in early phase of the infection is too low for detection. The mainstay of LB laboratory testing today is the detection of borrelia specific antibodies in patient serum and cerebrospinal fluid. Direct diagnostic testing methods, such as cell culture or nucleic acid amplification, suffer from low sensitivity, and are therefore not practical.

Our aim was to develop a novel method for the diagnostics of LB by utilizing nuclear magnetic resonance spectroscopy (NMR) and/or mass spectrometry (MS). Both NMR and MS are suitable for metabolomics based research, i.e. studying the different substances formed in metabolic processes inside the body. With these methods, our aim is to detect biomarkers that are specific for LB.

To test the suitability of NMR metabolomics for the diagnostics of LB, we performed a preliminary study with *Borrelia* infected mice. Both urine and blood samples were collected from the infected mice. The samples were measured with a 600 MHz NMR instrument and samples from non-infected mice were used as controls. The data was analyzed by using multivariate analysis. From this data, clear differences between the *Borrelia* infected and the control mice could be seen, for example in the concentrations of organic acids, glucose and some amino acids. However, none of these molecules could be used as a specific biomarker for LB, as all of the molecular differences can be regarded as general biomarkers for infection.

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Computer-Assisted 3D Structure Elucidation of Small Molecules Using Residual Dipolar Couplings and Isotropic ^{13}C Chemical Shifts

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The 2D structure (molecular constitution) of most small molecules can be in principle straightforwardly determined by manual or automatic analysis of a set of experimental data that includes the molecular formula, a series of 1D and 2D NMR experiments providing through-bond connectivity (COSY, TOCSY, HSQC, HMBC and ADEQUATE/INADEQUATE), and empirical chemical shift predictions. This is the main concept embedded in CASE (Computer Assisted Structure Elucidation) programs. Once the 2D structure is available, the determination of the relative spatial arrangement (configuration and preferred conformation) of all atoms in the molecule is a more challenging task commonly addressed by using NOE and 3J coupling constants analysis, as well as recent developments on DFT calculations of ^{13}C chemical shifts (DP4). The development of the application of Residual Dipolar Couplings (RDCs) to the configurational and conformational analysis of small molecules has matured enough in the recent years to perform this task in an almost straightforward way, without even the need of using NOE and 3J coupling analysis, for the analysis of rigid and semi-rigid small molecules. We have recently shown that it is possible to go from molecular constitution to configuration without human intervention by feeding the program with an SDF structure file from a CASE-based program and a table containing only one-bond proton-carbon RDCs.[1] The process involves a) generation of the configurational space, b) conformational analysis for each configuration, c) automatic superposition of conformers for single-tensor analysis, d) fitting of RDC data to a set of conformation/configurations using model selection to prevent overfitting, e) plot of a bar diagram with quality factors indicating the selection of the correct 3D structure. This work led to the creation of StereoFitter, a complete computational module that can perform multiple-nmr parameters fitting (scalar couplings, NOE-derived distances, chemical shifts, RDCs, RCSAs) to combinations of configuration/conformations using model selection (Akaike Information Criterion (AIC)) to prevent overfitting. StereoFitter can be trivially extended to the use of chiroptics parameters (ECD, VCD) to determine absolute configuration. Generation of configurations, conformational analysis, DFT geometry optimizations, and analysis of DFT GIAO calculations of NMR parameters (J couplings, Chemical Shift Tensors) is also performed by StereoFitter by interacting with third party computational chemistry packages. In the present work we would like to show that, after the determination of the molecular constitution by the CASE program, a combined use of RDCs and isotropic ^{13}C chemical shifts is enough to determine configuration, particularly useful for proton-deficient molecules. For the antimalarial drug Artemisinin,[2] the correct structure was determined using only ^{13}C chemical shifts. The whole process from 2D structure to the generation of 38 diastereomers, followed by DFT energy minimization and DFT GIAO calculation of CSA tensor for each diastereomer, and finalizing with the fitting of chemical shifts to the set of diastereomers, took only 2 hours in a 5 years old PC equipped with 2 Xeon Processor with 6 cores each. Results shown in Figure 1.

We have also applied the method to other relevant natural products such as the sesquiterpene lactone 10-epi-8-deoxycumambrin B [3] and the fungal metabolite Homodimericin A.[4] The structure of this natural product was determined using a combination of CASE, RDCs and RCSAs, since it has a hexacyclic core

with fourteen quaternary carbons, eleven of them contiguous. For this particular compound we used the data reported by the authors.[4] For Homodimericin A we observed that it is possible to use ^{13}C chemical shifts as an alternative to RCSAs to determine its configuration.

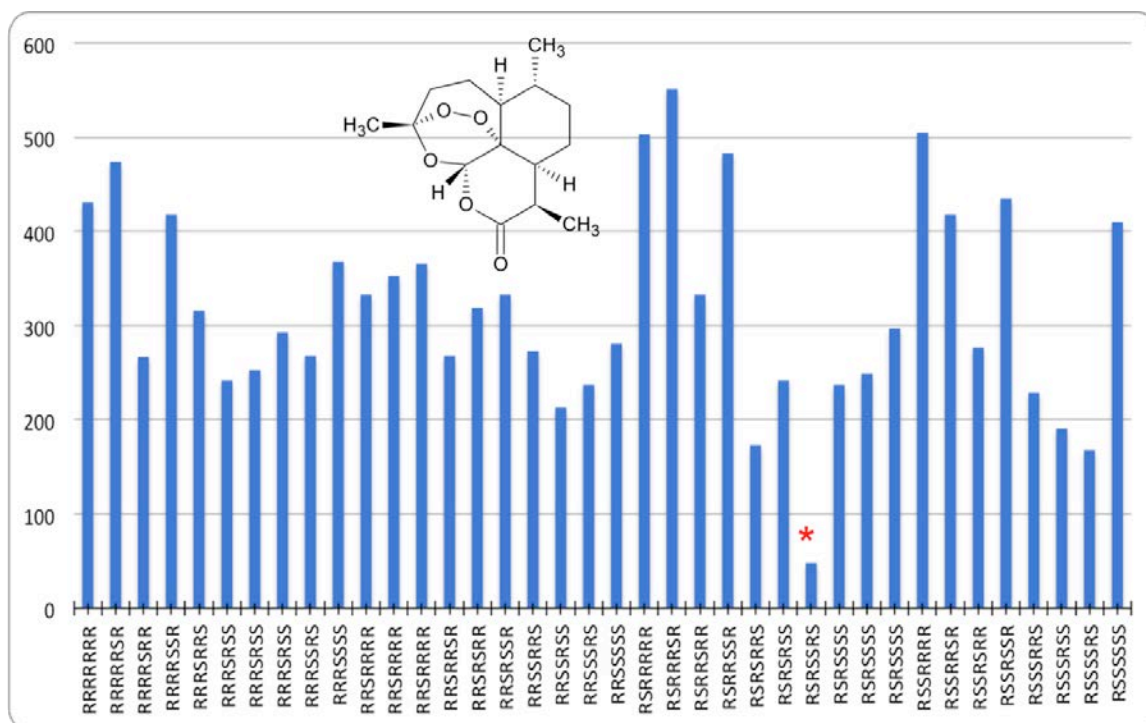


Figure 1. Determination of the configuration of Artemisinin using only isotropic ^{13}C chemical shift. Fitting DFT calculated chemical shift with the experimental data using StereoFitter. The red asterisk shows the correct structure with the lowest χ^2 value.

In contrast to methodologies such as DP4 our method allows the fitting of populations to all the available experimental data including chemical shifts. Potential overfitting problems are treated by using the Akaike Information Criterion (AIC) where models with increasing number of populations are penalized.

CASE analysis was performed using the MestReNova Structure Elucidator. Most of the results presented here used Poly(methylmethacrylate) (PMMA) based flexible gels, whose degree of alignment can be easily tuned by variable and reversible compression.[5]

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In situ illuminated Nanolitre NMR spectroscopy

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In-situ NMR detection of UV-vis assisted reactions find application in many different fields as photocatalysis, photochromics, photoisomerizations, and hiperpolarization techniques, among others [1]. However, commercially-available NMR setups for in-situ illumination suffer from many limitations mainly related to the difficulty of an efficient and uniform illumination of the entire sample volume. These setups deal with hundreds of microliters of sample typically contained in a 5 mm-NMR tube, therefore requiring high power light sources and complicated setups to bring the light uniformly along the sample tube. The high intensity laser brings additional problems related to light-assisted sample degradation. Essentially two different methods of guiding the light into the coil region of a superconducting magnet have been developed over the years, light from the bottom of the NMR probe or from the top. The latter option is the most used nowadays, although still encounters several conflicts as the magnetic field distortions caused by an insert into the sample volume and the light absorption by the solution above the radiofrequency coil region, which could decrease the stability of the sample and the observed signal, together with a loss of some sensitivity and resolution [2].

Here we present a novel setup for the illumination of nanolitre samples with low power light sources for the in situ NMR analysis of UV-vis light-assisted chemical reactions. The nanolitre sample are located underneath a reduced-diameter NMR coil, called microcoil, which is integrated on top of a microreactor allowing the light penetration typical of microchannels [3]. The use of these small NMR detectors not only offer an interesting approach to enhance NMR sensibility [4] but also, the capabilities of the microcoils of analyzing sample volumes in the nL ranges enable the rapid optimization of reaction conditions and an extremely fast determination of the kinetic parameters of reactions [5, 6].

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26

Round Robin Test for Applying the qNMR Method to the Japanese Industrial Standards

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Introduction

Verification of the method for quantitative analysis using ^1H NMR (^1H quantitative Nuclear Magnetic Resonance: ^1H qNMR) recently began. By setting optimised measurement conditions for quantification, it will be possible to determine precise quantitative values.[1,2,3,4,5] ^1H qNMR is a method for determining the quantities of substances based on the amount of proton nuclei. Therefore, it is used for deciding the accurate purities of reagents, residual pesticides, natural products, food additives, standard products, etc., and the absolute quantification of compounds that lack standards. In addition, we prepared a sample solution to which was accurately added an SI (International System of Units)-traceable Certified Reference Material (CRM), which was developed for ^1H qNMR as calibration standard. When this sample solution is measured using the ^1H qNMR conditions, accurate SI traceable quantitative values can be obtained. This method has been reported as the Accurate Quantitative NMR with Internal reference substance (AQARI) method.[1,7,8] ^1H qNMR in which this AQARI methodology is applied is already being adopted in Japan's official analytical methods such as the Japanese Pharmacopoeia and Japan's Specifications and Standards for Food Additives as an absolute purity determination method.[9] ^1H qNMR is a novel assay method with principles that differ from those of conventional chromatography. Rather than identifying and quantifying all impurities contained in a product as in the mass balance method, it is a method whereby accurate SI traceable quantitative values can be obtained effectively. Therefore, it is expected to be applied not only in the field of pharmaceuticals and food products but also in various fields requiring precise quantitative values such as the industrial chemical field. Based on such a background, we performed a Round Robin Test using the same substance to be measured and calibration standard for qNMR, and assessed the accuracy of this assay method aiming at the inclusion of ^1H qNMR in the Japanese Industrial Standards.

■Methods

1. Participating facilities

The study was performed at five facilities, namely, the National Institute of Health Sciences, National Institute of Advanced Industrial Science and Technology, the National Research Institute of Fisheries Science, JEOL RESONANCE Co., Ltd, and Wako Pure Chemical Industries, Ltd.

2. Materials

SI-traceable CRM, 1,4-bis(trimethylsilyl) benzene- d_4 (1,4-BTMSB- d_4 , Wako Pure Chemical Industries, Ltd.; certified value: mass fraction $99.9 \pm 0.6\%$) was used as the substance to be measured, and 3,5-bis(trifluoromethyl) benzoic acid (3,5-BTFMBA, the National Institute of Advanced Industrial Science and Technology; certified value: mass fraction $99.96 \pm 0.06\%$) as the calibration standard for qNMR. Methanol- d_4 (Wako Pure Chemical Industries, Ltd. (Purity (GC) mass fraction $\geq 98.5\%$ and deuterium content of $\geq 99.8\%$) was the deuterated solvent used for dissolving the substance to be measured and the calibration standard for qNMR.

3. Preparation of sample solutions

In preparing the sample solution, the mass of the substance to be measured and the calibration standard for qNMR were weighed such that at least twice the minimum weight W_{\min} [10] and the signal-to-noise ratio of the signals of the substance to be measured and the calibration standard for qNMR in the NMR spectrum were approximately equal. Methanol- d_4 was added so that the substance to be measured was ~0.1 w/v%. The minimum weight W_{\min} of the precision balance was determined separately using formula 1.

$$W_{\min} = \sigma \times 2000$$

Formula 1: The minimum weight W_{\min} was evaluated using aluminium pan (approx. 60 mg) which is used for the tare of the substance to be measured and the calibration standard for qNMR. The minimum weight W_{\min} was calculated by multiplying σ by 2000. σ : the standard deviation when the measurement of the weight of the aluminium pan was repeated 10 times 2000: the value when the safety coefficient (2) was divided by the permitted range (0.10%) in the measurement (= $2/(0.1/100)$)

4. NMR instrument, measurement conditions, and data process conditions

Six machines with proton (^1H) resonance frequency ≥ 300 MHz at each facility were used in the NMR instrument. The ^1H qNMR measurement conditions were the standard measurement conditions that were set based on the Japanese Pharmacopoeia conditions. In short, the nucleus to be measured was ^1H , digital resolution ≤ 0.25 Hz, spectral width ≥ 20 ppm including -5 to 15 ppm, spinning Off, flip angle of 90° , ^{13}C decoupling, relaxation delay ≥ 60 s, number of transients ≥ 8 , dummy scans ≥ 2 , and probe temperature maintained at 20-30 °C. The ^1H qNMR measurement conditions specific to each facility were set based on these standard measurement conditions to perform the measurements. The ^1H qNMR data process conditions were those conditions suitable for measuring the respective quantitative values set independently using the software of each facility.

5. System Suitability Test

A system suitability test was set up based on the documentations on the specification established in the purity determination for ^1H qNMR in the Japanese Pharmacopoeia.[11]

6. Evaluation methods

We checked whether the purity obtained for each facility was within the range of uncertainty of the substance to be measured (1,4-BTMSB- d_4 ; certified value: mass fraction $99.9 \pm 0.6\%$).

■ Results

The ^1H qNMR measurement conditions were set based on the procedure documented in the Japanese Pharmacopoeia. 3,5-bis(trifluoromethyl) benzoic acid (3,5-BTFMBA; certified value: mass fraction $99.96 \pm 0.06\%$) was used as the calibration standard for qNMR, and the system for determining the purity of 1,4-bis(trimethylsilyl) benzene- d_4 (1,4-BTMSB- d_4 ; certified value: mass fraction $99.9 \pm 0.6\%$) was used for the evaluation. As ^1H qNMR includes bias and variation factors in the respective operations for sample preparation, NMR measurement and data process, appropriate conditions were set for them. All analytical results lied within the range of uncertainty of 1,4-BTMSB- d_4 , confirming that ^1H qNMR functions as a method for determining the purity or content rate of general organic compounds.

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Time-Resolved NUS Interleaved Experiments with Online Processing and Analysis

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The present work combines three key ideas: 1) implementing time-resolved non-uniform sampling (TR NUS) for monitoring changes in non-stable samples, 2) interleaving NMR experiments within the framework of TR NUS procedure (enables one to acquire various types of spectra simultaneously), 3) processing and analysing spectra on the fly.

TR NUS [1, 2] is an approach to modify multidimensional NMR experiments for monitoring time-dependent processes occurring on the time-scale of tens of minutes to hours. As in conventional NUS, some random points of the indirect dimensions are probed, while others are skipped. However, since the sample and its spectrum are changing during the experiment, it is justified to use a very long, shuffled sampling schedule (even many times larger than the size of a full grid) and divide it into subsets that are “snapshots” of the process. Allowing for an overlap between these subsets, proper time resolution can be achieved. Each subset is then reconstructed with compressed sensing (CS) algorithms.

The results of an N-dimensional experiment can then be presented as a N+1-dimensional array, where the actual, physical time serves as an extra quasi-dimension. Also, these results can be viewed a “movie” of N-dimensional spectra with peaks slightly changing from one frame to another, thus representing the changes in the sample.

However, in the case of uncontrolled, irreversible changes in the sample the chosen measurement technique may turn out to be not optimal. Then, both sample and measurement time get wasted. Therefore, it would be best to conduct several experiments at the same time: for instance, record one-dimensional spectrum of high sensitivity between increments of well resolved but insensitive multidimensional spectrum; or interleave signal sampling by several different multidimensional techniques, as was shown for NMR methods aimed at the resonance assignment of proteins [3].

To improve the procedure even further and make it convenient for the user, we have created a system which automatically carries out the CS reconstruction of the NUS spectra simultaneously to their acquisition. This system starts the reconstruction for every frame when the necessary number of indirect dimension FID points is acquired; during this time the data for the next frame are being acquired, etc. It also processes the data and shows the spectra with a possibility to switch between frames. The work on peak-picking and peak-tracking simultaneous to acquisition is in progress, as well as the generalisation of the system to make it capable of treating interleaved experiments.

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28 Chromatographic NMR Spectroscopy with Hollow Silica Spheres

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NMR diffusometry comprises an ensemble of techniques designed to separate the signals of different molecules according to their apparent diffusion coefficients in solution. In some cases, the combination of dedicated pulse sequences and data processing can produce 2D “DOSY” maps displaying the chemical shift in the direct dimension and the apparent diffusion coefficient in the indirect dimension, something that closely reminds of a TLC plate. However useful in the analysis of complex mixtures, the success of this technique depends both on the absence of signal overlap, and on the existence of sizeable differences among the diffusion coefficients of the various species, a condition hardly met in crowded ^1H NMR spectra.

While signal overlap can be mitigated by pureshift spectroscopy, the separation in the diffusion dimension can be significantly improved by perturbing the analytes’ diffusion coefficient with an interacting matrix. To this aim, the use of a stationary phase made of solid silica has been proposed in order to exploit the partition equilibria which occur in a chromatographic column. [1] Owing to the intrinsic sample inhomogeneity, however, the practical implementation of this method, dubbed *chromatographic NMR*, relies on magic-angle spinning to remove the anisotropy of magnetic susceptibility across the sample.

In this context we have recently proposed the use of hollow microspheres to reduce the impact of field inhomogeneities. [2] The idea is founded on the observation that, in the presence of a uniform magnetic field, the effect of an isolated sphere on the external region is equivalent to that of a dipole located at the center of the sphere itself. When an NMR tube filled with silica powder and a solvent is put into the magnet, the magnetic field becomes inhomogeneous within small domains across the sample, a situation which is beyond the correction capabilities of the spectrometer shim system. However, if the same spheres are partly emptied to form a cavity, it can be proved that the magnitude of the associated dipolar field collapses as the shell of the sphere becomes thinner.

In this communication we describe the use of hollow silica microspheres in chromatographic NMR, from early experimental observations up to a numerical solution of the magnetostatic problem in model samples. The obtained results provide a significant understanding of how the performances of such systems depend on the internal sample geometry, together with the surface properties of silica. Indeed, the possibilities offered by hollow spheres are manifold: we expect that new materials can be envisaged to simultaneously exploit different supramolecular recognition abilities based, for example, on the concurrence of hydrophobic and dipolar interactions. Along this line, the properties of hybrid hollow microspheres are currently being investigated in our lab.

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Combining the best of NMR: Fast, Quantitative, Highly Resolved and Sensitive approach for metabolomics

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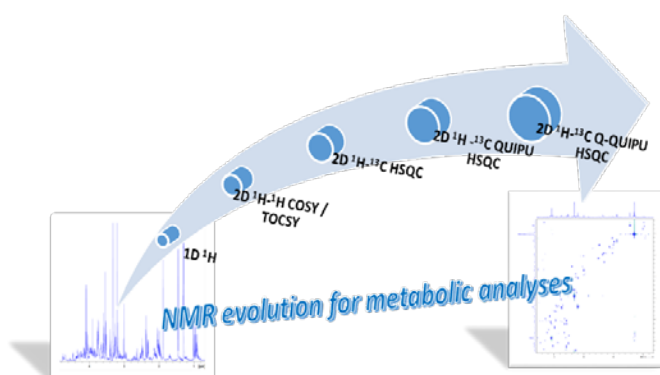
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A better understanding of living organisms requires a detailed and quantitative analysis of complex biological samples such as extracts, biofluids for organisms under investigation. Among the most efficient analytical techniques, NMR is one of the most used and makes it possible to quantify complex mixtures with an excellent repeatability and precision.

However, NMR is not the most sensitive technique despite its ability to give informative structural and quantitative data.

In the context of metabolic studies, complex mixtures are usually hard to decipher on a 1D spectrum (see Figure). Therefore, 2D NMR is a nice alternative for improving the spectral resolution along two dimensions. For highly overcrowded spectra, the ultimate solution is to record heteronuclear ^1H - ^{13}C 2D maps to benefit from the larger spectral window of ^{13}C . However ^{13}C nuclei are low abundant (1.1%), preventing the detection of low concentrated metabolites. Pushing the inherent limits of NMR is essential for investigating potential low concentrated metabolites within complex biological mixtures.

We suggested a new approach based on new quantitative ^1H - ^{13}C techniques called QUIPU HSQC [1] that can detect metabolites in the range of 100 μM within leaf extracts thanks to the sensitivity gains of 2 to 3 for ^1H - ^{13}C correlations in comparison with the conventional version. This technique has also been applied with success to better understand the photosynthetic and photorespiratory cycles by detecting sugar phosphates as biomarkers of these processes [2]. This approach has recently been improved through the Quick QUIPU HSQC or Q-QUIPU HSQC [3] combining spectral aliasing [4], NUS [5] and variable repetition times [6] methods. The time necessary to obtain quantitative data could be divided by a factor from 5 to 9 while preserving highly resolved spectra. The analytical performance of this new advanced tool dedicated to metabolomics is successfully shown on a model metabolite mixture and on breast cells cancer extracts. The new method opens perspectives in a variety of fields related to metabolomics.



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30

NMR Experiments for Several Receivers

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NMR experiments involving multiple receivers provide a unique way of increasing the sensitivity and information content of data recorded in a given period of time [1-3]. We present a comprehensive series of such experiments designed for simultaneous detection of abundant nuclei, such as ¹H, ¹⁹F and ³¹P, as well as samples enriched with magnetically active isotopes including ¹³C and ¹⁵N. The multiple receiver experiments are categorized into three main types – (a) parallel acquisition, (b) sequential acquisition and (c) interleaved experiments. The optimum implementation is shown to depend on the relaxation properties of the involved nuclei as well as the intrinsic sensitivity of the directly observed nuclei. We particularly focus on the basic NMR experiments involving the pairs of ¹H / ¹⁹F nuclei and ¹H / ³¹P nuclei not least because of the particularly important role that ¹⁹F and ³¹P NMR plays in drug discovery and pharmaceutical industry [3]. Essentially any of the basic 2D NMR experiments, such as COSY, NOESY, TOCSY, DOSY, HSQC, HMQC, HMBC, HETCOR or relaxation measurements that are routinely used in small molecule NMR can be easily adapted for and more efficiently recorded on systems equipped with multiple receivers.

Many of these experiments are amenable to further reduction of experiment time by combining them with other fast NMR techniques, such as Hadamard NMR, non-uniform sampling, spatial encoding or rapid pulsing methods. We believe that the multi-receiver technology will boost the development of new NMR experiments as well as NMR research in general, making the NMR instruments more efficient and making the NMR spectroscopy even more unique in the universe of analytical tools and experimental techniques.

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Structure-Property-Relationship for Amine Based CO₂ Absorption by Quantitative NMR Spectroscopy

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In a cooperation project between Evonik Industries, thyssenkrupp, and the Laboratory of Engineering Thermodynamics (LTD) of University of Kaiserslautern, novel amines for syngas purification by reactive absorption are developed and investigated [1]. All studied amines are derivatives of triacetone amine (TAA) and will be named “Evonik Amines” (EvAs) in this project. This interesting group of substances has not been considered before for the present application. In a screening, a large number of EvAs are synthesized, i.e. the chemical structure is tailored. Thermodynamic properties of aqueous solutions of the novel amines, both loaded with CO₂ and unloaded, which are particularly relevant for the industrial reactive absorption process are investigated: gas solubility and absorption kinetics, solid solubility, and liquid-liquid equilibria. The results show that even for very small differences in the chemical structure of the EvAs, major differences in their absorption behavior can be observed.

To gain a better understanding of these findings, aqueous solutions of several EvAs, loaded with CO₂ under high pressure, are studied with NMR spectroscopy. The experiments are carried out in a wide range of CO₂ loadings and temperatures, which are relevant for industrial applications. For this purpose, a robust method for the in situ analysis of the chemical equilibria in CO₂ loaded solutions, using quantitative NMR spectroscopy, is developed. Depending on the molecular structure of the amines, different reaction products are formed via different reaction paths when CO₂ is being absorbed. Thus, the ability of NMR spectroscopy to combine directly structure elucidation with quantitative analysis is important in this project to gain insights into the reaction network and chemical equilibria and finally into the performance of the amines in a reactive absorption process.

By comparing the results obtained for the different TAA derivatives the influence of changes in the molecular structure on the studied properties is elucidated. That knowledge gives a guideline for the synthesis of new amines.

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Hot and Cool RDCs in One Alignment Medium

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In recent years it has become more and more popular to measure residual dipolar couplings (RDCs) providing global structural information as well as the relative configuration of molecules. For this approach suitable alignment media are necessary and as far as biomolecules are concerned, a wide range of water-based media are available. However, when it comes to small organic molecules, the compatibility of the analyte with the alignment-medium and the respective organic solvent limit the eligible media. Although, several alignment media based on either liquid crystalline phases or anisotropic swollen, stretched or compressed gels (SAG) have been successfully applied in the RDC approach [1], the development of further alignment media is desirable.

We synthesized Poly- β -phenethyl-L-aspartate (PPLA), the constitutional isomer of Poly- γ -benzyl-L-glutamate (PBLG), which is a commonly used alignment medium in organic structure elucidation [2]. We have investigated the orientational properties of PPLA and determined excellent enantiodifferentiating abilities; by today the highest observed in a homopolypeptide-based alignment medium. Enantiodifferentiation is based on different diastereomorphous interactions between a chiral analyte and the P- and M-helix configuration or between both enantiomers of the analyte and one helix configuration, both leading to different mean orientations of the analyte.

PPLA possesses the special temperature dependent feature to undergo a reversible helix screw-sense inversion in the LLC-state without perturbing the orientational order of the α -helical backbone in the nematic arrangement [3]. The transition originates from small energy differences between left- and right-handed helices in polyaspartates. This thermoresponsivity implies the possibility to measure different orientations of one enantiomer of the analyte within the same sample by just raising the temperature.

Herein we report the synthesis of Polyphenethylaspartate and its outstanding abilities as thermoresponsive enantiodifferentiating alignment medium in NMR-spectroscopy.

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Time-Course Metabolomic Analysis of Yeast Unrestricted Growth by 2D ^1H - ^{13}C HSQC NMR and 1D ^1H NMR Spectroscopy

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Metabolomics is a field of ‘omics’ research that is primarily focused on the identification and characterization of small molecule metabolites in cells, tissues, organs and organisms[1]. Metabolomic approaches have been used for biomarker discovery in clinical studies[2], food analysis[3], environmental assessment[4], and phenotyping[5], among others. Due to the important role that metabolomics plays in bridging the phenotype-genotype gap in all these areas, it has become one of the most ambitious ‘hot topic’ at present.

One of the preferred instrumental techniques in metabolomics is Nuclear Magnetic Resonance (NMR) spectroscopy, due to the inherent advantages of the technique (simple sample preparation, non-destructive, robust,...).

Most of the studies have been focused on the analysis of one-dimensional proton (1D ^1H) NMR, while the analysis of other nuclei (such as ^{13}C and ^{31}P) and other NMR experiments (^1H - ^{13}C HSQC[6], 2D ^1H INADEQUATE[7],...) are still underrepresented. The preference of 1D ^1H NMR in NMR metabolomics lies in the fact that has good sensitivity in a short acquisition time, albeit it lacks spectral resolution, since it presents a high overlapping degree.

In this study, we have characterized the metabolome of a yeast strain grown in two different liquid media for a 3-day period with both 1D ^1H NMR and 2D ^1H - ^{13}C HSQC experiments. Dozens of metabolites, including amino acids, nucleotides, sugars and organic acids, among others, have been detected and quantified. The metabolic response over time has been connected to the particular composition of each culture medium, proving that cell adaptation does not only occurs under unfavourable or stress conditions, but also in order to minimize the energy cost of growth and development (i.e. by uptaking a metabolite from the extracellular medium instead of producing it by *de-novo* biosynthesis).

Finally, with this empirical example, we have concluded that the combined analysis of both 1D ^1H NMR and 2D ^1H - ^{13}C HSQC datasets is much preferable than the analysis of the two separately, since a better interpretation of the overall metabolome can be obtained with the combined approach analyzing the two datasets simultaneously.

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Design and Validation of a Compact Online NMR Module

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Monitoring chemical reactions is the key to process control. Today, mainly optical online methods are applied, which are calibration intensive. NMR spectroscopy has a high potential for direct loop process control [1, 2] while cutting the calibration and validation needs to an minimum and thus exhibiting short set-up times. Compact NMR instruments make NMR spectroscopy accessible in industrial and harsh environments for advanced process monitoring and control.

Intensified continuous processes are in focus of current research. Flexible (modular) chemical plants can produce different products using the same equipment with short down-times between campaigns and quick introduction of new products to the market. In continuous flow processes online sensor data and tight closed-loop control of the product quality are mandatory. Data analysis techniques are available but currently mostly used for off-line data analysis to detect the causes of variations in the product quality.

This is addressed within the EU's Research Project CONSENS [3] by the development and integration of a smart NMR module for process monitoring. The presented NMR module is provided in a mobile explosion proof housing and involves a compact spectrometer together with an acquisition unit and a programmable logic controller for automated data preparation (phasing, baseline correction), and evaluation. Such "smart sensors" provide the basis for the future project "Industrie 4.0", and Industrial Internet of Things (IIoT), along with current requirements to process control, model based control, or soft sensing. The module transforms the acquired online spectra of various technically relevant reactions to either conventional 4–20 mA signals as well as WiFi based OPC-UA communication protocols. This enables NMR-based advanced process control for the progressive or result in bloodcurdling discussions with plant managers along with automation and safety engineers.

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Label-Free NMR-Based Dissociation Rates Determination

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Protein functions can be accurately modified with small-molecule ligands. In addition to the equilibrium binding affinity of such ligands to biomacromolecules the kinetics and lifetime of the complex formation are of interest. Indeed, there is now growing experimental evidence linking ligand efficiency with ligand-receptor complex lifetime (τ), raising the interest in the determination of dissociation rate constants ($k_{\text{off}} = 1/\tau$).^[1]

Recently, the High Power Relaxation Dispersion (RD) technique has been developed in our group, allowing the detection of single-digit-microsecond chemical exchange events.^[2] Here we present the use of a 1D ¹H $R_{1\rho}$ experiment on unmodified ligands, from which binding dissociation can be easily extracted.^[3] This method has advantages over previously proposed CPMG RD techniques.^[4] First, ¹H spectra does not require expensive, and unpractical, isotope-labeled samples; second, the experimental time is significantly shorter; third, the ¹H nucleus is differently sensitive from ¹³C to the conformational state, e.g. rotameric states, reducing the weight of bound ligand slow dynamics (if any) that might obscure the binding event; and fourth, the use of $R_{1\rho}$ experiment instead of CPMG makes possible the use of any multiplet from the ligand due to the absence of J coupling modulation during the spin-lock period. Additionally, the use of the High-Power RD approach improves the detection limit from residence times of 500 μs to the single digit μs .

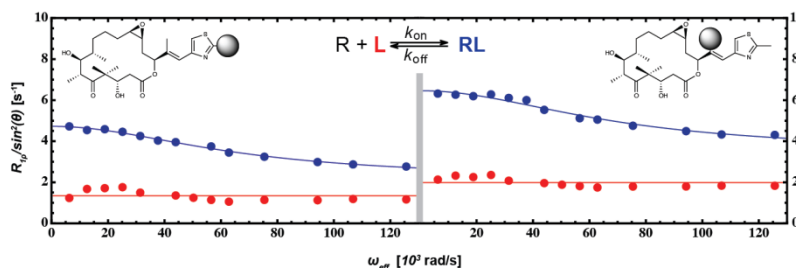


Fig. 1. Relaxation Dispersion profiles of Epothilone A in the absence (red, flat line) and in presence (blue) of tubulin heterodimers. The solid lines indicate the individual fit of each resonance. Globally, both resonances fitted to a $k_{\text{off}} = 6.88 \pm 0.47 \cdot 10^4 \text{ s}^{-1}$, which translates into a half-life $\tau = 14.52 \pm 0.93 \mu\text{s}$.

We show the applicability of our method with different small-molecule binding to two different target systems: the soluble bovine serum albumin and stabilized microtubules. Due to the inherent high sensitivity of ¹H 1D sampling different ratios of ligand to target biomolecule can be quickly acquired, allowing the deconvolution of the $\phi_{ex} = p_a p_b (\delta_a - \delta_b)^2 \omega_0$ term and therefore the extraction of the K_D together with the k_{off} .

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Reading Between the Lines – Automated Data Analysis for Low Field NMR Spectra

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For reaction monitoring and process control using NMR instruments, in particular, after acquisition of the FID the data needs to be corrected in real-time for common effects using fast interfaces and automated methods. When it comes to NMR data evaluation under industrial process conditions, the shape of signals can change drastically due to nonlinear effects. Additionally, the multiplet structure becomes more dominant because of the comparably low-field strengths which results in overlapping of multiple signals [1]. However, the structural and quantitative information is still present but needs to be extracted by applying predictive models.

We present a range of approaches for the automated spectra analysis moving from statistical approach, (i.e., Partial Least Squares Regression) to physically motivated spectral models (i.e., Indirect Hard Modeling [2]). By using the benefits of traditional qNMR experiments data analysis models can meet the demands of the PAT community (Process Analytical Technology) regarding low calibration effort/calibration free methods, fast adaptations for new reactants, or derivatives and robust automation schemes.

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Chiral Hemicucurbit[8]uril Complexation with Anionic Guests: an NMR Titration and Variable Temperature NMR Study

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Ion recognition and transport are of crucial importance in biological systems, which brings about the quest for synthetic receptors that are capable of binding ions under physiological conditions. Many macrocycles, including crown ethers[1], cryptands[2] as well as cucurbiturils[3], have a rich history of cation recognition, while development of anion receptors that are effective in protic solvents remains challenging.[4–7] Limited examples of such anion receptors include cyclopeptides that have binding affinity towards halides and SO_4^{2-} anions;[8,9] the Gibb's octa-acid cavitant that is capable of binding partially hydrated anions at alkaline pH values[10] and cyclodextrins that bind dodecaborate dianions according to their size[11]. The cucurbituril family has been explored as ion receptors due to their well-defined hydrophobic cavity.[12,13] Currently, babus[6]urils hold the record for the strongest anion binding ($K_a = 5.5 \times 10^7 \text{ M}^{-1}$) by a neutral host in an exclusively protic solvent.[14]

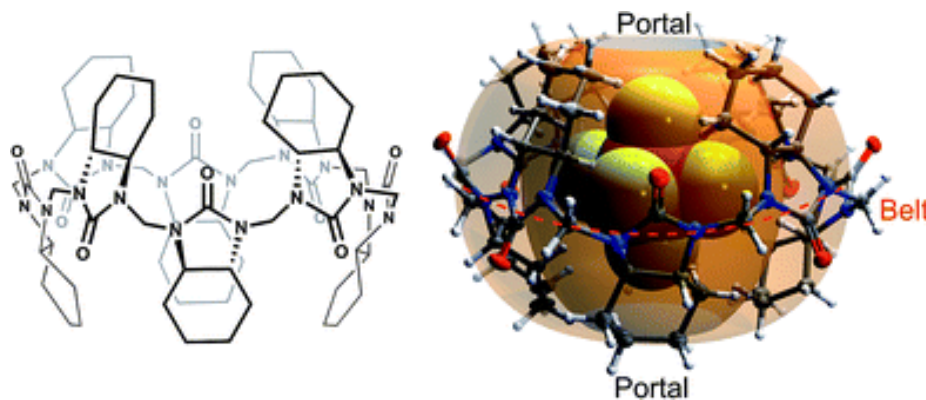


Figure 5 Molecular structure of (all-R) cyclohexanohemicucurbit[8]uril, cycHC[8] (left), and the X-ray structure of an inclusion complex with SbF_6^- (right).

Riina Aav's group previously showed the synthesis of the first 8-membered hemicucurbituril (**cycHC[8]**) that was synthesized by an approach utilising anion-templating (Fig. 1).[15] As larger anions, such as PF_6^- and CF_3CO_2^- , were effectively used as templates, we proposed that **cycHC[8]** could efficiently complex with other larger inorganic anions. Our complexation study included anions used in ionic liquids (BF_4^- , PF_6^- , SbF_6^- , CF_3SO_3^-)[16] and as oxidizing agents (ClO_4^- , IO_4^-)[17]. Additionally, such anions are considered environmental pollutants and therefore, binding of these anions in protic media is important from a biological and environmental point of view.[17,18] In this abstract and presentation, we focus on the NMR aspects, namely NMR titration experiments and variable temperature NMR studies, of **cycHC[8]** with anionic guests.[19]

We examined anion complexation with **cycHC[8]** in solution using ^1H NMR spectroscopy. The complex with SbF_6^- showed the largest complexation-induced chemical shift changes, together with significant broadening of peaks at room temperature. This suggests slow exchange on the NMR timescale with SbF_6^- , which induced our low-temperature NMR study for this system (details below).

Utilising NMR titration measurements, we determined association constants of binding with a number of anionic guests, brought in Table 1. For tetrahedral and octahedral anions, we observed growing affinity to the host with the increasing size of the guest, ranging from 48 M^{-1} for the smallest tested anion BF_4^- to $250\,000 \text{ M}^{-1}$ for the largest tested octahedral anion SbF_6^- . Furthermore, we observed that smaller anions BF_4^- and ClO_4^- , which are able to form only one or two C–H \cdots anion interactions in a given orientation, are bound with considerably lower affinities.

We were further able to establish that charge distribution around the anion plays an important role in complexation affinity. The binding of roughly octahedral CF_3SO_3^- is dramatically lower compared to the similarly sized octahedral guest SbF_6^- , despite of the several interactions between the host and the encapsulated CF_3SO_3^- apparent in our determined crystal structure. Due to the inherent symmetry of **cycHC[8]**, a possible argument is that binding might be stronger with anions having the charge equally distributed over the surface. To assess this hypothesis, a control experiment was conducted with CF_3CO_2^- , similar in volume to PF_6^- , but with a charge distribution resembling that of CF_3SO_3^- . The fact that CF_3CO_2^- , which effectively templates the synthesis of **cycHC[8]** (in acetonitrile) and has been shown by diffusion NMR to bind to **cycHC[8]** in chloroform, does not bind to **cycHC[8]** in methanol ($K_a < 10$), indicates the binding of anions to **cycHC[8]** in methanol to be sensitive to charge distribution around the anion.

Table 1 Association constants K_a for **cycHC[8]** inclusion complexes with anions, measured in MeOD at 288 K by ^1H NMR titration experiments.

Anion	Cation	$K_a \text{ (M}^{-1}\text{)}$
SbF_6^-	Na^+	$(2.5 \pm 0.7) \times 10^5$
PF_6^-	Bu_4N^+	$(2.8 \pm 0.4) \times 10^4$
PF_6^-	Na^+	$(2.0 \pm 0.2) \times 10^4$
ReO_4^-	Bu_4N^+	$(4.7 \pm 0.4) \times 10^3$
IO_4^-	Na^+	$(1.8 \pm 0.2) \times 10^3$
ClO_4^-	Bu_4N^+	$(4.7 \pm 0.2) \times 10^2$
BF_4^-	Bu_4N^+	$(4.8 \pm 0.4) \times 10$
CF_3SO_3^-	Bu_4N^+	$(3.9 \pm 0.5) \times 10$
CF_3CO_2^-	Bu_4N^+	<10

Subsequently, additional experimental insight into the kinetics of the complexation and the reaction pathway was gained by variable temperature NMR (VT-NMR) studies of SbF_6^- and PF_6^- , using a 2 : 1 host-to-guest ratio (Fig. 2). The complexation reaction order was determined by dilution experiments near the coalescence temperature (241 K and 253 K for SbF_6^- and PF_6^- , respectively).

Reaction rates remained constant upon dilution, indicating that the complexation process follows first order kinetics, characteristic for a unimolecular reaction. This suggests that the overall complexation reaction occurs via a low-energy pre-complex. Moreover, the complexation reaction rate constants for both SbF_6^- and PF_6^- were determined, allowing us to derive the activation parameters, using the Eyring equation. As expected, the complexation rate constant was an order of magnitude higher for PF_6^- than for the larger SbF_6^- , also in good agreement with our initial solution studies by NMR. The entropy of activation of the complexation is negative for both anions, as expected for the host and guest forming one host–guest complex.

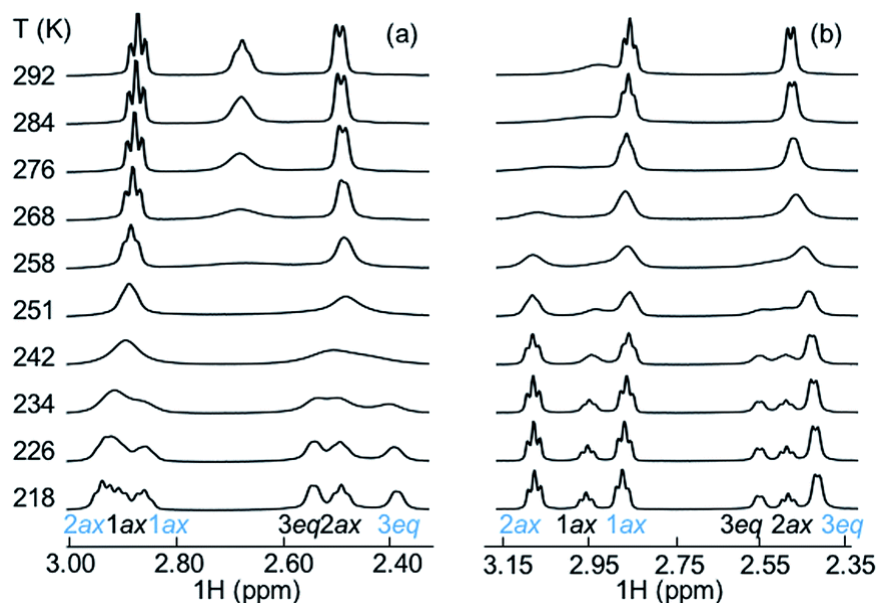


Figure 6 Evolution of proton resonances in the variable temperature NMR study (a) for PF₆⁻ and (b) for SbF₆⁻. The cycHC[8] and guest concentrations were 2.6 mM and 1.5 mM in MeOD solution, respectively.

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SHARPER NMR

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To maximise the NMR signal intensity, it is beneficial to remove the splitting due to J -couplings. However, limitations of NMR probe technology mean that decoupling in the traditional sense is not always possible. To overcome these limitations, a new NMR method referred to as SHARPER (Sensitive, Homogeneous And Resolved PEaks in Real time) was developed. The method removes *all* homo- and heteronuclear couplings of a selected signal, producing narrow singlets with linewidths approaching limits dictated by the spin-spin relaxation of the observed nucleus. The resulting increase in the signal-to-noise ratio, lowers the detection threshold, allowing minimal concentrations of solutions to be studied.

In addition, the SHARPER method eliminates limitations of the magnetic field inhomogeneity. This allows detection of inherently inhomogeneous systems, such as biphasic solutions, agitated gas liquid mixtures and gels.

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Towards the Optimization of the Identification and Quantification of Serum Metabolites

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The metabolic profiling of serum can provide important information about physiological and pathological states and may aid understanding of the mechanism of disease occurrence [1].

A quantitative analysis of the metabolites contained in serum samples can be performed using ¹H NMR spectroscopy [2]. However, due to the complexity of this matrix, NMR-based profiling has remained manual, resulting in a slow, expensive, and error-prone procedures that have hindered clinical and industrial adoption of metabolomics via this analytical technique.

Here we present an approach that exploits Mestrelab software, and in particular its SMA (Simple Mixture Analysis) plugin [3], which can quickly, accurately, and autonomously produce serum sample metabolic profile in a very short time. In particular, we have implemented a library of about 40 compounds which, given a 1D ¹H NMR spectrum, can autonomously determine and quantify the metabolic profile in less than 1 minute.

One of the greatest limitation related the NMR analysis serum samples depends on the fact that spectra include both sharp/narrow peaks, from small molecule metabolites, and broad peaks, from proteins and lipids. The size difference between macromolecules and small molecule metabolites provides an excellent basis for ultrafiltration of the serum. Nowadays, the easiest and most reliable method is to pass the sample through a 3 kDa molecular weight cut off (MWCO) micro-centrifuge filter in order to separate metabolites from proteins and other large molecules [4]. Nevertheless, this is a time-demanding procedure that implies risks in sample manipulation and loose of information regarding macromolecules contained therein. For this reason, we are exploring the possibility to filter NMR peaks *a posteriori* on the basis of peak linewidths as determined by global spectral deconvolution [5]. This approach will allow to save time in sample preparation and to retain information from the high molecular weight components of serum.

Acknowledgements

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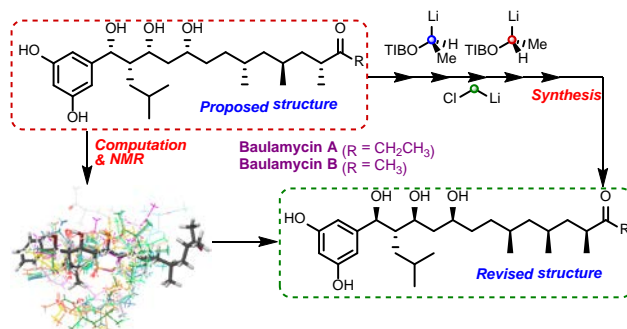
Synergy of Computation, NMR and Synthesis to Elucidate the Correct Structure of Baulamycins

Siying Zhong, Jingjing Wu, Paula Lorenzo, Muhammad Ali, Craig P. Butts, Eddie L. Myers and Varinder K. Aggarwal

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Structural elucidation of acyclic natural products, in particular the determination of relative configuration, still remains a challenge^[1] because the observed time-averaged NMR parameters, such as scalar coupling constants, are strongly influenced by the molecules' complex dynamic conformation(s) in solution state.^[2]

The configuration of a recently isolated polyketide Baulamycin A,^[3] which exhibits potent antibacterial activity, was reassigned using a combination of Density Functional Theory (DFT) calculations and synthesis. In this work, DFT calculations were employed to predict Boltzmann-averaged ¹H-¹H, ¹H-¹³C scalar coupling constants and ¹H-¹H distances. Comparison between the computed NMR parameters with the experimentally determined values eliminated up to 120 out of the possible 128 diastereomeric candidates. Finally, synthesis^[4] allowed the relative and absolute configurations of Baulamycin A to be positively identified.



Our work demonstrates the power of using a combination of computation and NMR in structural elucidation of acyclic natural products. Further developments in accurate conformational analysis by computation, together with new NMR experiments for accessing hard-to-measure NMR properties, would enable us to confidently and reliably determine the structures of flexible open chain-molecules exclusively *in silico*.

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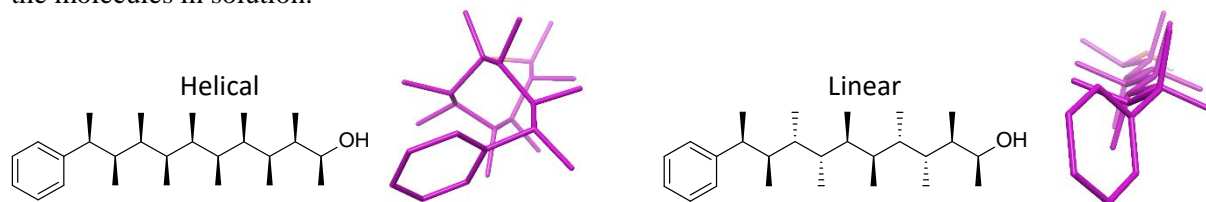
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Conformationally Constrained Hydrocarbon Inhibitors of the p53-Mdm2 Protein-Protein Interaction

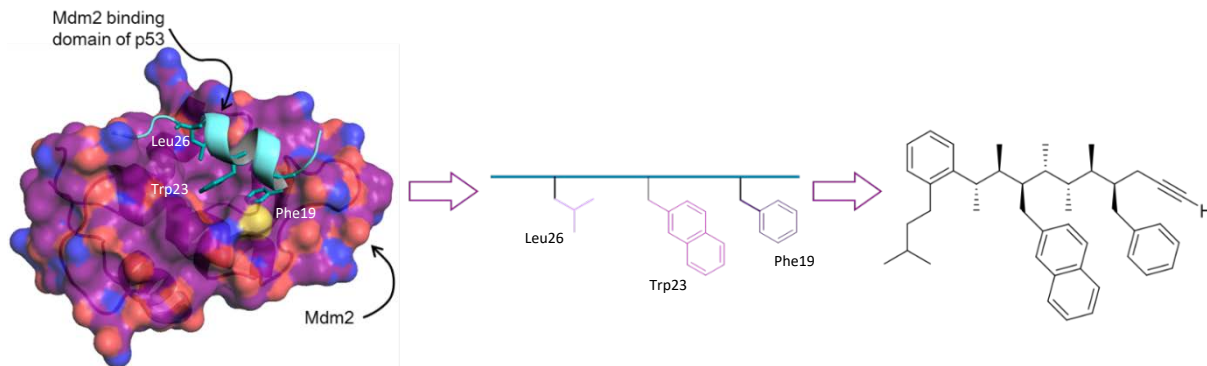
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We aim to develop a library of conformationally controlled hydrocarbons that can inhibit the p53-Mdm2 protein-protein interaction (PPI). Previously, we have established that contiguously methyl-substituted hydrocarbons, accessible through iterative lithiation-borylation, are conformationally constrained into either helical or linear conformations depending on the stereochemistry of the methyl groups. NMR data based on NOE-derived interproton distances and scalar coupling constants confirmed the conformation of the molecules in solution.¹



Both the linear and helical conformations project side chains onto the same face of the molecule with distances between them varying from 5-7 Å. These distances closely match those observed between the residues on an α -helix, making these potential α -helix mimetics, capable of inhibiting PPIs that are mediated through an α -helix, such as the p53-Mdm2 PPI. The p53-Mdm2 PPI is dominated by three 'hot-spot' residues on p53: Phe19, Trp23 and Leu26 which insert into a hydrophobic cleft on Mdm2.² We envisage carrying out an iterative lithiation-borylation sequence using the appropriate substituents at judicious points in the sequence to design a hydrocarbon framework that is capable of mimicking the three 'hot-spot' residues on p53. The conformation of the molecule must be retained on addition of the 'hot-spot' residues to ensure that the groups are located at analogous distances and angular relationships to those on p53.



To experimentally confirm their conformation in solution, we will use a combined NMR and computational approach, comparing experimental interproton distances derived from 1D-NOE and scalar coupling constants to those calculated at the DFT level of theory. EXSIDE and IPAP-HSQMBC will be used to

extract $^nJ_{\text{CH}}$ coupling constants. Finally, we will use well established biological assays to determine their binding affinity and specificity to the Mdm2-p53 interface. We hope that this project will serve as a strong foundation for the use of conformationally constrained hydrocarbons as scaffolds for the design of proteomimetics targeting a number of other important PPIs.

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Approaching DFT Accuracy for $^3J_{\text{CH}}$ Calculation Using Empirical Equations

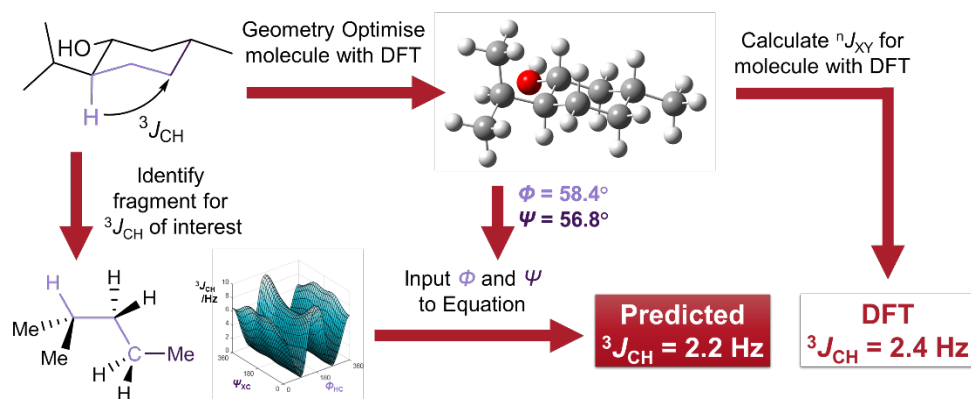
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NMR spectroscopy has a wide application in the determination of three-dimensional molecular structure and the use of three-bond scalar couplings is common in solution-state NMR. Scalar coupling between nuclei separated by three bonds can be described empirically by homonuclear (^1H - ^1H)¹ and heteronuclear (^1H - ^{13}C)^{2,3} equations relating the magnitude of the scalar coupling constant to the dihedral angle between the nuclei. Computational methods can also predict NMR properties such as chemical shift and scalar coupling constants for a given molecular structure⁴.

Over 140,000 density functional theory (DFT) calculations were used to examine the relationship between $^3J_{\text{CH}}$ and dihedral angles (Φ and Ψ) for >75 different molecular fragments. The effects of bond angle, substituent pattern and coupling pathway were examined. These DFT-calculated $^3J_{\text{CH}}$ were then used to identify and parameterize suitable equations relating $^3J_{\text{CH}}$ to the dihedral angles Φ and Ψ for different fragments.

The performance of this library of equations was tested using computationally determined $^3J_{\text{CH}}$ as shown below. This fragment-based approach achieved an accuracy of ~0.5 Hz compared to ~1.0 Hz for literature methods.



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Non-Animal Testing of the Skin Sensitization Potential of Chemicals

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The main step in skin sensitization is the reaction of small molecules with the proteins and peptides. These adducts are recognized by the immune system which results in the disease known as contact allergy. The current approach to detect the sensitization power of chemicals, the local lymph node assay [1], includes the use of laboratory animals. For this reason development of alternative testing methods for the skin sensitization potential of chemicals is desired. Several approaches have been developed, like QSAR modelling [2] or the Direct Peptide Reactivity Assay.[3]

This work aims to achieve a better understanding of the different mechanisms in the reaction of a chemical with a protein. In principle five different types can occur: Acylation, Michael-Addition, Imine forming and Nucleophilic substitution (either S_N2 or S_NAr). Amino acids and small peptides are used as test substances to identify the reaction mechanism as well as the reaction rate. For the structural characterization mainly NMR spectroscopy is used. Kinetic studies are performed with either NMR spectroscopy or with a HPLC chemoassay. For several chemicals it is not clear which reaction is the rate determining step, because in the HPLC chemoassay only the loss of the nucleophile signal is monitored. For the identification of products and mechanism NMR spectroscopy is very important. One example is the reaction of Trinitrobenzenesulfonic acid (TNBS) with amino acids or small peptides. This system had already been investigated and it was shown that the skin sensitization potency was underpredicted in correlation to the rate constant [4]. We could show that for TNBS different reactions occur, depending on the pH. The reaction is ten times faster at pH=7.4 then at pH=6 and more products can be observed.

Other molecules, like Acrolein, can have more than one reacting center. This substance can either react with Imin-formation or with Michael Addition. By NMR spectroscopy we were able to identify the preferred reaction mechanism, as well as the corresponding products.

As mentioned above many reactions are generating a product mixture, which depends on the number of reacting centers or the mechanism. For these mixtures it is important to use a separation processes before the structural studies. Therefore we try to develop a coupling method of HPLC with ultra-high resolution FT-ICR-MS. Together with NMR, this will improve our understanding of the reactions.

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Characterisation of Catechol and (4-fluoro)-phenylboronic Acid Binding System: Investigation by F19-NMR and Modelling of Equilibria

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The multi-step equilibria which are hallmarks of the reaction [1] between catechol and (4-fluoro)-phenylboronic acid are measured and characterised by F19-NMR in buffered aqueous media across a range of pH. The recorded species concentrations show very good agreement with the data values from a model of a linked set of equilibria for the reaction. Global curve fitting yields equilibrium constants for all steps which are consistent across all conditions tested for this reaction. This process provides a template for robust and facile interrogation of analogous equilibria in other systems.

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Intramolecular Hydrogen Bond in p-Substituted o-(N-Diethyl)aminomethylphenols: NMR Study

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The aminomethylphenols are commonly used as catalysts for supramolecular systems, epoxy resins and in the synthesis of medicaments with multiple pharmacological activities. The availability of intramolecular H-bonds causes the specificity of physicochemical properties of the aminomethylphenols.

A series p-substituted o-(N-diethyl)aminomethylphenols (o-DEAMPH), $\text{XC}_6\text{H}_5\text{CH}_2\text{Y}$ ($\text{X}=\text{p-OCH}_3, \text{CH}_3, \text{H}, \text{F}, \text{Cl}, \text{Br}, \text{COOCH}_3, \text{CN}$ and NO_2 , $\text{Y}=\text{o-N}(\text{C}_2\text{H}_5)_2$) were studied using Nuclear Magnetic Resonance spectroscopy (NMR). The chemical shifts of the bridging hydrogen atom in o-DEAMPH have been measured in CCl_4 solutions. Moreover, ^1H NMR spectra of compounds were obtained at B3LYP/6-311+G(d,p) level using the GIAO method. Excellent linear correlation ($R^2=0.9996$) observed between values the chemical shift of hydrogen atom obtained experimentally of ^1H NMR and calculated B3LYP/6-311+G(d,p). A linear relationship revealed the chemical shift of the hydrogen atom, δ_{OH} with the charge on the hydrogen atom, $Q_{\text{A}}(\text{H})$, calculated by the quantum theory atoms in molecules (QTAIM) ($R^2=0.9902$). It is found that a linear correlation between experimental assignment phenol hydroxyl proton chemical shifts, δ_{OH} and theoretical calculated values frequencies of the O-H stretching mode, ν_{OH} : $\delta_{\text{OH}} = 58.4151 - 0.0149\nu_{\text{OH}}$ ($R^2=0.9872$). The values hydroxyl proton chemical shifts, δ_{OH} also correlates linearly with the para-substituent constants, σ_{p} ($R^2=0.9882$). In addition, the intramolecular O-H...N bond energy was estimated from the empirical relationship. It is shown that from the standpoint of their strength the intramolecular O-H...N hydrogen bonds can be classified as strong interactions. The intramolecular hydrogen bonding energies of the considered systems range from 6.5 to 8.5 kcal/mol. Intramolecular hydrogen bond energy is linearly correlated with Hammett constants, σ_{p} ($R^2=0.9903$).

Moreover, numerous relationships between the hydrogen bond energy obtained from empirical ^1H NMR data and predicted geometric descriptors of hydrogen bond: $d_{\text{H}\dots\text{N}}$ ($R^2=0.9984$), $\angle \text{O-H}\dots\text{N}$ ($R^2=0.9979$) as well as topological descriptors of hydrogen bond strength: total energy density ρ^{BCP} ($R^2=0.9964$) and the density of potential energy V ($R^2=0.9980$) at the bond critical point of the proton-acceptor (H-N) distance (QTAIM) and the core-valence bifurcation index CVB (based in ELF, $R^2=0.9945$) were obtained. From these dependencies excluded the point corresponding to $\text{R}=\text{H}$. Relationships between of values hydrogen bond energy and various quantities characterizing of o-DEAMPH are discussed.

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Methanol Self-association and Preferential Solvation of Chelating Agents for the Extraction of Nuclear Fission products in Supercritical CO₂

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While supercritical carbon dioxide (sCO₂) is a promising solvent in heavy metal extractions due to its low toxicity, surface tension and viscosity, many chelating molecules exhibit limited solubility in sCO₂. The addition of volatile polar molecules to sCO₂, such as methanol (MeOH), improves the solubility of chelating molecules and metal-ligand complexes. MeOH self-assembly and preferential solvation of 2-ethylhexylphosphonic acid mono-2-ethylhexyl ester (HEHEHP), a chelating ligand, in sCO₂ at 19.7 MPa and 323 K was investigated with diffusion-ordered nuclear magnetic resonance spectroscopy (DOSY-NMR) and molecular dynamics simulations. DOSY-NMR pulse sequences contain a pair of pulsed field gradients that attenuate the NMR resonance and enable calculation of diffusivity for each NMR active component in the mixture. Diffusometry results were analyzed using a model that integrates the Stokes-Einstein relationship with self-association models to approximate the size-dependent, energy of cluster formation landscape. DOSY-NMR results are consistent with molecular dynamics simulations and suggest the formation of small clusters of MeOH limited to a composition of under 10 molecules in a cluster at high concentrations of MeOH.

Next, self-assembly of HEHEHP into dimers was quantified with DOSY-NMR in sCO₂ at 19.7 MPa and 323 K. The propensity of HEHEHP to self-associate into dimers in sCO₂ was evaluated with the integrated Stokes-Einstein, self-association model. The results suggest that HEHEHP is primarily composed of dimers at concentrations of over 40 mM in sCO₂. Finally, we compare the extent of dimerization of HEHEHP in sCO₂ containing 4 mole % of MeOH to the dimerization of HEHEHP in neat sCO₂ with a series of DOSY-NMR experiments. The experiments indicate that MeOH preferentially solvates HEHEHP, increasing both the solubility of HEHEHP in the supercritical fluid and the weight fraction of monomeric HEHEHP. Our results suggests that the increased amount of monomeric chelators may be an additional, synergistic factor in the enhanced heavy metal extraction in MeOH modified sCO₂.

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Molecular Networking Using ^{13}C NMR Can Facilitate Natural Product Structure Elucidation

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Structure elucidation of natural products is a complicated and challenging research problem, with variables of subjectivity including the experience of the spectroscopist, the particular structure(s) to be determined, and instrumental capabilities, to name a few. Modern methods, such as the incorporation of RDC and LR-HSQMBC experiments, advanced data analysis using computer-assisted structure elucidation, and other innovative approaches are changing the way that unknowns are characterized (when cost-permissive or otherwise feasible).[1-2] Similarly, the dereplication of natural products in mixtures using NMR, LC-MS, and other methodologies has had a profound impact on the efficiency and pace of new natural product isolation and discovery.[3-4] Recently, a method has emerged for the simplification of ^{13}C NMR data that allowed construction of a large-scale data library from values obtained experimentally, computationally, or even by extraction from literature reports.[5] Thus we have undertaken an informatics-based research project to evaluate whether or not the use of ^{13}C fingerprinting, followed by comparative array analyses, can facilitate natural product structure elucidation. Indeed, the evaluation of these data can be completed by several different metrics, including the calculation of Tanimoto coefficients, Pearson correlation coefficients, Jaccard indices, normalized euclidian distances, and cosine values. As a proof of principle, each of these computations was completed for a set of previously isolated and already characterized natural products (including many analogues from the same structural class). The results to be shown in this presentation indicate that not only can the simplified ^{13}C NMR data rapidly facilitate the classification of known natural products and analogues thereof, but furthermore that specific variations in functional group substitutions will also lead to molecular network connections that can provide other clues to the structure of unknowns.

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General Approach to Access Long-Range ^1H - ^1H RDCs

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Residual dipolar couplings (RDCs) are a powerful means for conformational and configurational analyses [1, 2]. For small molecules, ^1H - ^{13}C one-bond couplings are mostly exploited. However, the quality of, for instance, configurational analysis greatly depends on the number of RDCs that sample independent orientations, which may not always be provided by one-bond couplings alone. Ways to obtain long-range RDCs are thus in high demand. Long-range ^1H - ^1H RDCs would be an obvious choice, but are very challenging to access. The main reason is that, in alignment media such as PBLG, the high abundance of ^1H - ^1H RDCs results in broad multiplets that overlap and from which individual splittings are not resolvable. Here, we propose a general approach to obtain ^1H - ^1H RDCs.

We demonstrate that the recent PSYCHEDELIC experiment [3] gives access to long-range ^1H - ^1H couplings at a resolution close to the natural linewidth. PSYCHEDELIC delivers a 2D J-resolved spectrum with absorption mode lineshapes and with only ^1H - ^1H couplings to one or more chosen spins, thus allowing coupling measurement as simple doublets at pure shift resolution and resolving both the issues of spectral overlap and multiplet complexity. The experiment is based on PSYCHE [4], which allows broadband homodecoupling at good sensitivity and tolerance to strong coupling. The tolerance to strong coupling can be further enhanced by introducing frequency-swept 180° pulses applied during gradients [5]. Although PSYCHEDELIC delivers the magnitude of the ^1H - ^1H coupling, it offers no sign information, which is crucial for RDC interpretation. In isotropic samples, E.COSY or similar experiments can provide relative sign information, but in weakly aligned samples the overflow of ^1H - ^1H couplings complicate cross-peak analysis. Solutions based on P.E.COSY or z-COSY using similar principles as PSYCHEDELIC to provide relative sign information of only selected couplings will be introduced.

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50 Solution-State Structure Determination of Lactoferrin-Derived Peptides, Acting Against Influenza

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Bovine lactoferrin is a glycoprotein playing a key role in innate immunity against infections. Previously, we have demonstrated that selected peptides from bovine lactoferrin C-lobe are able to prevent both cell infection and Influenza virus hemagglutination [1]. These findings prompted us to further investigate the ability of lactoferrin derived peptides to inhibit Influenza virus infection. In this study, we present the structural investigation of new bovine lactoferrin C-lobe derived sequences and their ability to inhibit infection and viral hemagglutination.

By means of NMR spectroscopy, we have investigated the solution structures of new five peptides (**1-5**) derived from loop Ser418-Pro429 of Bovine lactoferrin (PDB ID: 3IB0) [2], in order to determine the structural elements responsible for binding towards the macromolecule. In details, we have resolved the solution structure (HFA/H₂O) of **1** (SKHSSLDCVLRP) and **2** (SLDCVLRP), finding a γ -turn for **1**, and β -turn for **2**. The global turn observed for **1** and are in agreement with conformation of the lactoferrin loop Ser418-Pro429 (PDB ID: 3IB0). In particular, the helix 3_{10} formed by Cys425-Leu427 of protein loop is overlapped with the γ -turn of **1** (Val9-Arg11) and β -turn of **2** (Cys4-Arg7). The tetrapeptides **3** (VLRP), **4** (SLDC) and **5** (SKHS) gave rise to very low number of inter-residue NOE effects, due to their expected highly flexibility, hampering the solution structure determination. However, we may suggest that the preferred conformation of **3-5** is similar to the spatial arrangement observed for **1**, **2** and the loop Ser418-Pro429 as highlighted by the biological activity of **3-5**. Indeed, The tetrapeptides showed an anti-Influenza activity in a concentration range femto- to picomolar. Our data indicate that these peptides may constitute a non-toxic tool for potential applications as anti-Influenza therapeutics.

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Study of Ambroxol Hydrochloride and Excipients in Infant Syrups by ^1H NMR

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Ambroxol Hydrochloride (AH) is a drug with mucolytic activity used as an expectorant and found in commercially available pharmaceutical preparations in the form of drops, injections, syrups and tablets and is widely used in pediatric patients. International pharmacopoeias advocate methods for the determination of AH only in pure form, which makes it impossible to apply these methods in formulations, and the excipients may interfere with the quality control analysis. A promising technique for this type of case is Nuclear Magnetic Resonance¹⁻³. The objective of the present study is to use the Nuclear Magnetic Resonance (NMR) technique in the characterization and quantification of the active principle, in addition to investigating the presence of ethanol in children's syrups with ambroxol hydrochloride as the active compound. The ERETIC method provides a reference signal, synthesized by an electronic device, which can be used for the determination of absolute concentrations after a previous calibration.⁴ Ten commercial samples were analyzed in triplicate, and were performed the characterization of the main compounds present in the formulations. Seven samples showed characteristic ethanol signals and based on the qualitative analysis of the spectra the triplet (CH_3 of ethanol) was selected as quantifiable signal for ethanol. For AH, a singlet was selected as a quantifiable signal. According to National Formulation of the Brazilian Pharmacopoeia, 2nd edition Rev. 02, syrups must not present ethanol in their composition, and their minimum presence must be declared in the package leaflet. However, through the study in question, it was possible to quantify the ethanol in the formulation without the declaration, and one of the samples had a value of 1.8 °GL, therefore being classified as an alcoholic beverage according to a Brazilian decree 6.117/2007. There was also the quantification of the AH content, where it was possible to observe that only one of the samples had the declared content of 2.7 mg/mL in 120 mL of solution, what could affect the therapeutic effect of these syrups.

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Interactions between Diet and Microbiota Studied by NMR-Based Metabolomics

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The gut microbiome is a comprehensive ecosystem that regulates many physiological functions. It affects the evolution of the immune system, influences cell proliferation, and ensures metabolism of bile salts as well as indigestible polysaccharides and proteins [1]. An individual's diet is one of the main factors affecting the gut microbial composition, and the catalytic activity of intestinal microbiota determines an individual's health status. Hence, investigating the interactions between the gut microbiome and diet is crucial for understanding how serious metabolic diseases develop, e.g. metabolic syndrome, colorectal cancer, and others.

Lately, NMR-based metabolomic analysis of fecal extracts has become a well-established platform for the study of the gut microbiota and host metabolism through the analysis of the endproducts of their metabolism. It is advantageous to analyze feces as it has immediate contact with the colon and is easily sampled, which makes it a relevant material for exploring both the gut microbiota metabolism as well as its co-metabolism with the host.

In this study, we focused on the impact of different microbial colonizations and diets on the metabolic composition of fecal samples in the experimental model of BALB/c and C57BL/6 mice respectively. Water fecal extracts were analyzed by 600 MHz NMR spectrometer using 1D-NOESY pulse sequence. Metabolites identification was supported by information extracted from J-resolved, COSY, and HSQC spectra. Statistical analysis of the acquired data was performed using a combination of univariate (parametric and non-parametric tests) and multivariate (principal component analysis (PCA) in addition to the partial least square discriminant analysis (PLS-DA)) approach.

Firstly, we compared the fecal metabolome of conventionally colonized (CV) male mice fed by two different diets: a standard murine diet and a diet with excess protein and fat ("Western-type diet", WTD). The WTD significantly increased the levels of short-chain fatty acids (SCFAs; namely acetate, propionate and butyrate), which are an important energy source for cells and possess other properties favourable for the host organism. Enhanced protein breakdown might explain the increased concentrations of alanine, glutamate, and 5-aminovalerate in WTD-treated mice. Additionally, we observed the elevation of fumarate and succinate, indicating more intensive carbohydrate breakdown in mice on WTD.

Secondly, we were looking for differences in fecal metabolome of germ-free (GF), monocolonized (*Escherichia coli* Nissle 1917) and CV mice, all fed the same standard diet. Statistical analysis of NMR spectra was able to unequivocally differentiate among samples from these three groups. Obviously, the largest differences were observed between CV and GF mice. These included elevated levels of SCFAs (butyrate, propionate and acetate) in CV mice, as their production from polysaccharides or proteins is conditioned by the presence of specific bacteria. On the contrary, the absence of bacteria in GF mice resulted in higher levels of saccharides, e.g. raffinose. Therefore, raffinose, which can be digested only by

certain bacterial strains, may serve as a good marker of microbial activity [2]. Additionally, NMR spectroscopy allowed us to distinguish CV mice from *E. coli* Nissle colonized mice. This observation is of particular importance since the probiotic bacterium *E. coli* Nissle has been reported to have a positive effect on maintaining remission of ulcerative colitis [3].

In summary, our data show several interactions between diet and microbiota beyond the obvious relations between substrates and enzymes. Next, our metabolomic results will be correlated with abundance of particular colonies residing the gut or gut mucosa transcriptome. A follow-up study will then focus on monitoring how these differences in microbial metabolism influence the development of intestinal inflammation and colorectal cancer in this mouse model.

Acknowledgements

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Probing Torsional Flexibility Via RDC-Driven Molecular Dynamics Simulations

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Residual Dipolar Couplings (RDCs) have established themselves as an efficient NMR tool for the structure elucidation of organic molecules [1]. Unlike standard short-range NMR parameters such as chemical shifts, ³J-couplings and NOEs, RDCs make it generally possible to correlate nuclear pairs that are far apart in the molecular structure, hence providing global constitutional, configurational and conformational information.

Interpretation of experimentally determined RDCs is, however, far from being a trivial task, especially in the presence of conformational flexibility. To this end, different strategies have been developed. A promising and conceptually new approach is based on Molecular Dynamics simulations with Orientational Constraints (MDOC) [2] implemented in the COSMOS software [3] and successfully applied to stereochemical assignment of rigid and flexible molecules [4].

In this contribution, the potential of RDC-driven MD simulations is explored in the study of the torsional flexibility of small non-steroidal anti-inflammatory drugs, members of the salicylate and profen families. Realistic conformational distributions are obtained with high-quality fits of the experimental one-bond and long-range couplings used as constraints. Contrary to most existing methods, the MDOC approach does not require any optimized structure or *a priori* knowledge of the conformational surface. This feature, together with the possibility of combining isotropic and anisotropic constraints as well as the reasonable computational cost, make the COSMOS software an appealing candidate in the search of a versatile and general tool for the structure elucidation of any flexible organic molecule.

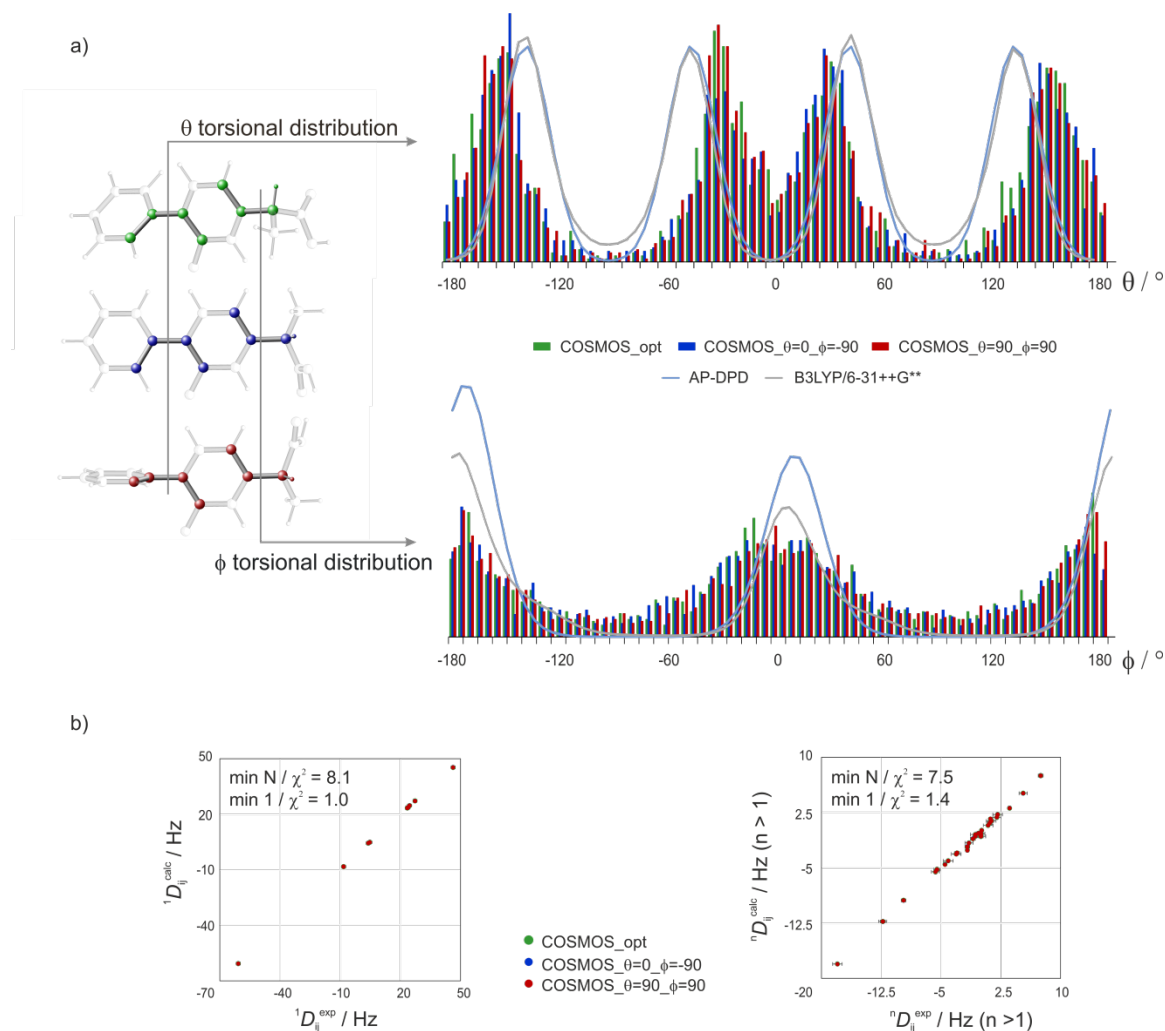


Fig. a) Torsional distributions obtained for the fluorinated anti-inflammatory drug flurbiprofen in 20-ns MDOC simulations with COSMOS software, starting from three different geometries and using as constraints RDCs measured in PBLG/THF- d_8 together with 3J -couplings, and comparison with results obtained *via* theoretical calculations at DFT level and a different conformational model known as AP-DPD approach [5]. **b)** Experimental *versus* back-calculated one-bond and long-range RDCs for the same MDOC simulations.

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Insights into Amorphous Solid Dispersions of Felodipine Using Solid-state NMR Spectroscopy: Miscibility and Molecular Interactions

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Phase separation in amorphous solid dispersions (ASDs) is still not clearly understood on the sub-nanometric scale [1], and further systematic investigations are still required. The phase separation behavior is influenced by many factors, such as the composition of drug-polymer blend, the type and the strength of drug-polymer interactions, and the method of preparation [2-4]. We demonstrated the use of solid-state NMR spectroscopy (SSNMR) to evaluate the role of the strength of drug-polymer hydrogen bonding (H-bonding) on the compositional homogeneity in ASDs of felodipine (FEL), a poorly water soluble drug, with poly(vinylpyrrolidone), or PVP, poly(vinylpyrrolidone-co-vinylacetate), or PVP/VA, and poly(vinylacetate) or PVAc. The dispersions were prepared at various drug loadings (50% to 90% w/w) via melt quenching. The blend scale miscibility was studied by examining the proton spin-lattice relaxation times in the laboratory and rotating frame ($^1\text{H } T_1$ and $T_{1\rho}$) for the drug and the individual polymer for each set of ASDs. Domain sizes were estimated via spin diffusion. The experimental data was used to elucidate the influence of the strength of drug-polymer H-bonding on the phase behavior of resulting dispersions. It was found that FEL:PVP and FEL:PVP/VA systems exhibited weak signs of nano phase separation especially for the compositions with lower polymer loadings with considerably small domain sizes. Whereas FEL:PVAc system showed pronounced signs of nano phase separation as the polymer amount decreased with larger domains. The extent of H-bonding in amorphous FEL and FEL:Polymer blends within each set of ASDs was quantified via deconvolution. The carbonyl region was used to selectively analyze the evolution of the various populations of coexisting species in the samples. The order of the strength/extent of drug-polymer H-bonding interactions was PVP > PVP/VA > PVAc. It was suggested that the strength of drug-polymer H-bonding interaction is one of the key factors in controlling the phase behavior of ASDs.

Our findings indicate that SSNMR is a useful tool for evaluating the spatial homogeneity with sub-50 nm resolution, where other conventional techniques fail, and drug-polymer interactions for the compositions of pharmaceutically relevant systems.

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Characterization of Multicomponent Crystal Forms of Neuroleptic Drugs by NMR Crystallography

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Launching new products in the market is very expensive and extremely time consuming. Envisaging these aspects, pharmaceutical industry has shown a high interest on improving the efficacy of “old” drugs by enhancing their physicochemical properties (solubility, dissolution rate and bioavailability) without changing their pharmacological behavior. This has been addressed using several approaches including the preparation of multicomponent drug crystal forms (salts and co-crystals), developed over the last years. [1-3]

Understanding how molecular systems self-assemble in the solid-state is a great challenge, with particular relevance in pharmaceutical sciences.[4] Herein, we present an experimental NMR, single-crystal / powder X-ray diffraction (XRD), and computational study of the supramolecular assemblies of Gabapentin (GBP) and amantadine (AMA); two active pharmaceutical ingredients used to treat neurodegenerative diseases, such as epilepsy and Parkinson, respectively.[5,6] For GBP, an amino-acid based drug, there are few co-crystals reported to date[7] and here we report the preparation of two organic “solid” ionic liquid drug systems featuring an unusual lamellar arrangement. The effect of crystal packing interactions was investigated and quantified using SSNMR in tandem with periodic and cluster DFT calculations, by means of a stepwise *in silico* dismantlement of the 3D packing. To better understand the synergy between weak and strong hydrogen bonds, several computer cluster fragments were evaluated based on calculated packing-induced chemical shifts.[8] In the case of AMA, the very low aqueous solubility and the bioavailability problems related to the use of the hydrochloride and sulfate salt forms,[6] lead us to prepare a set of new molecular salts through the combination with safe organic co-formers. The structural determination was carried out using powder XRD for the compounds where it was not possible to grow crystals. DFT calculations were also used in tandem with SSNMR for structure validation.[9]

Thermal stability analysis were performed, in both systems, using DSC-TGA and hot-stage microscopy. For Ama salts, preliminary solubility studies were carried out, showing an improvement of the drug aqueous solubility.

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Progress in Multi-Spectra Automatic Structure Verification

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Automatic Structure Verification is fast becoming an important part of NMR data evaluation software packages such as Mnova and others. Its basic goal is to answer, in a qualitative as well as quantitative way, the question

“Is this molecular structure compatible with these NMR data?”

Naturally, the query one makes, the answer one gets. These things are fuzzy in logic because of imperfections and impurities in the spectra, nmr parameters prediction errors, solvent effects, etc. The complexity is close to that of an artificial intelligence, and the scoring is critically dependent on the query itself.

But the problems extend beyond that, involving what one intends by “NMR data”. As we know well, it is one thing to have a single ¹H spectrum, for example, and another one to have a pair of spectra of different kinds, such as ¹H and ¹³C, or ¹H and HSQC, and a still more different one if the spectra (of presumably the same compound) include an arbitrary subset of any of the “200 and more” NMR experiments such as, for example ¹H, ¹³C, HSQC, COSY, and HMBC.

How does one proceed with the automatic analysis in such cases? It is well known that the likelihood of multiple “solutions” decreases sharply when one combines several spectra of different kinds. We also know that even ‘human’ analysis is in these cases anything but linear and standard: it requires multiple “passes” through the spectra and a non-trivial search for correlations (or lack of correlations) of various orders. We also expect that critical points can get exacerbated and build up beyond a threshold (false negatives). Since no spectrum is perfect, how many spectra it takes before any structure gets ruled out?

Over the last few years we have built a considerable body of experience [1] with these problems. How to design a software machinery to iteratively analyse a set of any number of NMR spectra of various kinds but presumably of the same compound (or better the same sample) and make it extract and refine all information they might contain in a synergetic way, and then score that information database against an assumed, hypothetical molecular structure.

In this presentation we would like to share with you some of this.

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Numeric Atom-Centered Basis Sets for Magnetic Response Calculations

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First principles calculations have become extremely useful for assigning the chemical shift peaks and J-couplings to specific nuclei. One of the difficulties that theoretical NMR calculations usually face is having good quality basis sets that are accurate in the region near the atomic nuclei, which has great influence on the NMR parameters. To date, several all-electron models have been proposed, which are based on commonly used basis functions such as Gaussian-type orbitals (GTOs), projector-augmented waves (PAW), linearized augmented plane waves (LAPW), and Slater-type orbitals (STOs).

In this work, we explore the use of numeric atom-centered orbitals (NAOs) [1] as basis sets for magnetic response calculations, and compare their accuracy to Gaussian-type orbitals. The NAOs have numerically tabulated radial functions and therefore their radial shape is fully flexible. This includes retaining the correct shape of the orbitals near the nucleus without a significant increase in the computational cost. Furthermore, following [1], the NAOs can be constructed to be strictly localized, which can lead to linear scaling with system size, which becomes beneficial for large systems. We focus on three magnetic response properties that are characterized by different degrees of localization of the perturbation: the magnetizability (defined as the second-order electronic response to an external field), NMR shieldings, and J-couplings. We present details of our implementation, in a nonrelativistic formalism, and use a test set of molecules with light atoms up to chlorine. For magnetizabilities, the NAOs demonstrate faster convergence than GTOs. For shieldings, the general-purpose NAOs show similar convergence as GTOs that are specifically optimized for the calculation of shieldings. J-couplings pose a significant challenge for basis set convergence, because the Fermi contact term requires extreme flexibility of the basis orbitals near the nucleus, as has been illustrated by strategies pursued by the most advanced GTO basis sets available [3].

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In situ MAS-NMR Studies of the Dissolution of Gibbsite and Precipitation of Lithium Hydrotalcite in High-Level Nuclear Waste Processing

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The Hanford site contains high-level radioactive waste (HLRW) that was produced during World War II and the Cold War during nuclear weapon manufacturing. HLRW is composed of dynamic colloids perturbed far from equilibrium from exposure to both ionizing radiation and extreme alkalinity. Colloidal aluminum hydroxides, known as gibbsite, is a major HLRW component. Hanford personnel are actively looking for solutions to process HLRW. One potential solution is to store the waste as glass through a process called vitrification. However, vitrification of material with high aluminum content forms polyphasic glasses with poor material properties. The vitrification strategy therefore necessitates removal of large amounts of aluminum species through dissolution processes. The aluminate in solution would then be precipitated as lithium hydrotalcite. We have completed in situ ^{27}Al , magic angle spinning, nuclear magnetic resonance spectroscopy (MAS-NMR) studies to investigate the dissolution of gibbsite and precipitation of lithium hydrotalcite to emulate the processing of HLRW. Gibbsite dissolution and precipitation of lithium hydrotalcite in aqueous solutions of lithium hydroxide (LiOH) was observed with in situ ^{27}Al MAS-NMR. In situ ^{27}Al MAS-NMR measurements were carried out on a Varian-Inova 500 MHz NMR spectrometer at 25°C using a 7.5 mm MAS probe and a homemade 7.5 mm zirconia rotor sealed with Teflon tape. Gibbsite (0.5 M) had a particle size of 400-700 nm and was dispersed in 3 M LiOH and loaded into the NMR spectrometer. Analysis of in situ NMR spectra suggests gibbsite dissolution produces tetrahedral aluminate species that interact with lithium ions in solution. Aluminate interactions with lithium are consistent with the progressive deshielding of the tetrahedral resonance. Line deconvolution was utilized to separate the broad gibbsite signal from the sharp lithium hydrotalcite resonance. The presence of two kinetic regimes during the dissolution of gibbsite suggests the phase transformation occurs through the intercalation of Li ions. Work is underway to conduct in situ ^7Li studies to investigate the formation of ion pairs and Li intercalation during the dissolution and precipitation process. A fundamental understanding of the dissolution of gibbsite and precipitation of lithium hydrotalcite will assist facilities in HLRW processing.

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Defining 'Gas-Tight:' An Examination of the Oxygen-Permeability of Several NMR/EPR Tubes

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Numerous research applications exist which require exclusion of air and/or moisture from an NMR sample. In the absence of a glovebox or similar inert atmosphere, these approaches become even more challenging. J. Young tubes (or comparable LPV tubes) are the most widely used gas-tight NMR tubes. These tubes are simple to use and seal reliably, however, they do not allow for syringe puncture, require a special glass tube to accommodate the valve, and have other limitations. In this investigation, we employed redox indicators to determine the rate of O₂ diffusion into NMR/EPR tubes with J. Young/LPV valves, screw-caps, omni-fit valves, and septa. We compared these diffusion rates to the innate rate of O₂ diffusion into an uncapped tube. Experiments with uncapped tubes demonstrated that O₂ diffusion through aqueous solutions is moderately slow. Samples in LPV tubes remained below the experimental limit of O₂ detection for hours, only showing minor oxygen contamination after one day. Tubes sealed with screw-caps and septa showed intermediate oxygen permeability. The implications of this experimental data and the practical constraints associated will be discussed.

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Evaluation of NMR Signals of Isoquinoline Alkaloids: An Experimental and Theoretical Approach

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The Amaryllidaceae family lies between the 20 most applied plant families as therapeutic agents against various diseases. An important characteristic of this family is the presence of isoquinoline alkaloids, exhibiting several biological activities such as antitumor, antibacterial, antifungal, antimalarial, antiviral, analgesic, and antiparasitic and acetylcholinesterase inhibition [1].

The innumerable studies in the literature on the occurrence, isolation and structural determination of natural products, has made them interesting objects of study in nuclear magnetic resonance (NMR) spectroscopy [2], which is a very important and well-established tool for the structural analysis of organic compounds [2-4]. These isoquinoline alkaloids homolyorine-type isolated from Amaryllidaceae plants are reported in the literature when substituted at C-2, with an OH or OCH₃ group in α position. However, the spectra of these compounds are not simple, there are overlapping ¹H and ¹³C NMR signals and many couplings that hinder the unequivocal assignment of all ¹H and ¹³C NMR signals and the determination of the relative stereochemistry of this group.

The development of experimental NMR techniques has been accompanied by advances in computing and theoretical methods (especially the DFT methods), which permit shielding tensors and spin-spin coupling constants to be calculated with highly reliable results [3,4].

We present here a complete NMR assignment for some isoquinoline alkaloids. These studies were performed using 1D and 2D NMR techniques and compared with the theoretical predictions of the chemical shifts and coupling constants using DFT calculations.

GIAO model at DFT/B3LYP level of theory using the cc-pVTZ basis (with and without solvent) set was employed for calculations of ¹H and ¹³C NMR chemical shifts (δ), using Gaussian03 program [6].

From correlation between the theoretical and experimental data, it was concluded that the model used was effective for calculating the shielding tensors for the compounds studied. From hippeastrine, for example, it was found that the mean deviation (MD), standard deviation (SD) and linear correlation factor (R) for all the calculations were generally better when using solvent from δ ¹H and without solvent from δ ¹³C. In addition, the results showed a slight indication that the OH group is in α position, as reported in the literature.

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Effect of Nitrogen Lone Pair: ${}^3J_{\text{HF}}$ versus ${}^5J_{\text{HF}}$

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Fluorine atom is present in a large range of pharmaceutical compounds such as voriconazole, an antifungal drug. As ${}^{19}\text{F}$ is an interesting probe in NMR studies, the J_{HF} is a suitable parameter for correct structure assignment of those drugs.[1] Despite the voriconazole assignment has been reported,[2] information about the magnitude and sign of ${}^nJ_{\text{HF}}$ are not provided. In the case of the pyrimidinyl moiety of voriconazole, the long-range scalar coupling between proton and fluorine nuclei, ${}^5J_{\text{HF}}$ (2.40 Hz), is observed to be larger than ${}^3J_{\text{HF}}$ (1.86 Hz). Model molecules, such as fluorobenzene, *N*-methyl-2-fluoropyridine, *N*-methyl-3-fluoropyridine, 3-fluoropyridine, 5-pyrimidine, and 2-fluoropyridine were chosen to evaluate the trend of ${}^3J_{\text{HF}}$ and ${}^5J_{\text{HF}}$. Spectral-aliased HSQC [3] was used to determine the relative sign between J_{CF} and J_{HF} . Theoretical calculation of J_{HF} coupling showed a similar trend observed experimentally. The coupling pathways for ${}^3J_{\text{HF}}$ and ${}^5J_{\text{HF}}$ couplings were decomposed using NLMO approach.[4] The main electronic factor that explain the observed trend is the high delocalization character of nitrogen lone pair, which has positive and negative contributions to ${}^5J_{\text{HF}}$ and ${}^3J_{\text{HF}}$, respectively.

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Unraveling the Selectivity for the Knoevenagel Reaction

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Aldol condensations represents a versatile class of reactions that involve the formation of carbon-carbon bond. Among these, the Knoevenagel reaction [1] is the condensation of carbonyl (usually aldehydes) and activated methylene compounds, leading to unsaturated products. Reactions with ethyl cyanoacetate as methylenic compound usually leads only to *E* isomer. Some authors [2,3] suggest that selectivity is due to steric constraints between substituents in the reaction intermediate. In this work, reactions of 2-methoxybenzaldehyde were conducted with both ethyl cyanoacetate and ethyl acetoacetate, to evaluate the effect of bulkiness of substituents.

In the reaction of ethyl cyanoacetate, only *E* isomer was observed. While a mixture of isomers (*E:Z*, 4:6) was observed in the reaction of ethyl acetoacetate. A sample analyzed (¹H NMR) one day after its purification shown enriched *E* isomer (*E:Z*, 9:1). The same sample was analyzed after four days and the 4:6 (*E:Z*) ratio was restored. This result suggests a thermodynamic equilibrium between *E* and *Z* isomers of Knoevenagel reaction. The selectivity observed for cyano-derivatives should be due to a highly favored isomer in such equilibrium. To test this hypothesis, Metternich and Gilmour methodology [4] was applied to induce conversion of (*E*)- to (*Z*)-cyano-derivatives, and then the behavior of this mixture was monitored by ¹H NMR (17.1 h). It was observed spontaneous interconversion of these isomers, corroborating to the hypothesis of thermodynamic equilibrium.

The product ratios of Knoevenagel reactions under study can be assigned to thermodynamic equilibrium between isomers. The existence of such equilibrium suppresses any preferences due to reaction mechanism, since after enough period, the ratio between isomers can be predicted considering only the relative energies of each isomer.

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Unravelling hydrogen bond network in theophylline-pyridoxine salt co-crystal by solid-state NMR

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Pharmaceutical co-crystals represent an emerging class of crystalline solids formed by at least one API (active pharmaceutical ingredient) and other one or more appropriate components (GRAS), laying in the same crystal lattice and held together by weak interactions (hydrogen bond, halogen bond, π -stacking).[1,2]

Solid-state nuclear magnetic resonance (SSNMR) spectroscopy is a versatile technique that can provide essential information about structure, packing and intermolecular interactions.[3] Hence, SSNMR is increasingly proving to be a valuable complementary method to single crystal X-ray diffraction (SCXRD), usually considered as the best tool for structural studies.[4,5]

Herein, we report an experimental and computational investigation by means of SSNMR and DFT (density functional theory) calculations of a theophylline-pyridoxine co-drug, a co-crystal formed by two APIs usually given in co-therapy. The co-drug has been initially characterized by SCXRD, IR and Raman spectroscopy and then an exhaustive SSNMR study has been performed. SSNMR spectra have provided several information, such as the number of independent molecule in the unit cell (Z'), the purity and crystallinity of the sample, the neutral or ionic nature of the adduct and hydrogen bonding properties. Several advanced 2D SSNMR spectra such as ^1H DQ MAS, ^{13}C - ^1H HETCOR, ^{14}N - ^1H J- and D-HMQC were acquired, taking advantage of the resolution and sensitivity improvement provided by indirect detection pulse sequences and very fast MAS at 70 kHz. These experiments, supported and completed by DFT calculations, have been fundamental to accurately determine the position of hydrogen atoms and thus to elucidate the complex HB network and to define the ionic character of the drug-drug salt co-crystal.

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Quantitative NMR (qNMR) with new Certified Reference Materials for ^1H , ^{31}P and ^{19}F

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Over the last decades quantitative NMR (qNMR) spectroscopy has become an important tool for the content determination of organic substances and the quantitative evaluation of impurities. Since the signal intensity is directly proportional to the number of protons contributing to the resonance, qNMR is considered as a relative primary method [1-3]. Quantitative NMR in combination with metrological weighing was optimized to show the power of the measurement method [4]. The implementation of qNMR in new fields of application (e.g. metabolomics, biomarker discovery, physiological pathways) brings along complex molecules and systems, thus making the usage of ^1H -qNMR more challenging. The answer to the problem is given by the use of other NMR active nuclei, namely ^{31}P and ^{19}F . We discuss three classes of qNMR CRM, based on different NMR active nuclei. The certification concept for ^{31}P -qNMR CRM shows how traceability to the SI can be established by using different nuclei [5]. We could adapt this approach to establish traceability for ^{19}F to primary Standard Reference Material (SRM) from the National Institute of Standards and Technology (NIST) and to primary Certified Reference Material (CRM) from the National Metrology Institute of Japan (NMIJ). The suitability of ^{19}F for qNMR is underlined by a detection sensitivity that is comparable to that of protons and a large chemical shift range (300ppm) [6].

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Towards the NMR characterization of Bombesin interaction with tumor cells expressing GRP Receptors

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Bombesin (BN) is a 14-residue peptide originally isolated from the amphibian *Bombina orientalis* [1]. It belongs to a family of peptides showing a variety of biological activities in numerous tissues and cell types, [2] exerted through their interaction with the Gastrin-Releasing Peptide Receptors (GRPR), transmembrane G-proteins coupled receptors triggering different signaling transduction pathways, resulting, among which, in the stimulation of cell proliferation. GRPRs are significantly involved in the pathogenesis of different human cancers [3], and are recently emerged as tumoral markers in early prostate and breast cancers diagnosis [4]. For these reasons, the research of new GRPR ligands as antagonists or carriers for cytotoxic and imaging molecular tools might be a promising strategy for the treatment and diagnosis of human tumoral malignancies [5].

In this scenario, structural data about BN binding to GRPR are required for the design and synthesis of high affinity receptor ligands, but, unfortunately, they are not yet available. BN conformation has been studied in various solvents demonstrating that it adopts an unordered structure in aqueous media and in dimethyl sulfoxide [6], while a partial helical structure has been observed in aqueous solutions containing TFE [7]. According to proposed models, this is the conformation that, probably, BN presents when anchored to biological membranes.

With the aim to verify the truthfulness of this hypothesis, we studied the effect of d25-SDS (a biological membrane mimetic) on BN by CD and NMR spectroscopy. As for BN-GPCR interaction, the heptapeptide BN(8–14) has been shown to be the minimal carboxyl fragment interacting with the receptor, the same experiment were performed also on the BN C-terminal heptapeptide.

Moreover, to discover the structural determinants of BN interaction with GRPR, the binding of both BN and BN(8-14) to human prostate carcinoma cell line (PC-3) over-expressing the receptor has been studied through on-cell STD-NMR experiments.

Acknowledgements

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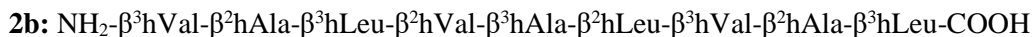
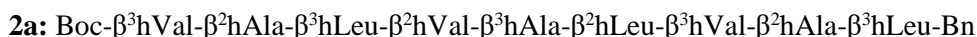
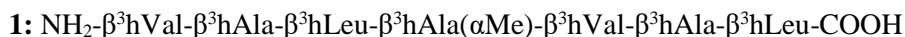
Studying the Conformational Ensemble of β -Peptides Using RDCs, ROEs and J-Couplings

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NMR spectroscopy is the method of choice for determination of the three-dimensional structure of molecules in solution. Most commonly, the available NMR data are thereby attributed to a single dominant molecular conformation. But molecules in solution are constantly subjected to conformational changes and representing the conformational ensemble as one single structure can lead to over-restraining and thus to misinterpretation of the data. Efforts to overcome this problem have mainly been focused on large biomolecules. For small and medium-sized molecules the low density of available restraints still renders a more adequate description of the conformational ensemble difficult. The incorporation of RDCs in the structure determination protocol and the use of more exact ROE- and NOE-derived distance restraints are promising measures to get closer to this goal.

We have studied the solution structure in methanol of representative members from two families of β -peptides exhibiting differential flexibility: The β^3 -heptapeptide **1** and the mixed β^2/β^3 -nonapeptides **2a** and **2b**. Peptide **1** forms a stable 3_{14} helix in methanol [1]. Earlier studies of **2a** suggest that a 12/10 helix is the dominant conformation of the terminally protected β^2/β^3 -peptide while deprotection to **2b** is believed to lead to an equilibrium between 10/12 and 3_{14} helix [2, 3]. It is known that β^2/β^3 -peptides can exhibit antimicrobial activity, and only recently they were found to penetrate the lipid bilayer of eukaryotic cells [4, 5].



We have determined RDCs of compounds **1** as well as **2a** and **2b** in stretched polyvinyl acetate gel in methanol. We have used this data together with ROE- and J-coupling derived dihedral angle restraints in a series of simulated annealing calculations in Xplor-NIH. The results are discussed with respect to our ability to more accurately define long-range structure and detect local and more extended flexibility in these compounds [6].

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A homochiral Polyglutamate with mesogenic sidechains as enantiodifferentiating alignment medium in NMR spectroscopy

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Within classical NMR spectroscopy in isotropic solution there are only few established possibilities to elucidate the spatial structure. The use of anisotropic NMR parameters – especially residual dipolar couplings (RDCs) – offers access to further structural information. However, partial alignment of an analyte with respect to the magnetic field is required [1]. This can be achieved by inter alia dissolving the analyte in lyotropic liquid crystalline (LLC) phases. These can be based on homopolyglutamates, i.e. poly(- γ -benzyl-L-glutamate) (PBLG) or poly(- γ -ethyl-L-glutamate) (PELG). Their helical chirality allows for enantiodifferentiation as different diastereomorphous interactions take place [2].

To obtain a better understanding of these interactions and the orientation process we decided to introduce a mesogenic biphenyl group into the sidechain. As mesogenes themselves can form LLC-Phases, we were wondering whether the rigidity and therefore the tendency to form LLC Phases can be increased. In previous works it was reported that the polymer with biphenylgroups shows surprising temperature- and solvent-dependent chiroptical changes [3]. This allows for the investigation of the impact of the sidechain on the analyte's orientation.

Therefore Poly(-*p*-biphenylmethyl-glutamate) (PBPMG) was synthesized and investigated towards its orienting properties. Herein we report about the performance of PBPMG as a versatile new enantiodifferentiating alignment medium with temperature tunable properties.

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Investigation into measurement uncertainty in Pure Shift NMR

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After many decades as a fringe analytical technique, qNMR has been gaining mainstream traction, primarily due to its acceptance as a primary reference technique and thus able to fulfil ISO Guide 34 criteria for generation of certified reference materials. LGC recently obtained ISO17025 calibration status for quantification of pure (>90%) organic materials by qNMR, allowing faster and more cost effective characterisation of reference materials.

The application of 1D qNMR in situations where the NMR spectra are less well resolved, due to structural complexity or multicomponent mixtures, can present significant challenges when high accuracy is required. To allow resolution of analyte signals, users are turning to alternate NMR experiments that afford improved resolution through additional dimensions and/or spectral editing. Unfortunately, the disadvantage of using such techniques for quantitative purposes is that they typically introduce analyte specific biases that are not present in classic 1D qNMR experiments. These techniques therefore do not offer the same degree of traceability to the international system of units required for ISO guide 34 reference materials. Understanding the limitations in the measurement techniques and the associated measurement uncertainty is vital in ensuring that the analysis is fit for purpose. In order to claim SI traceability of multiple pulse NMR techniques, these biases and additional sources of measurement uncertainty need to be quantified.

The use of quantitative HSQC based experiments, to alleviate signal overlap by dispersion of along the ^{13}C dimension, is well documented as its inherent biases have been thoroughly investigated. Pure shift NMR techniques have recently attracted a lot of attention and can improve signal resolution by removing homonuclear coupling. The majority of the literature relating to pure shift methods is focused on the qualitative features of this technique. This poster will address the implementation of pure shift methods for quantitative analysis and provide an assessment of its strengths and limitations in comparison to quantitative 1D and HSQC methods. It will discuss biases and uncertainties arising from a range of sources including method parameters and physical and spectral properties of the analyte and standard.

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WebCocon: From Correlations to Constitutions in the WWW

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The use of computers for structure elucidation has been shown numerous times to help researchers to improve the quality of the determined structures. Most of these Softwares are integrated into commercial packages, commonly combined with NMR data handling or data analysis. But there are a few Softwares available for download, which mostly lack in user interface capabilities. Usually they come alone, without integration with other tools that could help in the improvement of their output.

Almost 15 years ago we presented WebCOCON (Web - Correlations to Constitutions), which is based on COCON [1,2], a closed source software for structure elucidation. Until today, it's the only service that is offered for free in the WWW, and continuously undergoes changes. It was the first CASE software that used molecular modeling in order to improve the results. MD was selected due to its speed (<1s per structure), and all suggested molecules are subjected to it. If the MD fails, the structure is excluded from the result set (our statistical filter), otherwise the total energy calculated is used for the final ranking [3].

Today several CASE packages offer the integration with DFT calculations, which results in a much higher overhead (>2h per structure). WebCOCON has been integrated with DFT for the best 5 structures only, after the MD calculation. But, the quality of the NMR data calculated by freely available DFT packages is much lower when compared to commercial packages, so that for now we gave up on it.

From early on WebCOCON offered a unique feature: "Structure Discussion" based on theoretical NMR correlation data [4]. For this purpose a molecule is submitted to the server, which first generates theoretical NMR correlation data and then runs a structure elucidation with this data. Interestingly, for many molecules alternatives can be found that can not easily be distinguished. This kind of application can be regarded as procheck (and others), but for small molecules.

By now the complete WebCOCON interface is done in Javascript, resulting in a much improved user's experience.

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Tautomerism and Conformational Studies of Fluorinated 4-Aryl-1,5-Benzodiazepinones

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Although 1,5-benzodiazepines are compounds with interesting biological properties, they can be considered orphan molecules, in the sense that their pharmacological potential has not been fully explored [1].

Continuing with our studies on new scaffolds, we present here our results concerning three 4-aryl-6,7,8,9-tetrafluoro-NH-1,5-benzodiazepine-2-ones (Ar= phenyl, 2-fluorophenyl, 2-chlorophenyl) and their corresponding N-methyl derivatives [2,3].

The NH parent compounds have been prepared by reaction of 1,2-diamino-3,4,5,6-tetrafluorobenzene with ethyl benzoylacetate, ethyl (2-fluorobenzoyl)acetate and ethyl (2-chlorobenzoyl)acetate [4]. Iodomethane/K₂CO₃/KI in N,N-dimethylformamide was used to obtain the N-methyl compounds [5]. The structural characterization by multinuclear NMR (¹H, ¹³C, ¹⁵N, ¹⁹F) both in solution and in the solid state, permitted us to study the imine/enamine tautomerism concluding that in the NH compounds a mixture of both tautomers exists in solution. In solid state, only the imine (-N=C(Ar)-CH₂-CO-) tautomer is present.

To understand these results B3LYP/6-311++G(d,p) theoretical calculations were carried out. The imine tautomers correspond to the most stable structures and the calculated geometries are similar to the three experimental ones obtained by X-ray diffraction analysis, but the aryl rings are more twisted. Concerning conformational studies, the experimental inversion barriers have been determined by variable temperature ¹H NMR. The values for the NH compounds are rather low, about 48 kJ·mol⁻¹ and the measurement was carried out in THF-*d*₈ at low temperatures. For the N-methyl ones the barriers are quite high (about 70 kJ·mol⁻¹) and the experiments were performed in DMSO-*d*₆ at high temperatures. In both cases theoretical calculations provided consistent values [6].

The application of this methodology, which combines the use of NMR spectroscopy and DFT theoretical calculations, to novel 1,5-benzodiazepinones is now in progress.

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71

New NMR methods for Resolving overlapping spin systems and accurate determination of chemical shifts

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Nuclear magnetic resonance (NMR) spectroscopy is a powerful technique widely used not only for structure determination but also for dynamic studies of molecules. Its contribution in the field of metabolomics has been significant despite its lower sensitivity. An important step in the analysis of NMR spectroscopic data is accurate estimation of the chemical shift values of peaks observed in the spectrum and the identification of the individual components. However, it is a challenging task hindered by severe signal overlap, with weak intensity of some resonances further compounding the issue. This problem is frequently encountered in metabolomics and systems involving large proteins and their complexes, where the sample contains a mixture of a large number of compounds with varying S/N. The resolution in NMR is directly proportional to the duration for which the signal is acquired and is considered to be limited by the intrinsic linewidth of the peaks. For achieving good resolution, the NMR signal or the free induction decay (FID) is acquired typically with a large number of sampling points and processed using apodization, linear prediction or line-shape fitting. However, these methods become unsuccessful if the signal decays rapidly due to transverse relaxation resulting in large linewidths. While approaches such as involving dual receiver system have been established for rapid NMR data acquisition but resolving broad or overlapping peaks remains a bottleneck for application of NMR.

We address this problem on the basis of higher-order spectra estimation technique to accurately estimate the chemical shifts and achieve high resolution. Particularly for metabolomics field we have developed a new phase modulated HSQC-TOCSY NMR experiment which facilitate identification of metabolites for resonance assignments. By suitably adjusting the ^1H chemical shift evolution period to a specified value in a conventional 2D [^{13}C - ^1H] HSQC-TOCSY experiment, the phase of the cross peaks of different metabolites are modulated facilitating their recognition. The results of these studies will be presented.

72 High-resolution in 2D heteronuclear experiments: spectral reconstruction based on chemical shift encoding in F1 dimension

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Obtention of high resolution in carbon dimension of 2D heteronuclear experiments has always been a problem since they are indirectly detected. An alternative to incrementing the number of acquired points, and thus the experimental time, in order to achieve higher resolution is to implement spectral aliasing [1]. Aliased spectra are obtained by reducing the F1 spectral window, which increases resolution by a factor proportional to the reduction of the spectral window. The main disadvantage for acquiring an aliased spectra are the chemical shift ambiguities introduced, making impossible to determine the true chemical shift of an aliased signal. Different methods have been reported to overcome these ambiguities [2,3].

We present a general approach to resolve chemical shift ambiguities due to spectral aliasing by encoding signal's chemical shifts along the F1 dimension. The inclusion of an additional specific t_1' evolution time block without quadrature discrimination results in a signal split proportional to the ^{13}C chemical shift. Further processing with an external computer program is used to identify the different splitting partner and therefore to calculate the true chemical shift based in the distance separating them. A full-width reconstructed spectrum is obtained by placing the splitted peaks into their previously calculated true chemical shift. Reconstructed spectrum has the same resolution as the aliased spectrum, that is typically 10-100x higher than standard heteronuclear spectra.

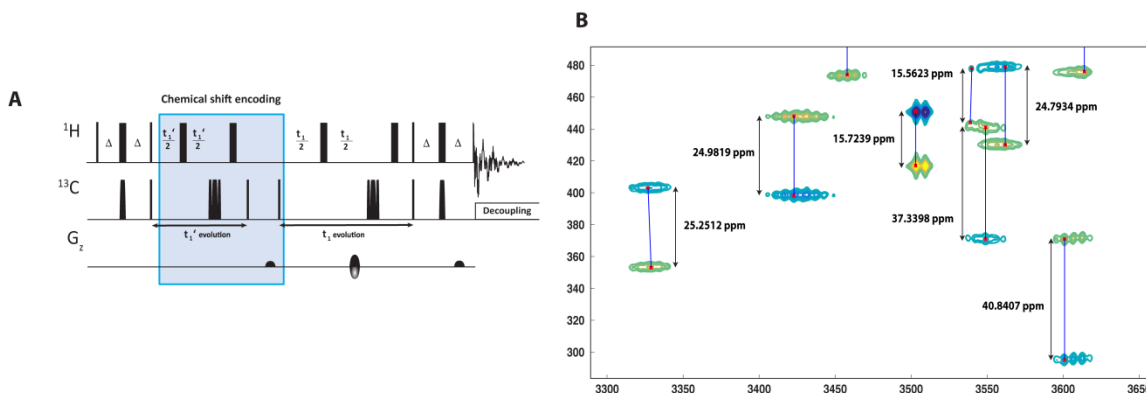


Figure 1. A) HSQC pulse sequence with additional signal evolution for F1 chemical shift encoding. B) Example of an aliased edited HSQC spectrum processed with 4k/512 points. After the individual peaks are identified, they are matched by pairs and their precise and accurate true chemical shift are determined.

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Conformational Deconvolution of NECA in Solution

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NECA is a 5' modified adenosine analogue and shows interesting biological properties as a synthetic non selective adenosine receptor agonist. The NECA conformation with the purine ring *syn* to the ribose together with an internal hydrogen bond, was indicated to be a major conformer in solution (DMSO) by NMR and hence was suggested as being the form responsible for receptor biological activity [1]. This conformer is also the form found in the single crystal x-ray of NECA, further supporting it as a low energy form [2].

However, recent work demonstrated that NECA bound to a Thermostabilised Human Adenosine A_{2A} Receptor [3] with a very different *anti* orientation of the purine ring.

Hence, further NMR data was collected on NECA in solution and the restraints obtained were deconvoluted to yield a conformational ensemble. The preliminary results of this work are outlined.

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New software kit for reaction monitoring using interleaved acquisition for high and low NMR

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The reaction monitoring is crucial and booming field of NMR. However most available solutions concentrate on utilizing series of 1D spectra to monitor the changes in the system.

Different approach is present in the poster by Alexandra Shchukina titled “Time-resolved NUS interleaved experiments with online processing and analysis”. In this work she presents the idea of acquiring of 2D randomly sampled spectrum interleaved with 1D. Allowing to have better insight in the changing system.

The work presented here shows the new software kit allowing user to utilize the idea presented by A. Shchukina on their own systems. The new program consists of two main parts:

1. Module/macros controlling the spectrometer acquisition allowing user to acquire interleaved spectra using simple GUI.
2. Processing module (combined with CS processing from MDDNMR) which allows online processing, displaying and analysis of spectra during, or post acquisition. The program is written in python allowing use of it on almost any OS and is providing user-friendly GUI.

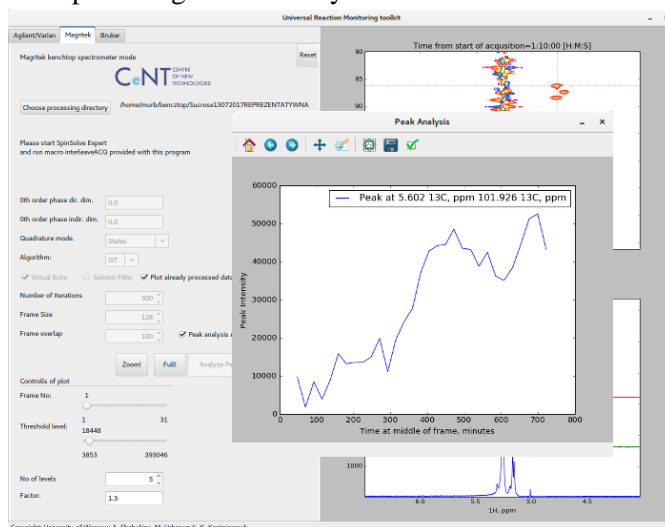


Figure 7: Screenshot of interface during development

Currently the software toolkit is working on Agilent and Magritek spectrometers (Bruker version will be available later) therefore working on both High-field and benchtop systems.

The first beta version of toolkit will be released on SMASH 2017.

75 A cross-platform format to associate NMR-extracted data (NMReDATA) to chemical structures

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10. University of Bristol, United Kingdom
11. Bruker BioSpin GmbH, Rheinstetten, Germany
12. ACD/Labs, Toronto, Canada
13. Mestrelab Research, Santiago de Compostela, Spain
14. Merck and co. New-Jersey, USA
15. Magnetic Resonance in Chemistry, Wiley, Chichester, UK

An open initiative involving all the major players of computer-assisted structure elucidation (CASE), including methodology specialists, software and database developers and the editorial board of Magnetic Resonance in Chemistry, is addressing the old problem of reporting and sharing the assignment of 1D and 2D NMR spectra of organic molecules.[1]

Our approach aims to solve some of the problems encountered with the “full analysis” of organic compounds. Usually, they are reported in chemistry journals using an image of the chemical structure, a text-based assignment of the 1D ¹H and ¹³C spectra and a set of tables listing the correlations found in 2D spectra such as COSY, HSQC and HMBC. In the best case, images of the spectra (of uneven quality and resolution) can be found as Supplementary material. This is unsatisfactory.[2,3]

We introduced a data format to associate the data extracted from the “full NMR analysis” (1D ¹H and ¹³C, COSY, HSQC, HMBC, etc.) and the structure of the identified compound. The file uses the SD format, that is compatible with .mol files (a quite commonly format used to draw chemical structures). The NMR-extracted data (chemical shift, coupling and assignment) are encoded as so-called “tags”, that are included in the .sdf files. These “tags” are not visible when displaying the molecules but can be accessed by specialized software such as CASE software and analyzed by the database during the importation of the data. These .sdf files including NMReDATA will be generated by future releases of computer-assisted structure elucidation software and have multiple roles:

- 1) They make the link between the atoms of the structure and the signals found in the spectra (assignment).
- 2) They list, for each 1D and 2D spectrum, the spectral parameters in a defined format (chemical shifts, couplings, integrals, 2D correlations)

3) They combine the data extracted from the spectra into an aggregated table (list of chemical shifts, coupling network, etc.).

4) They include links from the spectral data to the original files of the spectra (located in a local files folder in a database).

These files will be uploaded on a database together with the associated spectra as embargoed NMR data. A link to the data will be included in the manuscript submitted to scientific journals. The reviewing will be facilitated by the fact that the spectra, the extracted data and the structure will be accessible in an usable electronic format. The reviewers will use their favorite CASE software to assess the assignment. Once the paper is accepted, the spectra and the extracted spectral data become openly accessible to everybody.

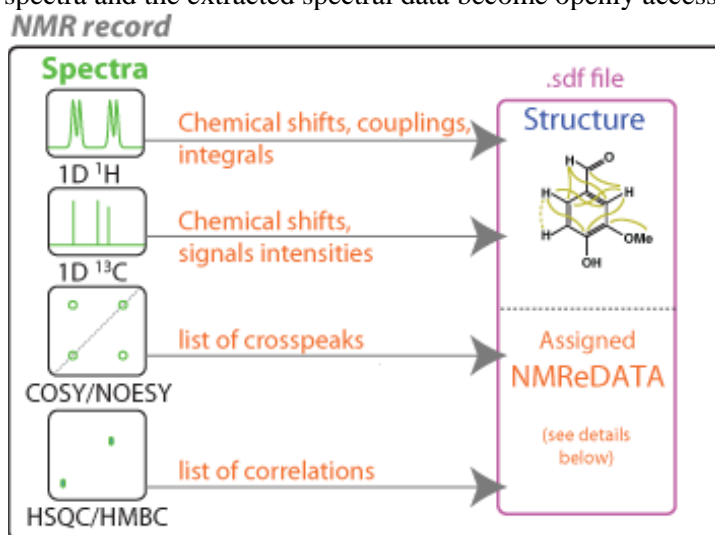


Fig. 1 Schematic representation of an NMR record including the files of the NMR spectra, and the SDF file containing the structure and the assigned NMR data extracted from the spectra.

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Human Serum Albumin and Drugs a Love Story Told by NMR

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Plasma protein binding affects the time that a drug stays in the body and can also influence the drug's efficiency. Only the unbound fraction can have a biological effect or be metabolised/excreted. Because of that, estimating the unbound fraction of a drug candidate is of capital importance in the drug discovery process.

Human serum albumin (HSA) is the most abundant protein in the blood circulation, it is 600 μM concentrated in plasma and most drugs have some affinity for HSA that results in sequestration of compound in serum. Furthermore, this 65 kDa protein presents various and diverse drug-binding sites: specific and no specific. In fact, HSA is able of soaking up ligands in a non-specific manner. This conduces to apparently discordant results of diverse individual ligand binding studies, which are difficult to generalize, and the great quantity of data are perplexing.

Thus, considering the relevance of evaluating the HSA binding of pharmacological compounds, we present here a systematic NMR study for valutatating the binding site and the affinity of several representative drugs to HSA. We have used 1D ^1H line-broadening [1] and competition-STD experiments [2] for quantitative affinity measurement, and 2D ^1H - ^{13}C correlation experiments for determining the binding site in HSA [3], with the aim to have a clearer picture of the interaction of diverse drugs with this abundant protein in plasma.

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Simplifying complexity: a new approach for the analysis of fluorine-containing mixtures by NMR

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In recent years, interest in fluorine-containing compounds has significantly increased, mainly due to their biological activity as anticancer, antidepressant and antiviral drugs, amongst others.[1] NMR spectroscopy has proven to be a powerful tool for the analysis of fluorine-containing mixtures.[2,3] However, most current methods fail to extract structural information for individual components in a mixture, and new methods to tackle such problems are needed.

Here we propose a novel two-step 1D experiment for obtaining structural information from simplified ¹H spectra showing only those protons that are in a spin system coupled to a selected fluorine. The first step is a doubly selective ¹⁹F-¹H reverse INEPT block, transferring coherence from a selected fluorine to a selected proton. The second is a TOCSY transfer to all the protons that correlate with the selected HF pair. We show the power of this method for a mixture containing four fluorinated pharmaceuticals. From this complex mixture, a set of clean 1D ¹H TOCSY subspectra is obtained, one for each fluorine resonance.

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78 ¹H qNMR for Multiple Compounds Determination in Commercial Energy Drinks

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The global market of energy drinks was valued at USD 43 billion in 2016 and it is expected to witness a high growth in the next years [1] and due to this, the quality of the products is essential. Considering the analytical techniques that are employed for this purpose, the advantages of NMR spectroscopy can provide a great differential in the energy drink analysis.

In this context, the aims of this work were the development and validation of a ¹H qNMR method to determinate multiple analytes in commercial energy drinks and to explore the NMR technique potentiality for analyses of non-targeted compounds, in order to identify possible some undeclared compounds.

The development of qNMR method was carried out according to EUROLAB Technical Report 1/2014 “Guide to NMR Method Development and Validation – Part 1: Identification and Quantification” [2]. The procedure of validation and its performance criteria were based on the Q2 R1 Guideline and FDA recommendation, respectively [3,4].

The procedures of analytical methodology (accuracy, repeatability, intermediary precision, specificity, range, linearity, detection and quantification limits) were satisfactory evaluated since the values obtained were with the FDA performance criteria [4]. Six compounds (caffeine, sucrose, glucose, citrate, threonine and ethanol) were identified and simultaneously quantified in several commercial products. The quantification was carried out directly in the matrix diluted in D₂O (1:1 v/v) employing maleic acid and quinine (Certified Reference Material – RMs) as internal and external standard, respectively. The ethanol identified and quantified was not declared in the label of any products.

Therefore, the identification of undeclared compound show as the NMR technique can be useful to investigate possible adulteration. Besides, the ¹H qNMR method developed in this work can contribute for the quality control improvements of commercial energy drinks in a simple and fast way.

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Computer Assisted Structure Elucidation of Two Biflavonoids from the Leaves of *Ochna Mauritian*

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As part of a phytochemical investigation of the leaves of *O.mauritiana* we have isolated, using a combination of silica gel flash chromatography and preparative layer chromatography (PLC), two biflavonoid compounds (1 and 2) from the organic extract of this plant. Approximately 12.5 mg of 1 and 2.5 mg of 2 were placed in NMR tubes and dissolved with DMSO-d₆ in order to elucidate their structures. High-resolution mass spectra were also recorded in order to establish the molecular formulae.

Both compounds were relatively proton deficient resulting in few correlations in long range 2D NMR experiments. The repeated ring structure made some of the ¹³C peaks appear very close together, which also limited the usefulness of conventional 2D experiments. Together with the presence of many exchangeable protons, the elucidation of the structures proved to be rather challenging. Here we describe the advanced computation method used for elucidating the structures using a series of 1D and 2D NMR experiments.

Initially, a search was performed on a database with predicted ¹³C spectra, or ca. 92 million compounds in search of a spectrum similar to the ¹³C spectra of 1 and 2. The spectrum of 1 matched that of a previously reported structure, while there was no match for the spectrum of 2.

Computer Assisted Structure Elucidation (CASE), together with band-selective versions of the HMBC experiment were employed to assign the NMR data and unequivocally determine the structures and thus confirming the molecules. Additionally, several closely related structures were ruled out either by poor agreement between predicted and experimental ¹³C chemical shifts, or by additional derivatization experiments.

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Comparison of Public Chemical Structure Databases for Structure Dereplication and Elucidation

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The capability to quickly and reliably separate and identify active components in natural product mixtures—identified through bio-assay and/or mass spectrometry guided fractionation—is critical to successful natural product-based drug discovery. Dereplication refers to the process of screening active compounds early in the development process in order to recognize and eliminate those compounds that have been studied in the past. This process enables scientists to proactively decrease the number of structures that will need to be fully elucidated and focuses testing on true ‘unknowns’.

One question that should be addressed before starting a dereplication project is ‘where can a comprehensive list of all the known structures be found’? It is generally recognized that the largest collections of such structures exist in PubChem and ChemSpider. PubChem [1] is a database of chemical molecules and their activities against biological assays. It is maintained by the National Centre for Biotechnology Information, is part of the National Library of Medicine, and contains more than 93 million structures. ChemSpider [2] is a database of chemicals, that is owned by the Royal Society of Chemistry and currently contains approximately 59 million structures.

The ^{13}C NMR spectrum of a compound can be considered a fingerprint since it is virtually unaffected by conditions such as pH, concentration, and solvent effects that can cause ^1H NMR spectra to vary. It is also largely magnetic field independent, since there are no couplings that could cause variations in stronger or weaker fields. Searching databases of predicted ^{13}C NMR spectra for similarity to the unknown is, therefore, a very powerful and reliable strategy [3] for structure dereplication. The search can be enhanced by including search terms such as molecular formula (expanded to cover MF ranges) and by accommodating for missing or extraneous peaks in the NMR spectrum. It is also very common to use such databases to identify structural fragments in the case of genuinely unknown structures.

In this work we will compare PubChem and ChemSpider in terms of overlap between the two databases, the types and variety of compounds contained therein, and their overall effectiveness in structure dereplication and elucidation

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Small probe molecules, Trimethyl phosphine and ^{15}N -Acetonitrile, as probes of acidity in FCC catalysts

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FCC catalysts responsible for the conversion of heavy fractions of crude oil into useful products like fuels (gasoline, naphtha, LPG). They are composed of zeolite Y and a matrix of silica-alumina, kaolin and clay. Of them, the zeolite Y plays an important role in the catalytic activity through the Brønsted and Lewis acid sites.

One of the most important aspects in the behavior of these catalysts is their continuous loss of activity due to the clogging of the pores by the formation of coke and the poisoning of the active sites by the deposition of metals such as Ni and V, causing the inactivation of the same. To counteract these effects, the catalyst is subjected to regeneration cycles to improve it and again is feedback it to the FCC unit.

Small molecules can be used as probes to measure the acidity in these samples. ^{15}N -Acetonitrile in function with ssNMR spectroscopy is a powerful technique for the structural characterization. The isotropic chemical shifts of ^1H and ^{15}N of acetonitrile depends of the interaction with acid sites (Brønsted and Lewis). An upfield shift of ^{15}N resonance will be related with an increase in acid strength of the Brønsted sites.

The acidity of FCC catalysts hydrothermally deactivated and different regeneration cycles was analyzed. Acetonitrile (15N/99%) and trimethyl phosphine were used as probe molecules. CPTOSS experiments allow us to identify the different Brønsted and Lewis acid sites in the samples, ^{31}P -one pulse experiments give us the quantity of acids sited presents in the materials and, HETCOR $^{15}\text{N}\{^1\text{H}\}$ and $^{31}\text{P}\{^1\text{H}\}$, allows the unambiguous assignment of the Brønsted sites. Also, was revealed decreasing structural properties and change of the number of Brønsted acid sites of the FCC catalysts after hydrothermal deactivation.

Keywords: FCC catalyst, solid state NMR, ^{15}N -acetonitrile, deactivation.

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A Novel qNMR Technique: Quantitative Global Spectrum Deconvolution (qGSD)

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Global Spectrum Deconvolution (**GSD**) is a technique for an automatic peak analysis of NMR spectra (1). GSD analyses frequency-domain NMR spectra and produces a highly accurate list of NMR peaks with refined peak parameters including position, amplitude, linewidth and shape. GSD has been at the core of Mnova data analysis for many years, and has proved to be highly reliable and fast. It is widely used in Mnova as a preparation step for many advanced analysis routines like multiplet analysis, automatic assignments, automatic structure verification, etc.

Deconvolution of NMR spectra is an attractive technique for quantitative analysis: accuracy is often adequate, and it can provide realistic integrals even for overlapping lines. However, the accuracy of the quantitation depends on how well theoretical models describe experimental NMR resonances. As a default, GSD uses a Generalized Lorentzian function to model the experimental lineshapes. This is a very flexible model that covers a broad range of shape variations for NMR resonances. However, careful analysis (2) shows that even such a flexible model cannot precisely fit all possible experimental lineshapes, as evidenced by non-zero fitting residuals. As a consequence, due to its model-free nature, for well resolved resonances, the classical point-by-point integration method may produce higher accuracy quantitation compared to model-dependant procedures.

To have the “best of both worlds”, we would like to determine integrals that have the accuracy of traditional integration with the resolving power that comes from GSD, together with access to an analysis capability of partially overlapping signals.

To improve GSD quantitation, we propose a new technique of “quantitative GSD”: **qGSD**. It is based on careful analysis of the residuals after GSD processing, and correcting GSD lineshapes in a way which minimizes the residuals. Preliminary results of qGSD applied to real 1D NMR data will be presented. We will show that for well-resolved spectral lines qGSD can offer quantitation precision which is significantly improved over conventional GSD, and approaches or meets the precision of the classical sum integration. At the same time, the technique also provides a good quality quantitative analysis of the lines within overlapped regions.

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83 F1-decoupled CLIP-COSY experiment

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The presented research work focuses on the novel F1-decoupled CLIP-COSY [1] experiment. The experiment is very attractive for a structural chemist seeking for a fast assignment of signals in highly resolved spectra and in addition it is also an interesting case study for a NMR methodologist.

Based on in-phase magnetization transfer during the mixing, it is compatible with different kind of decoupling blocks to be applied in the middle of the t_1 evolution. While the use of the selective modulated pulse (*nemoZS*) [2,3] or the PSYCHE [4] element ensures optimal sensitivity, the application of the BIRD [5] filter introduces some interesting features in this experiment.

The BIRD, selectively inverting only the ¹³C satellites and thus decoupling them from all other protons, inserts an additional ¹³C isotope into the spin system under investigation. This alters the J coupling interactions and very often allows to neatly decouple signals which would otherwise be classified as strongly coupled. The presence of a distinct isotope allows to draw some parallel with heteronuclear experiments, in particular it gives some flexibility on the type of J coupling interaction to be observed and/or eliminated either by the homonuclear decoupling or broadband decoupling during acquisition.

It will be shown on the example of the BIRD-CLIP-COSY how this can be used to fully reveal multiplet structure of signals not easily accessible by other methods.

We also propose a solution to diminish the one drawback of the CLIP-COSY experiment related to the dependence of the signals intensity on the length of the in-phase transfer period. We suggest to increment the constant delay into a third dimension and Fourier transform the data. The sum of signals along the new dimension results in the CLIP-COSY where the intensity variation is drastically reduced

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1H Chemical Shift Variations in complex Mixtures at different Concentrations

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The chemical shifts of a compound are known to vary due to certain experimental conditions, specially the temperature and the solvent used. Only in some cases variations have been observed with changing concentrations, usually due to aromatic interactions of the observed compound. Also, changes of chemical shifts have been observed due to binding to proteins. But, the general expectation is that for normal experimental conditions, chemical shifts vary only slightly with the concentration of the analyte. Hence, spectra of mixtures can easily be compared among each other and compounds in mixtures can be identified.

This approach works, for example, for lyophilized blood plasma, but it did not work with coffee preparations. Our coffee preparations were produced using a normal household coffee machine, a fixed volume of coffee was lyophilized, and later a known amount was solubilized in D₂O. Our first observation was that the chemical shifts of the caffeine signals of the mixture did not match the values observed in the spectrum of pure caffeine in D₂O. Next we acquired NMR spectra with varying concentrations.

In none of the spectra the chemical shifts of the caffeine signals matched. We observed that the signals that were shifting significantly, >0.15 ppm, could be found in the aliphatic region, whereas in the aromatic region the shift was only about 0.05 ppm. No other signals shifted in the aliphatic region, different from the aromatic region, where more changes were observed. In no concentration of our coffee preparations we could observe the chemical shifts of caffeine observed in a reference spectrum in D₂O.

This has two consequences: first, the assignment of the peaks can not be done by simple comparison of the pure compound spectra. Second, statistical analysis of this spectra is not possible, in this case even excluding the aromatic region completely does not resolve this, since changes in the aliphatic region were observed as well. This takes away one of the most powerful tools that spectroscopists have at hands for analysis and comparison.

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Stop Flow Approaches to in situ Analysis of Reactions

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The work presented will centre around our development of in situ NMR analysis techniques for reactions of interest to the synthetic chemist; these will be exemplified in the context of our recent investigations of reactions employing reagents based on boron and silicon.¹⁻³ The overall project that these developments have contributed to, aims to gain a better grasp of the fundamental physical and chemical processes that facilitate and govern these reactions, thereby allowing "informed control" of the processes, either directly, or indirectly via modulation of undesired side reactions. Mechanistic interrogation via analysis of reaction kinetics and other physical organic parameters is the key to this insight, and gaining sufficient quantity and quality of data is pivotal. The bulk of our data is obtained by UV, IR, MS, and most especially NMR, in concert with strategic isotopic labelling and computational analysis.

We will focus on the problems we faced when reactions became too fast, or too sensitive to be reproducible. This resulted in the conventional techniques for in situ analysis of reactions by IR and NMR either being of limited use in terms of reaction life-time, precision, or highly inefficient in terms of the physical handling aspects of accurate reaction assembly.

We thus began to explore new approaches and devices to give us traction in the analysis of these systems. These developments have allowed the design, assembly and interfacing of new devices with IR and NMR instruments. The presentation will reveal details of these new devices. We will show how they allow the exploration of synthetic organic reaction kinetics in situ, not just quickly and efficiently, but also in a way that the devices can be rapidly coupled / decoupled to standard IR and NMR instruments, thus requiring instrument access only when the device is being used.

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Real-time NMR in Human Cells

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NMR has for a long time been used to study metabolism in cells. However human cells have always been challenging as the conditions needed to keep cells viable are more difficult to achieve in an NMR system than for *Escherichia coli* bacterial or yeast cells. We have recently described an experiment where we have embedded primary chronic lymphocytic leukaemia cells in an agarose matrix and measured metabolic turnover for an elongated period of time [1]. In this system, we observed fast and reversible adaptation of these cells to lowering oxygen levels.

In order to be able to better control the conditions of such real-time experiments we have now explored a flow-based approach where cells are placed in a flow-tube, and medium is circulated over a period of 12h. In collaboration with Bruker we have adapted the InsightMRTM flow-unit for this purpose. By adding a bioreactor in the flow-path we can control oxygen and CO₂ tension. This approach has been tested for different human cell lines.

This new approach offers unprecedented opportunities to test the response of patient derived cells to altering conditions or the exposure of drugs, and may open new avenues in the context of personalised medicine.

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87 Spontaneously hypertensive rats on high-fat diet as a model of metabolic syndrome: NMR metabolomic study

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Metabolic syndrome is a cluster of pathologies that includes insulin resistance, type 2 diabetes mellitus, obesity, dyslipidemia, hypertension which when present in combination markedly increase the risk of cardiovascular morbidity and mortality [1]. Spontaneously hypertensive rats (SHR) are one of the most common animal models used to study essential hypertension in humans. However, it was demonstrated that high-fat (HF) diet increased body weight, serum cholesterol and blood pressure in SHR and these changes in metabolic parameters were associated with cardiovascular dysfunction in these animals [2]. SHR on HF diet can be used as a model of metabolic syndrome [3]. The aim of our study was to characterize for the first time the SHR-HF model using NMR-based metabolomics in combination with the standard biochemical parameters.

SHR-HF model was studied in the context of both obesity and hypertension. The normotensive Wistar-Kyoto (WKY) rats served as the closest genetic control for the SHR. Male SHR and WKY rats were supplied with a standard low-fat (LF) and a HF diet, establishing four experimental groups: hypertensive and normotensive, both lean and obese. Urine samples were collected overnight after 12 weeks of HF diet. NMR spectra were acquired using Carr-Purcell-Meiboom-Gill pulse sequence with the presaturation. Pre-processed spectra were subjected to multivariate statistical analysis (Principal component analysis (PCA), Partial least square – discriminant analysis (PLS-DA)) and univariate parametric and nonparametric tests.

PCA and PLS-DA models display substantial separation of all experimental groups: diets are clearly differentiated by the first principal component, genetic strains by the second one. Urinary metabolic profiles of obese animals show attenuated concentration of hippurate, trigonelline, fumarate, phenylacetylglycine, 3-indoxylsulfate, citrate, and methylamine, and increased levels of tartrate, methylnicotinamide, glucose, taurine, dimethylamine, and allantoin compared to the corresponding lean controls. Changes of these metabolites are manifested to a different extent depending on the genetic strain. To identify a potential relationship between the urine metabolic composition and pathologies clustered in metabolic syndrome, the metabolite concentrations were also correlated with significantly changed biometric and biochemical parameters.

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Proton NMR Spectroscopy in Investigative Product Analysis

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The speedy and reliable identification and quantification of unknown contaminants in both finished products and raw materials is crucial in understanding production issues and quickly solving potential quality issues. It can be critical to quickly acquire a top-level screen of a contaminated substance, where comparison to a control sample can identify the class of compounds that could be present. In addition, there is also a need to understand the composition of products however, successful de-formulation of particular complex matrices can require application of numerous techniques, each providing information about a different class of compound, which can take time.

NMR spectroscopy is a flexible technique with varied applications, which can provide useful information about a large number of chemical compounds that may be present in situations such as these. Simple sample preparation procedures and short analysis times also mean results can be obtained quickly. Examples of the application of NMR spectroscopy in the analysis of food, cosmetic and pharmaceutical products will be displayed, including contaminants screening, product de-formulation and investigation into product quality issues.

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Measuring Spin Relaxation Rates of Low-Gamma Nuclei using Satellite Exchange NMR Spectroscopy

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NMR spectroscopy is well established as the leading technique for the structure elucidation of small molecules and for studying their dynamic behaviour in solution. In particular, spin relaxation rates can be correlated with the global and local dynamics of a molecule. Measurement of the longitudinal relaxation time constants (T_1 s) for transition metals has the potential to provide valuable information on their complexes relating to their co-ordination geometry, motional properties and molecular interactions.

However, there exists scant evidence of longitudinal relaxation data of low magnetogyric ratio (γ) transition metal nuclei in the literature, much of which was collected many decades ago. There has been limited progress in this area due to a number of factors, including the poor detection sensitivity of low- γ nuclei and often the need for specialised NMR probes operating at the low resonance frequencies appropriate to observe metals such as ^{103}Rh , $^{107/109}\text{Ag}$, ^{183}W and ^{187}Os . These factors make the direct measurement of the relaxation properties of such nuclei largely impractical using standard instrumentation.

In this work we demonstrate the potential to measure relaxation rates of low- γ nuclei without the need to observe them directly or even to apply rf pulses to them, making these measurements accessible with conventional broadband probes. This can be achieved through the observation of “spin-state satellite exchange” in 2D EXSY (Exchange Spectroscopy) spectra of a J-coupled, high- γ partner nucleus. We demonstrate this approach by measurement of the longitudinal relaxation rates of ^{195}Pt (as a model system) and $^{107/109}\text{Ag}$ metal centres in organometallic complexes through observation of scalar-coupled ^{31}P nuclei. Excellent agreement is demonstrated for T_1 data collected for ^{195}Pt using direct observation with the conventional inversion-recovery sequence.

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NMR studies of hydrogen cleavage by Solid-phase frustrated Lewis pairs

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Hydrogenation is important in many chemical transformations and frustrated Lewis acid pairs (FLP) offer a metal-free catalyst for such reactions.¹ In the solution phase FLPs have shown extensive catalytic hydrogenation activity, but for industrial processes a solid catalyst is preferred for reasons such as; catalyst separation, the use of continuous gas or liquid feed fixed bed reactors or slurry-phase reactors.² Here we report the preparation and characterisation of a silica supported FLP $[\equiv\text{Si-OB}(\text{C}_6\text{F}_5)_2][\text{P}^t\text{Bu}_3]$ and its reactivity on exposure to hydrogen, deuterium and CD_3OD characterised by ^1H , ^2H , ^{11}B , ^{19}F and ^{31}P Solid State NMR spectroscopy.³

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Automatic Concurrent Phase & Baseline Correction in 1D NMR Spectra

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The algorithmic problem of phase correction (PC), and that of baseline correction (BC), of 1D NMR spectra have been both tackled many times over the last half a century, by many authors.

There are many algorithms which emulate the manual procedure. Basically, they all 'fit' the parameters which describe the phase (ph0, ph1) and the baseline (various parametrized models) so as to maximize some quality assessment of the corrected spectrum. Historically, the employed 'quality functions' included peak heights, negative peak lobes, DISPA patterns symmetry, selected baseline points, peak ablation, etc.

Here we propose a radically different type of 'quality function' Q to be optimized. It is based on the histograms of the spectrum (real and imaginary parts) which turn out to be very sensitive to both phase and baseline distortions.

This permits us to:

1. Carry out the phase and baseline corrections simultaneously. In traditional approaches these are always intended as separate evaluation steps and carried out separately (first phase and then baseline), even though all NMR spectroscopists know that the two corrections interfere with other. We think that our approach neatly overcomes this problem.
2. Carry out both corrections on both the real and imaginary parts of a spectrum. So far, the baseline correction was always done only on the real part, a fact that can have various adverse effects on other evaluation tasks.
3. Enhance the objectivity of the corrections, especially considering that in practice one often faces situations with multiple acceptable 'solutions'.

We describe the algorithm we have developed and we illustrate the results achieved.

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Structure Revision and Conformational Analysis of the Fluxional Microginin 674

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The cyanobacterium *Microcystis aeruginosa* is well-known for the production of numerous bioactive modified peptides such as the hepatotoxic microcystins or the proteases inhibitors cyanopeptolins and microginins.¹ The latter metabolites are a class of linear peptides that contain generally four to six amino acid residues including the 3-amino-2-hydroxydecanoic acid (ahda). To date, around 50 structural variants are known but only 13 microginins have been fully characterized by NMR.²⁻⁴ Microginin 674 (**1**) was obtained from a culture of *Microcystis aeruginosa* (SAG 14-85). This tetrapeptide exhibited two tyrosines, the ahda residue and an *N*-methyl-methionine. Interestingly at room temperature, most of the 1D NMR signals of **1** were splitting, suggesting the existence of diastereoisomers or rotamers. The presence of the latter was first confirmed by changing the NMR solvent and then by variable-temperature NMR experiments (VT-NMR). Moreover, structures of **1** and a commercially available microginin 674 differ in the amino acids assemblage with an inversion between the *N*-Me-Met and Tyr2.⁵ The lack of NMR data provided by the companies and the difficulties in NMR data analysis induced by the presence of rotamers, question the structure proposed for this natural product. We report herein the structure revision, but also the experimental and theoretical conformational analysis of microginin 674 allowing the establishment of the thermodynamic parameters involved in the exchange process between the two rotamers.

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NMR of new metal complexes of curcuminoids with Mg, Zn, Cu, Mn, Ga, Gd and Sm

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Curcumin is a secondary metabolite with historical roots in its usage for culinary or therapeutic purposes. It has gained an outstanding relevance for its implications as a systemic antiinflammatory agent and its potential as an anti-cancer agent.[1,2] Also, derivatives diacylcurcumin (**DAC**) and tetrahydrogenated diacylcurcumin (**DAC-H**) have attracted wide attention due to their biological and complexation properties.[3]

Following our chemical studies of both ligands we prepared new metal complexes of **DAC-H** with Mg, Zn and Cu, which were studied in liquid state by high-resolution NMR. Also, new metal complexes of **DAC** with Mn, Gd, Ga and Sm were prepared and studied by NMR in both liquid and solid state. **DAC-H** complexes showed clear spectral changes caused by the metal binding to the diketone site. The metal ions were chosen after their presence in other complexes with biological properties.

Our previous crystal homoleptic structures showed the stoichiometry ML_2 for the complexes including the new Manganese complex reported herein. The vinylic proton signal of the ligand was used as a probe for complexation in DMSO, while in CP-MAS NMR, the profile of carbonyl signals and signal broadening provided a useful criteria of complexation. The case of trivalent metal ions is discussed.

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In Situ Monitoring of the Ascorbic Acid Electrooxidation by Ultrafast NMR

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Ascorbic acid (AA) electrooxidation is a known reaction, however this compound has many applications as the antioxidant agent in food and drink industry.¹ In this work we chose this reaction to be applied to a Electrochemistry and Nuclear Magnetic Resonance (EC-NMR) coupled system. The magnetoelectrolysis effect was also evaluated because the electrochemical reaction was performed *in situ*.² A promising methodology for the real-time monitoring of these reactions is the Ultrafast NMR (UF-NMR)^{3,4}, capable of recording 2D NMR correlations in a fraction of a second. The objective of the present study is to apply UF-NMR experiments to the real-time monitoring of *in-situ* AA electrooxidation reaction. The electrochemical system consisted in a glass capillary with two coaxial Pt wire coils as work and auxiliary electrodes, the pseudo-reference electrode was also a glass capillary, but with Ag wire. Cyclic voltammetry and chronoamperometry techniques were used to evaluate the magnetoelectrolysis effect and oxidate the AA in real time, respectively. During all the electrochemical process, in order to monitor in real-time the evolution of the oxidation of the AA, many ultrafast COSY (UF-COSY) were continuously performed every 41s, each one corresponding to an eight-scan experiment. Analyzing the most representative UF-COSY spectra, it was possible to note the gradual consumption of the AA during the course of the electrooxidation. After 15 minutes, the ¹H NMR correlations of the product, dehydroascorbic acid, were observed in the UF-COSY spectrum. It was confirmed by a HSQC experiment that the dehydroascorbic acid detected was present in the hydrated bicyclic and monocyclic forms. Also, it was detected another stable product formed on this condition and it still remains uncharacterized. We are performing other kinds of UF-NMR experiments to continue to investigate this electrochemical process, and also conventional NMR experiments to perform the characterization of the formed unknown compound.

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In the Colombian mountains, local farmers produce one of the best coffees in the world. The varieties of Arabica coffee and Robusta coffee are grown in special areas of the country and their quality, flavor and fragrance are recognized all over the world. Different methodologies by NMR and IR have been developed for the chemical analysis^{2,3} of aqueous extracts of Colombian coffee to confirm its origin¹ and distinguish it from adulterated coffee. Also, low field NMR has been used to establish the origin of Robusta and Arabica varieties.⁴

In this work, Colombian coffees were analyzed by ssNMR and their chemical compositions were correlated with the quality properties.

Samples used for solid state NMR spectroscopy do not require extraction steps previously to the analysis. All the samples were measured using a ¹³C/CP-TOSS pulse sequence to suppress the rotational echoes, contact time was optimized to obtain the best conditions to the ¹H polarization transfer at 10kHz rotation speed. Because in solids the line broadening by ¹H dipolar coupling results in an excessive line broadening, the chemical analysis can't identify individual chemical compounds, however it is possible to analyze the chemical changes of the samples on basis of the main chemical groups.

ssNMR results in combination with statistical analysis shows that it is possible to classify the samples on basis of quality criteria.

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The Role of Non-Uniform Sampling (NUS) in the NMR Characterization of Bile Acid Biomarkers

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2-D NMR is the quintessential analytical technique in the structural characterization of organic compounds. However, these experiments often have long acquisition times and can be limited in sensitivity and/or resolution. Reducing the required number of data points used in the second and third dimension through various non-uniform sampling (NUS) schemes has been used with protein and peptide samples to enhance resolution and decrease the acquisition time of multidimensional experiments. In 2-D and 3-D experiments, these models can result in a 75% reduction in acquisition time. Many of these gains can also be realized in the analysis of small molecules. The reduction in acquisition time by using NUS can be utilized to promote higher sample throughput (through a reduction in total acquisition time), increased sensitivity (by using more pulses per f1 data point) or increasing resolution by adding more data points in the f1 dimension. All of these approaches have been evaluated using common homonuclear and heteronuclear experiments used in the structural elucidation of small molecules (¹H-¹H COSY, ¹H-¹³C HSQC and ¹H-¹³C HMBC).

Recently, a biomarker project highlighted the utility of NUS in 2D small molecule NMR. The search for novel bile acid (BA) biomarkers of liver organic anion-transporting polypeptides (OATPs) led to a LC-MS method that supported the analysis of 30 different BAs, which included 15 different 3-O-sulfates. Many of these O-sulfates were not commercially available and required either chemical or biological synthesis. The resulting synthetic products need to be structurally characterized and in some cases quantitated via NMR prior to their use in a quantitative LC-MS assay. Because of the structural redundancies within bile acids high resolution 2D NMR was required. Using NUS acquisition schemes on these samples facilitated structural elucidation of the site of sulfation while saving a factor of seven or more in acquisition time.

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Natural bioactive-protein interactions studied by ligand-observed NMR

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Molecular recognition of small molecules by proteins is fundamental to most processes in living organisms. Small molecule-protein interactions are involved in transport, regulation, and metabolism in the cell, having direct implications in the molecular aetiology of diseases, as well as in therapeutic target discovery.

Natural compounds have long inspired drug development given their potential bioactivities. Plants are therefore an important source of bioactives with a wide chemical diversity. Cytokinins, N^6 -substituted purines, are a class of phytohormones promoting plant growth and development. Besides their role in plant physiology, little is known about the biological effects of cytokinins in humans.

To study the potential interactions of bioactives with human proteins, ligand-observed NMR was set-up. The interaction of a series of cytokinins N^6 -benzyladenine (BA), N^6 -benzyladenosine (BAR) and N^6 -benzyladenosine phosphate (BARMP) with the model protein, human serum albumin (HSA) was monitored. Different approaches were tested for this purpose: Water Ligand Observed via Gradient Spectroscopy (WaterLOGSY), relaxation-edited (T1rho) NMR and saturation transfer difference (STD) NMR. In all approaches, the 3 studied bioactives coordinate to HSA, with slightly different binding propensity, according to the different substituents. In addition, to minimize the amount of required protein, miniaturisation of the sample from 5 to 1.7 mm tubes was attempted.

With this study on cytokinins-HSA interactions, we hope to demonstrate the potential of ligand-observed NMR screening in the field of natural bioactives and bioactive mixtures.

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Assessment and Validation of Various Flow Cell Designs for Quantitative Online NMR Spectroscopy

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Compact nuclear magnetic resonance (NMR) instruments make NMR spectroscopy and relaxometry accessible in industrial and harsh environments for reaction and process control. Robust field integration of NMR systems have to face explosion protection or integration into process control systems with short set-up times. This paves the way for industrial automation in real process environments. [1, 2]

The design of failsafe, temperature and pressure resistant flow through cells along with their NMR-specific requirements is an essential cornerstone to enter industrial production plants and fulfill explosion safety requirements. NMR-specific requirements aim at full quantitative pre-magnetization and acquisition with maximum sensitivity while reducing sample transfer times and dwell-times. All parameters are individually dependent on the applied NMR instrument. Luckily, an increasing number of applications are reported together with an increasing variety of commercial equipment. However, these contributions have to be reviewed thoroughly.

The performance of sample flow cells commonly used in online analytics and especially for low-field NMR spectroscopy was experimentally and theoretically investigated by ¹H-NMR experiments and numerical simulations [3]. Here, we demonstrate and discuss an automated test method to determine the critical parameters of flow through cells for quantitative online NMR spectroscopy. The setup is based on randomized setpoints of flow rates in order to reduce temperature related effects. Five flow cells and tubings were assessed and compared for high-field as well as low-field NMR spectrometers.

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Collaborative study for determination of okadaic acid in solution by qNMR spectroscopy

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Okadaic acid is one of the diarrhea shellfish toxins. An instrumental analysis has been introduced into the testing of diarrhea shellfish toxins in many laboratories all over the world, where the regulatory level of the okadaic acid group is set as 0.16 mg/kg at CODEX in 2008. Certified reference materials (CRMs) of the diarrhea shellfish toxin are required for the instrumental analysis, and CRMs which concentration is determined by qNMR with external standard method [1] are provided from NRC Canada. qNMR uses a different structural compound as a reference material, and the method is practically used for rapid purity determination. The internal standard method is considered more accurate than the external standard method in qNMR, and there are some reports about comparison between these two methods [2, 3]. We already established qNMR with internal standard method for standard solution [4]. This approach is expected to provide an accurate determination for low concentration solution of precious sample such as diarrhea shellfish toxins.

In this research, we studied the optimization of sample preparation and measurement conditions of qNMR spectroscopy with internal standard (IS) for the determination of low concentration of okadaic acid in solution. Moreover, a collaborative study within three laboratories was performed and the accuracy and variation factors of qNMR spectroscopy were discussed.

Okadaic acid was produced and purified by the optimized method in previous work [5]. Purified okadaic acid was dissolved in methanol and prepared to approximately 350 mg/L methanol solution. 1,4-bis(trimethylsilyl) benzene-*d*₄ (1,4-BTMSB-*d*₄) as an IS from Wako Pure Chemical Industries, Ltd. was the CRMs whose purity was (0.999±0.005) kg/kg (*k*=2). The measurement sample with an internal standard was prepared based on our previous work [4]. All of mass were weighed by micro balance (XP56V, METTLER TOLEDO, Giessen, Germany). This sample solution was measured by NMR spectroscopy. Measurement temperature was regulated at 23 °C. Typical qNMR experimental parameters were as follows: 59523.8 Hz (99.2 ppm) spectral width, 4.0 s acquisition time, 13.0 μs (90°) pulse width, 60 s relaxation delay, and 512 transients acquired. ¹³C decoupling was used in all measurement. Data processing was performed using MestReNova ver. 8.1.

The concentration of okadaic acid solution of approximately 350 mg/L was successfully determined with only 2.5 % of standard uncertainty by qNMR with using the internal standard method. This result was equal to the uncertainty reported from NRC Canada in spite of a low concentration sample of about 1/10 in the report. The measurement accuracy in this approach was confirmed from this result. In this inter-laboratory test, the determined values of okadaic acid were in good agreement among three laboratories by

selecting adequate signals with sufficient resolution. The obtained value was validated because the difference from collaborative study was equal to the measurement standard uncertainty.

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Combining Presaturation, ^{13}C -Filtering and PROJECT for Effective Solvent Suppression of Protonated Organic Solvents

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Presaturation is one of the most popular solvent suppression methods in the analysis of small organic molecules. This appears to be because it is highly frequency-selective, requires no specialist spectrometer hardware and requires minimal customization for any specific solvent. The frequency selectivity of presaturation leads directly to two significant disadvantages. First, solvent signal from regions of magnetic field inhomogeneity is not suppressed [1]. Second, ^{13}C satellites of solvent resonances are not suppressed.

This poster demonstrates that the PROJECT spin-echo sequence [2] can be used to overcome both of these disadvantages. First, phase cycling of the 180° pulses suppresses solvent signal from regions of magnetic field inhomogeneity in the same way as in NOESY-PRESAT, PURGE and FLIPSY. Second, adding only three ^{13}C pulses to the PROJECT sequence re-introduces evolution of J_{CH} allowing conversion of ^{13}C satellite signals into unobservable or dephased coherences. The resulting ^{13}C filtering suppresses a broad range of J_{CH} . These two features allow acquisition of high-quality proton spectra in protonated organic solvents such as DMSO and acetonitrile.

The main disadvantage of presaturation-PROJECT is that, like NOESY-PRESAT, it requires a 90° excitation pulse. Therefore, a long relaxation delay is required to obtain resonance integrals independent of T_1 relaxation rates. However, this also allows a longer and hence more selective presaturation pulse [3].

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